Food capillary suspensions

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Abstract

When a small amount of a secondary immiscible fluid is added to a particle suspension, the rheological properties of the suspension can alter dramatically. The addition of the secondary liquid can create a sample-spanning particle network due to capillary bridges formed between the particles, inducing a transition from a fluid-like to a gel-like state. This results in an increase in yield stress and low shear viscosity up to several orders of magnitude. Accordingly, also the stability of the suspensions alters dramatically. Particles that are stabilized in a sample-spanning network due to capillary bridges do not sediment, even after long term storage. Capillary suspensions can be created whether the secondary fluid preferentially wets the particles or not. Mixtures where the secondary fluid is the better wetting liquid (three phase contact angle is smaller than 90°) are referred to as pendular state suspensions. If the bulk fluid is preferentially wetting, the contact angle is greater than 90° and the suspension is in the so-called capillary state. Capillary suspensions can be used in a wide range of applications, including processing of porous ceramics, preparation of crack free films or the formulation of novel food products. This latter topic is the focus of this thesis.

The present thesis is divided into two parts. First, oil continuous capillary suspensions are studied. It will be discussed how capillary forces can be used to control the stability and flow behavior of food products. Simple model systems of particles in oil suspensions are investigated to determine the parameters that affect the network formation and the resulting rheological properties. For this, two food systems were chosen: corn starch and cocoa particles in vegetable oil. Water is used as the secondary liquid in both admixtures. Food suspensions were compared to simpler model systems made from CaCO₃ and polyvinylchloride (PVC) particles.

It is shown that capillary suspensions offer new opportunities for the formulation of food products. A clear transition from a liquid or very weak gel with no secondary phase added, to a strong gel-like state when water is added to the oil continuous suspensions made from cocoa or starch granules can be seen. This indicates that strong capillary forces act in this food model systems as has been reported earlier for other suspensions.

The wetting behavior of the particles with water and oil has been studied using confocal microscopy as well as contact angle measurements using the sessile drop method. Various rheological parameters of capillary suspensions are documented including oscillatory and steady rotational rheometry data.

Texture modifications can be done by adapting the amount of the secondary fluid and the wetting properties like the contact angle the two fluids form with the particles. On the other hand, the secondary fluid viscosity has no influence on formation and flow behavior of capillary suspensions. The adsorption of water by particles from natural sources during storage can dramatically influence the properties of oil continuous suspensions since capillary bridges form spontaneously from adsorbed
water layers. Suspension rheology is similar if the water is absorbed prior to suspension preparation by conditioning the particles or added to the suspension afterwards. These findings lead to the conclusion that the formation of these capillary suspensions is energetically favorable and not controlled by mechanical energy input.

Capillary bridges offer an additional advantage to chocolate or other temperature sensitive systems by increasing the heat stability of the products. This stability has been documented via optical experiments and was also measured using rheological setups. When water is incorporated to a model chocolate made from cocoa butter and cocoa particles, the blocks preserve their shape during one hour at 50°C. Pure model chocolate however melts after 30 minutes. This heat stability is due to the particle network created with the addition of water. The cocoa butter is trapped in between the formed network and the shape can be maintained. Increased volume fraction of secondary fluid as well as higher particle loading lead to a more stable product.

In the second part, I focus on the flow behavior of aqueous cocoa suspensions and how this can be adjusted by temperature or addition of a secondary liquid. A thorough rheological characterization is provided and potential mechanisms causing particle network formation are discussed. For this particles have been extracted with various solvents and the influence of added secondary fluid on the flow behavior is investigated.

When cocoa particles are dispersed in water and stored at a temperature of 30°C, a network of particles is formed which leads to an increase in low shear viscosity and a gel-like, creamy texture. Heating the sample further up to 40°C or above results in a network collapse and the suspension behaves fluid-like again. The critical factor controlling this behavior is the residual cocoa butter in the particles. At 30°C, the cocoa butter is in a semicrystalline state and bridges of this cocoa butter between the particles are formed via a diffusion process (similar to sintering) or just by sticking together upon contact due to cocoa butter that has leaked to the surface. The cocoa butter is completely liquid at 40°C and partly forms emulsion droplets in the water phase that are not connected to the particles as shown via confocal imaging. The key role of cocoa butter for network formation, texture and flow of the aqueous suspensions has been proven by extracting it from the particles. Particles, where the fat has been removed show no temperature dependent change in flow behavior.

Adding a secondary fluid to the cocoa suspension results also in an increased yield stress attributed to the formation of a network of particles even at room temperature. This bridging phenomenon only appears for particles with residual cocoa butter content. This leads to the assumption that the secondary fluid dissolves the cocoa butter and, similar to the heated particles a soft and sticky semi-crystalline fluid is created that glues the particles together. This process is faster for short chain solvents with a polar end-group compared to longer chain fatty acids and can be correlated to the speed at which the respective fluid dissolves the cocoa butter. Less polar substances like silicone oil or
sunflower oil do not induce a particle network formation, which is attributed to the poor and slow solubility of cocoa butter in these fluids.

SANS experiments also revealed structural differences between samples stored at different temperatures. Samples stored at 30°C or at 20°C, but with added oleic acid exhibit a scattering peak corresponding to a characteristic length scale of about 50 Å whereas the characteristic length reduces to about 35 Å for a pure cocoa suspension stored at 20°C. This length scale is attributed to the cocoa butter layer on the particle surface and completely vanishes upon storage at 40°C.

This research should encourage the development of low fat cocoa spreads that are not based on gums, hydrogels or other polysaccharides, but provide structural stability through a particle network formed within the suspension, resulting in a creamy texture.

Finally, it has been shown how the flow behavior of oil and water continuous food suspensions and correspondingly their creaminess and texture can be tuned in a wide range via appropriate preparation method and addition of an adequate, immiscible secondary fluid. This may open up a new pathway for the formulation of novel food products based on cocoa or other finely dispersed particles.
Zusammenfassung


Die Textur kann durch die zugegebene Menge an Zweitphase und die Benetzungseigenschaften wie Grenzflächenspannung und Kontaktwinkel verändert werden. Jedoch hat die Viskosität der
Zusammenfassung

zugegebenen Zweitphase keinen Einfluss auf die Netzwerkstruktur und somit auch nicht auf die Fleßeigenschaften wie Fließgrenze oder Viskosität. Untersuchungen zu unterschiedlichen Mischungen aus hydrophilen und hydrophoben Partikeln führten zu keinem allgemeingültigen Ergebnis.


mittels n-Hexan konnte die Schlüsselrolle der Kakaobutter bei der Netzwerkbildung bestätigt werden. Suspensionen, die mit extrahierten Partikeln hergestellt wurden, zeigten keinen Einfluss der Lagertemperatur auf das Fließverhalten.


Neutronenstreuexperimente zeigten strukturelle Unterschiede der Suspensionen bei unterschiedlichen Lagerbedingungen. Proben, welche bei 30°C gelagert wurden, und Proben mit zugegebener Zweitphase, die bei 20°C gelagert wurden, zeigten ein Streumaximum, welches eine charakteristische Länge von ca. 50 Å entspricht. Diese reduzierte sich zu 35Å bei Proben ohne Zweitphase, welche bei 20°C gelagert wurden. Dieses Streumaximum wurde auf die Kakaobutterchicht auf den Partikel zurückgeführt und verschwand ganz bei einer Lagertemperatur von 40°C.

Diese Forschungsergebnisse sollen die Entwicklung von fettreduzierten Lebensmitteln unterstützen, welche nicht mittels Hydrogelen oder anderen Polysachariden hergestellt werden, sondern allein durch das Partikelnetzwerk Stabilität und eine cremige Textur erhalten.

Schussendlich wurde gezeigt, wie das Fließverhalten von öl- und wasserbasierten Suspensionen im Bezug auf ihre Textur und Cremigkeit durch die richtige Herstellungsmethode und passende Zweitphase verändert werden kann. Dies eröffnet neue Möglichkeiten um innovative Lebensmittel auf Kakaobasis oder anderen dispergierten Partikeln zu formulieren.
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1 Introduction

1.1 State of the art

When a small amount of a secondary immiscible fluid is added to a particle suspension, the rheological properties of the suspension can alter dramatically. The addition of the secondary liquid can create a sample-spanning particle network due to capillary bridges formed between the particles, inducing a transition from a fluid-like to a gel-like state. This results in an increase in yield stress and low shear viscosity by several orders of magnitude. Accordingly, also the stability of the suspensions alter dramatically [1]–[5]. Particles that are stabilized in a sample-spanning network due to capillary bridges do not sediment, even after long term storage [1]. Capillary suspensions can be created whether the secondary fluid preferentially wets the particles or not. Mixtures where the secondary fluid is the better wetting liquid (three phase contact angle is smaller than 90°) are referred to as pendular state suspensions. If the bulk fluid is preferentially wetting, the contact angle is greater than 90° and the suspension is in the so-called capillary state. Capillary suspensions can be used in a wide range of applications. This could be processing of porous ceramics [6], coatings of crack free films or formulation of novel food products which will be described in this thesis.

The effect of water added to oil continuous suspensions for food applications has been previously discussed by Johansson and Bergenståhl for sugar (pendular state) and solid fat (capillary state) particles [7]–[9]. There, particle agglomeration with the addition of water was reported resulting in faster sedimentation of the fat or sugar particles, but no sample-spanning network was observed. This is similar to the so-called spherical agglomeration technique used to separate solids in coal and ore preparation [10], [11]. A focus on chocolate products can be found in literature, proposing that water can form capillary bridges between sugar particles and therefore a sugar network is created that prevents the fat phase from spreading while it melts [12], [13]. Several patents have been filed on that topic [14]–[17]. Work has been done to investigate the rheological behavior of sugar networks or the effect of surfactant concentration in model chocolate dispersions when water is added to the suspensions [18], [19]. Corresponding preparation methods [12], [20] for chocolate products have been discussed. Other particle/liquid/liquid combinations that form sample-spanning networks for food application as well as the influence of the wetting properties on the rheological behavior remain unexplored.

For the capillary state in capillary suspensions, calculations and experiments have shown that clusters of different shapes are formed within the suspension. Cluster configuration is highly dependent on the amount of secondary liquid and its wetting behavior [21]. With higher amounts of secondary liquid, particles tend to form octahedral clusters that have a stronger cohesion than tetrahedral clusters, which
are more favored for lower secondary liquid contents [21]. First approaches have been made to determine the strength of a capillary water bridge between two glass spheres in oil, showing that stronger forces are measured for secondary liquids with higher viscosity [22]. It also has been shown that the strength of a capillary suspension in the pendular as well as in the capillary state is proportional to the inverse of the particle size. Temperature can also alter the yielding and flow of capillary suspensions dramatically via its effect on interfacial tension and wetting properties. The addition of surfactant can prevent the formation of capillary bridges which results in a change in flow behavior of the capillary suspension [2], [3].

One current trend in food science is focusing on low fat products, to support healthier eating without the lack of taste and familiar mouthfeel. The texture of low-fat food products must be adjusted to mimic regular full fat products in order to gain consumer acceptance [23]. A similar mouthfeel and the stability of regular fat products can be achieved by producing emulsions with similar droplet size as full fat products [23]–[25]. Particle stabilized emulsions have been widely studied as a pathway for making low calorie food products. Especially fat particles and crystals are known to form stable Pickering emulsions [26]–[28]. Chocolate products using cocoa particles as emulsion stabilizers have also been used to incorporate water in chocolate products [16], [29]–[34]. The stability of Pickering emulsions is influenced by particle shape and size, and especially by surface properties like the wettability of the particles by the surrounding fluids [35]. No study has been found that uses a hydrophobic substance to create a sample spanning network induced by capillary bridges between particles in a suspension for food products. The addition of a fatty acid to the water continuous suspension of cocoa particles may result in capillary bridging or lead to the adsorption of the fatty acid to the surface then causing hydrophobic interactions between the particles. The latter were found to lead to spherical agglomeration in CaCO$_2$ suspensions [36]. The combination of both mechanisms could also be possible [37]. Capillary bridges can be induced by addition of a secondary fluid, but may also be created using fat already incorporated in the food particles. It has been reported that food particles like ground seeds with an oil content form agglomerates during, e.g., roasting due to the oil released. Particles stick together since oily capillary bridges are formed between them [38]. These investigations were restricted to dry granular matter and no work has been done yet regarding the behavior of such particles upon heating in suspension.

Natural organic particles are often used as Pickering stabilizers for food products but the wetting behavior seems to be ambiguous. For cocoa particles it was found that higher surface energy fluids like water exhibit higher contact angles than hydrophobic substances with a lower surface energy [39].

1.2 Scope of this work

The present thesis will be divided into two parts. First, oil continuous capillary suspension will be studied. It will be discussed how capillary forces can be used to control the stability and flow behavior
of food products. When particles are dispersed in oil the capillary force may be used to stabilize the particles or to introduce a higher viscosity by simply mixing a small amount of a secondary liquid to the suspension instead of using other more complex rheology control agents.

Simple model systems of particles in oil suspensions are investigated to determine the parameters that affect the network formation and the resulting rheological properties. For this, two food model systems were chosen: corn starch and cocoa particles in vegetable oil. Water is used as the secondary liquid in both admixtures. Food suspensions will be compared to simpler model systems from CaCO$_3$ and polyvinylchloride (PVC) particles. The effect of secondary phase viscosity and contact angle on suspension strength is systematically investigated. Since food products are hardly made out of one particle type, mixtures of hydrophilic and hydrophobic particles will be described. The influence of adsorption of water to the particles prior to dispersing in the oil phase and the resulting flow properties will also be discussed. It has been found that cocoa particles dispersed in cocoa butter do also form sample spanning networks when aqueous liquids are added which results in an increased heat stability. Melting behavior is documented with optical and rheological methods.

In the second part, we focus on the flow behavior of aqueous cocoa suspensions and how this can be adjusted by temperature or addition of a secondary liquid. A thorough rheological characterization is provided and potential mechanisms causing particle network formation are discussed. For this particles have been extracted with various solvents and the influence of added secondary fluid on the flow behavior was investigated. Structural analysis was performed using confocal and electro scanning microscopy as well as computer tomography and small angle neutron scattering.

This research should encourage the development of low fat cocoa spreads that are not based on gums, hydrogels or other polysaccharides, but provide structural stability through a particle network formed within the suspension, resulting in a creamy texture.
2 Theoretical background

This chapter will give an overview of the general theoretical background of the research in this thesis. First, particle interactions with a focus on capillary forces will be described. Then the phenomenon of capillary suspensions and the re-wetting behavior of the liquid phase at the particle surface will be explained. For the preparation of capillary suspensions mixing conditions play an important role, so an overview is also given here.

2.1 Particle interactions and stabilization in suspensions

Dispersed systems are heterogeneous mixtures from a dispersed phase in a bulk phase. If this dispersed phase is made from solid particles a suspension is created. Mixtures of two immiscible liquids are called emulsions.

The rheological behavior of the particle suspension is influenced by the particle volume fraction $\phi$, which is defined as follows,

$$\phi = \frac{V_d}{V_d + V_b}$$

(2.1)

with $V_d$, being the dispersed phase and $V_b$ the bulk phase of the system. Also particle form and particle interactions play an important role in suspension flow behavior. Particle interactions can be attractive or repulsive and mainly influence the stability of a suspension against sedimentation. Common particle interactions are extensively discussed in literature [40] and are summarized in the following:

- Van-der-Waals attraction
- Brownian motion
- Electrostatic repulsion
- Steric interactions
- Form closure
- Capillary forces

For this thesis steric interaction can be neglected, since particles are beyond the colloidal range and are not sterical stabilized. In oil continuous suspensions also no electrostatic interactions need to be taken into account. Interactions due to particle shape (form closure) as could be assumed for cocoa particles might have an influence on particle agglomeration, but this will be neglected in this thesis and therefore will not be discussed here.

Van-der-Waals forces result from dipole-dipole interactions induced by electron fluctuations at the atomic level. These Interactions can be permanent/permanent (Keesom interactions),
induced/permanent (Debye interactions) and induced/induced interactions (London interactions). Van-
der-Waals interactions can be calculated according to Hamaker [41] for two spheres of equal radius \( r \) and a small interparticle distance \( s \ll 2r \) [40]:

\[
F_{vdw} = -\frac{A_H r}{12 s^2} \tag{2.2}
\]

With \( A_H \) being the Hamaker constant which is a function of particle material and typically varies between \( 10^{-19} \) and \( 10^{-21} \) J [40]. The Van-der-Waals force is inversely proportional to the squared particle distance \( s \) and therefore strongly decreases when particles separate.

Capillary forces are well known in wet granular material as a particle bridging mechanism. In suspensions they have been known to cause particle bridging in spherical agglomeration processes [10] and have recently been introduced to also cause sample spanning network formation in suspensions [1].

The capillary force depends on the particle radii, the interfacial tension \( \gamma \) between the fluid and the surrounding gas in contact with the particles and the wetting angle \( \theta \) the fluid forms against the solid surface. In Figure 1 two spheres connected with a liquid bridge are shown.

![Figure 1: Schematic image of two spheres connected with a pendular liquid bridge. On the left particles are on contact and on the right they have the distance \( s \) (adapted from [42]).](image)

The force \( F_{cap} \) between two equally-sized spheres of radius \( r \) connected by a pendular bridge is given by [46], [47],

\[
F_{cap} = \pi r \frac{\cos \theta}{\cos \theta_{cap}} \left( 2r + \frac{d_{cap}}{2} \right) \tag{2.3}
\]

If it is assumed that the particles are in contact and that the pendular bridge is small compared to the particles \( s \ll r \) this is simplified to [43], [44]:

\[
F_{cap} = 2\pi r \Gamma \cos \theta \tag{2.4}
\]
For larger droplets, the force will depend on the volume of the wetting fluid, with various corrections given based on the filling angle and shape of the droplet. Additional corrections are available for surface roughness and particles that are not in contact [43], [45]. A detailed description of capillary forces in wet granular material is given by Herminghaus [42].

### 2.2 Capillary suspensions

Capillary suspensions can be described as a ternary mixture made up of a liquid bulk phase, a solid dispersed phase and a secondary immiscible fluid, that is fine dispersed in the system. This secondary fluid is forming capillary bridges between the particles, resulting in a capillary network within the suspensions. The formation of the particle network is changing the rheological behavior from a fluid-like state to a gelled state [1]. Capillary suspension can be subdivided into the pendular state and the capillary state. In pendular state suspensions the added secondary fluid preferentially wets the particles, whereas in capillary state the bulk fluid is the preferentially wetting fluid. For both suspensions, the added secondary liquid must be able to re-wet the particles, meaning that the liquid must be able to remove the bulk fluid from the particle surface (see section 2.3). Both the capillary state and pendular state are controlled by the capillary force and are strongly influenced by changes in the amount of secondary fluid and the interfacial and wetting properties of the ternary system.

![Figure 2: Ternary diagram of various particle–liquid–liquid systems showing stability regions as a function of the relative volume fractions (left). Corresponding diagrams for each state are plotted on the right [49].](image)

In Figure 2 capillary suspensions are compared and distinguished from other ternary liquid/liquid/solid systems. Capillary suspensions are fundamentally different from particle-stabilized (Pickering) emulsions. In stable Pickering emulsions, the particle volume \( r^3 \) is typically much smaller than the droplet volume \( V_d \), so that \( V_d/r^3 \gg 1 \) [46], [47], whereas in capillary suspensions the particle has a volume smaller or in the same range as the bridging volume \( V_d/r^3 \leq 1 \) [21]. In Pickering emulsions, capillary forces are stabilizing the emulsion droplets from coalescing and gelation is caused primarily
through the Van-der-Waals force acting between adjacent particles [48]. In capillary suspensions, the gelation is caused directly by the capillary force, creating a sample spanning particle network. Spherical agglomeration is often created when the secondary fluid volume fraction in pendular state suspensions is increased [6], but also a mechanical energy input is necessary.

The secondary fluid content in a capillary suspension is characterized by the so-called saturation of the preferentially wetting fluid $S$,

$$S = \frac{V_{\text{preferentially wetting fluid}}}{V_{\text{total fluid}}} \quad (2.5)$$

which is close to zero for the pendular state and approaches one for the capillary state [1], [44]. Saturation $S$ is chosen in analogy to wet granular material, where saturations is described as the ratio between total pore volume (air) and the total volume between the particles (fluid filled and unfilled pores). In the present case, the fluid that wets the particles less well corresponds to the air in wet granular media. Capillary suspensions where the secondary fluid is better wetting ($\theta < 90^\circ$) have a saturation $S$ close to 0, whereas in mixtures where the bulk fluid is the better wetting fluid ($\theta > 90^\circ$) saturations close to 1 are obtained.

The capillary force in a pendular state suspension can be calculated analogue to capillary bridges in wet granular matter (see eq.(2.4)), but surface tension must be replaced by the interfacial tension of the two liquids and the contact angle must be measured in the three phase system [1]–[3], [21], [50]. In capillary state suspensions the strength of the particle clusters can be calculated using the theory of Pickering emulsions. A single particle that is placed at an interface of two fluids is held with the energy $E$ given by [35],

$$E = \pi r^2 \Gamma_{1,2} \left(1 \pm \cos \theta_{1,2}\right)^2 \quad (2.6)$$

where the sign in the parentheses is negative for contact angles $< 90^\circ$ and positive for angles $> 90^\circ$. For removing this particle from the interface this Energy $E$ must be spend. This theory can be applied to capillary suspensions when the capillary network is seen as connected clusters of particles. In each cluster the particles are held at the interface of a small droplet as has been shown by Lauga and Brennes [51] who evaporated small particle stabilized emulsion droplets to form clusters connected by capillary forces, as shown in Figure 3.

Figure 3: Clusters formed by evaporation of Pickering stabilized droplets [51].
The yield stress \( \sigma_y \) of a pendular state suspension is related to the capillary force [3], [44],

\[
\sigma_y = f(\phi)g(V, s) \frac{\Gamma_{1,2} \cos \theta_{1,2}}{r}
\]

(2.7)

where \( f(\phi) \) is a function of the volume fraction of the particles and depends on the number of contacts per particle. The volume of the bridge \( V \) and the distance between the two particles \( s \) are included in the function \( g(V, s) \).

Few investigations have been performed on the formation of particle clusters and networks formed in capillary suspensions. Calculations have been performed by Koos et al. [21] using the computational code surface evolver by Brakke [52]. Close packed particle clusters have been calculated resulting in a state diagram showing minimum energy regions for particle clusters. It is shown that a rearrangement of particle clusters is happening when the amount of secondary fluid is increased. Tetrahedral structures are formed for low secondary fluid volume increasing up to combinations of octahedral and tetrahedral structures. Structures have been investigated up to a maximum of \( n = 7 \) particles. Further calculations on particle clustering [53] and network formation [6], [54] can be found in literature.

![State diagram of minimum energy regions as a function of contact angle \( \theta \) and secondary fluid volume \( V_i \).](image)

**Figure 4**: State diagram of minimum energy regions as a function of contact angle \( \theta \) and secondary fluid volume \( V_i \).

### 2.3 Wetting and rewetting

The basics of interfaces must be understood to explain the wetting behavior of a liquid on a solid surface. A system of two fluids strives to always find their state of minimal potential energy, meaning to minimize their interface. Work must be applied to the system to expand the interface, which results in an increased inner energy or free enthalpy \( G \). The interfacial tension \( \Gamma \) or interfacial energy per area can be defined as:
At a curved interface the pressure difference between the two phases must be taken into account. This is expressed via the Young-Laplace equation, which equals zero for even surfaces,

\[ \Delta p = \Gamma \left( \frac{1}{r_1} + \frac{1}{r_2} \right) \]  

(2.9)

with \( r_1 \) and \( r_2 \) being the principal curvatures of the surface. The interfacial tension of a solid surface with a liquid cannot be measured directly. It is calculated via a force balance at the three-phase contact line. The wetting of a liquid on a solid substrate can therefore be described by eq. (2.10), which is called the Young equation:

\[ \Gamma_{2S} - \Gamma_{1S} = \Gamma_{12} \cos \theta_{1,2} \]  

(2.10)

with \( \theta_{1,2} \) being the equilibrium three-phase contact angle, where the drop shape on the solid corresponds to a minimum free energy [55]. Surface tensions \( \Gamma_{1S} \) and \( \Gamma_{2S} \) are measured between the solid and the two fluids “1” and “2”, \( \Gamma_{12} \) corresponds to the interfacial tension between the two fluids. For the application in capillary suspension these two fluids refer to the bulk and the secondary fluid. In order to form a capillary suspension the added secondary fluid must be able to rewet the particles meaning to replace the attached liquid from the particle surface. Displacement of fluid 2 by fluid 1 is feasible if the difference between the two wetting tensions is positive [56]:

\[ \Gamma_{2V} \cos \theta_{2V} - \Gamma_{1V} \cos \theta_{1V} > 0 \]  

(2.11)

Interfacial tensions \( \Gamma_{2V} \) and \( \Gamma_{1V} \) as well as corresponding contact angles \( \theta_{iV} \) are measured against air. The contact angle \( \theta_{1,2} \) the two liquids form with the solid particle can be calculated using the modified Young-Dupré equation [1], [57]:

\[ \cos \theta_{1,2} = \frac{\Gamma_{2V} \cos \theta_{2V} - \Gamma_{1V} \cos \theta_{1V}}{\Gamma_{12}} \]  

(2.12)

Corrections of the contact angle for porous and rough surfaces can be made using the model by Wenzel [58] as well as the model of Cassie and Baxter [59]. For heterogeneous surfaces as used in this study the Cassie and Baxter model is more appropriate. The idea behind this model is to define the contact angle of a droplet sitting on a composite material, where the droplet is in contact with an area fraction \( \zeta_1 \) of material 1 and \( \zeta_2 \) of material 2. This result in a corrected contact angle \( \theta_c \) calculated as

\[ \theta_c = \zeta_1 \cos \theta_1 + \zeta_2 \cos \theta_2 \]  

(2.13)
To apply this to a rough surface it is assumed that the droplet sits on the peaks of the surface leaving the surface valley free of the liquid. So the droplet sits on a composite material made from the substrate and the surrounding fluid. If the surrounding fluid is air or oil and the droplet is water, the contact angle the surrounding fluid forms with the wetting liquid is 180°. In this case the formula simplifies to [59], [60]:

\[ \theta_c = \zeta_1 \cos \theta_1 + \zeta_1 - 1 \]  

(2.14)

### 2.4 Droplet breakup and mixing conditions

When the secondary fluid is mixed to the particle suspension it needs to be finely distributed. The simplest approach is to assume laminar droplet breakup. For this the theory of Grace [61] can be applied. The Grace diagram allows determining the necessary shear rate for the breakup of spherical droplets in a quasi steady homogeneous flow. For this the capillary number \( Ca \) can be taken into account, describing the relationship of viscous forces and the surface tension between two immiscible fluids:

\[ Ca = \frac{\eta \dot{\gamma} r}{\Gamma} \]  

(2.15)

The critical capillary number \( Ca_{crit} \) describes the size \( r_{crit} \) of droplets formed at given \( \dot{\gamma} \), \( \eta \) and \( \Gamma \). For capillary numbers smaller than \( Ca_{crit} \) only droplet deformation can be detected.

**Figure 5: Droplet breakup criteria for a single droplet undergoing shear and elongational stress** [61].

Mixing a suspension in a beaker using a dispenser mixer blade turbulent mixing conditions can be assumed. Here droplet breakup origins from eddies that superimpose the main flow. A closer look on turbulence theory can be found in the literature [62], [63]. For an estimate of droplet size of the secondary fluid that is mixed to the bulk suspensions in a beaker using a mixing blade, the theory of a stirrer vessel can be applied. Here the minimum droplet diameter accessible in emulsion preparation
can be calculated by applying dimensional analysis [64]. This leads to a dependency of the minimum mean droplet diameter $d_{dr,m}$ on mixing conditions [65]:

$$d_{dr,m} = N^{-1.2} d_{mix}^{-0.8} C_1 \left[ \frac{\Gamma}{\rho_i} \right]^{-0.6} \left( 1 + C_2 \phi_d \right)$$  \hspace{1cm} (2.16)

with $N$ being the rotational speed, $d_{mix}$ the diameter of the mixing blade, $\phi_d$ the volume fraction of the dispersed phase and $C_1$ and $C_2$ constants that must be determined experimentally. Experimental tests as well as theoretical considerations of turbulence procure the constants needed for full characterization of the system. It can be seen that the minimum droplet diameter is a function of mixing conditions ($N, d_{mix}$) that were kept constant for all preparations as well as material properties like interfacial tension and liquid phase density, which are varied for different systems.
3 Materials and methods

3.1 Materials

In this section an overview of the different particles and liquids used in the experimental section will be given.

3.1.1 Particles

Different particles were used in this study. Starch and cocoa particles represent food systems, while calcium carbonate and polyvinylchloride (PVC) particles serve as model system in the first part of the thesis (oil continuous systems). The particle size was measured using a LALLS (Low Angle Laser Light Scattering, Sympatec HELOS H0309) while the particles were suspended in paraffin oil or water and subjected to ultrasonic dispersion using a Sympatec QUIXEL unit. The particle size distributions of particles described below are plotted in Figure 6.

![Figure 6: Particle size distribution of particles used in this study.](image)

Cocoa particles

Cocoa solids are a product obtained by the processing of cocoa beans. The cocoa tree is growing in West Africa, South-East Asia and South America. The beans are fermented and dried before they are shipped to the chocolate manufacturing countries. Especially, the fermenting process which is done directly after harvesting, can influence the cocoa flavor. The process is hardly industrialized and the method strongly depends on growing county. In the chocolate factories the beans are roasted, winnowed and ground. The cocoa butter is pressed out of the product leaving the cocoa press cake as a residual. The cocoa powder is now produced by milling the cake. The fat content of the cocoa powder is adjusted in the pressing process step and can vary between 10% and 22%. Since cocoa particles are
a natural product and highly dependent on origin country of the beans, weather conditions and fermenting process, quality and properties are varying with every batch [66].

Two types of cocoa particles were utilized in this study. As a standard, cocoa powder 10/12 SN donated by ADM Schokinag GmbH Mannheim was used. The cocoa particles were non-alkalized and had a fat content of 11%. All ingredients are listed in Table 1. As a comparison a second product was used. Cocoa particles (20-22) provided by Storck GmbH, Halle (Saale) were also non-alkalized and had a fat content of 22%. Cocoa particles exhibit a density of $\rho_{\text{cocoa}} = 1.44 \text{ g/cm}^3$.

| Table 1: Ingredients of cocoa solids (10/12 SN) provided by ADM Schokinag. |
|-----------------|---|---|---|---|---|
| Ingredient | Protein | Sugar | Starch | Fat | Fibre |
| Per 100g | 22.0g | 2.0g | 10.0g | 11.0g | 31.1g |

As the processing of cocoa implies, every cocoa particle is different in shape, size and composition. They exhibit a very rough and porous surface and can be agglomerated as can be seen in Figure 7.

![Figure 7: Scanning electron microscopy (SEM) image of cocoa solids](image)

Starch

Starch granules are the major storage of carbohydrates in plants. They are quasi-crystalline, water insoluble and dense [67]. The two components amylase and amylopectin are forming the starch granule. Amylose consists of 1000-10000 D-glucopyranosyl units that are bond with $\alpha$-D-(1-4) glucopyranosyl units and is a mostly linear molecule. Amylopectin is highly branched and is also linked together by $\alpha$-D-(1-4) glucopyranosyl residues, but with (1-6) bonds at the branch points [68]. Amylose as well as amylopectin can form crystalline structures that origin from a helix or doublehelix structure of the molecules. Amorphous and crystalline structures are arranged in a lamellar structure that form layers in the granule [69].

The starch granules used in this study were maize starch G*Gel 03401 provided by Cargill Deutschland GmbH, Krefeld. The density of the starch is given to be $\rho_{\text{starch}} = 1.5 \text{ g/cm}^3$. 
Calcium carbonate particles

Fumed calcium carbonate particles are used in this study as a model system. Particles are used with their naturally hydrophilic surface (SOCAL 31) with an average mean particle diameter of $x_{50} = 70$ nm as well as hydrophobically modified particles (SOCAL U1S1) with an average mean particle diameter of $x_{50} = 80$ nm. Both products were produced by Solvay Chemicals, Hannover, Germany. SOCAL is an odorless, white powder with a density of $\rho_{SOCAL} = 2.71 \text{ g/cm}^3$. The hydrophobic particles are modified using fatty acids which are bond to the particle surface.

PVC/PVA particles

Polyvinylchloride (PVC) is an amorphous, plastic polymer that is used in a wide range of applications. Polyvinyl acetate is also a thermoplastic polymer. Particles Vinnolit C 12/62 V used here are prepared from a mixture of PVC and PVA (12% PVA) and are provided by Vinnolit GmbH, Ismaning, Germany. The particles have a spherical and non-porous shape and a narrow particle size distribution (see Figure 6). The density of the particles is $\rho_{PVC} = 1.38 \text{ g/cm}^3$.

3.1.2 Hydrophobic substances

Sunflower oil

Commercial sunflower oil was used in this study (“Ja!” Rewe Markt GmbH). Sunflower oil consists of triglycerides, which have free fatty acids attached to glycerol backbone. In sunflower oil they are mainly formed from double unsaturated linoleic acid (C 18:2) and unsaturated oleic acid (C18:1). A small fraction of free fatty acids (~1.8 %) are present in the oil. Measured viscosity was $\eta = 39 \text{ mPa s}$ with a density of $\rho = 0.92 \text{ g/cm}^3$.

Cocoa butter

Cocoa butter is a mixture of different triglycerides, which vary in fatty acid composition and therefore exhibit different properties in melting behavior and range.

Table 2: Different crystal forms of cocoa butter (adapted from [66]).

<table>
<thead>
<tr>
<th>Greek label</th>
<th>$\gamma$</th>
<th>$\alpha$</th>
<th>$\beta_2^\prime$</th>
<th>$\beta_1^\prime$</th>
<th>$\beta_2$</th>
<th>$\beta_2$</th>
</tr>
</thead>
<tbody>
<tr>
<td>Roman label</td>
<td>I</td>
<td>II</td>
<td>III</td>
<td>IV</td>
<td>V</td>
<td>IV</td>
</tr>
<tr>
<td>Temp. range</td>
<td>16-18°C</td>
<td>22-24°C</td>
<td>24-26°C</td>
<td>26-28°C</td>
<td>32-34°C</td>
<td>34-36°C</td>
</tr>
</tbody>
</table>

loose packing | dense packing

unstable form | stable form
Most cocoa butter consists of 35% oleic acid (C18:1), about 34% stearic acid (C18:0) and 26% palmitic acid (C16:0). Because of the different triglyceride compositions, the fats show polymorphism. This means that they can crystallize in a number of different ways, where some crystallize in double-chain packing and others in triple-chain packing. Cocoa butter can possess six different crystal forms that are labelled with Greek or Roman letters [66]. An overview of the different forms and melting temperatures are given in Table 2, with $\gamma$ being a crystal form in this section. Cocoa butter used in this thesis was purchased from Spiceworld GmbH, Salzburg, Austria.

Other hydrophobic substances

In Table 3 the properties of hydrophobic substances used in this thesis are summarized.

Table 3: Properties of hydrophobic substances used as secondary fluids for water continuous suspensions.

<table>
<thead>
<tr>
<th>Substance</th>
<th>Purchased from</th>
<th>Chemical Formular</th>
<th>Density</th>
<th>Viscosity</th>
</tr>
</thead>
<tbody>
<tr>
<td>Linoleic acid</td>
<td>Sigma-Aldrich, St. Louis, USA</td>
<td>C_{18}H_{32}O_2</td>
<td>0.89 g/cm^3</td>
<td>27.0 mPa s</td>
</tr>
<tr>
<td>Oleic acid</td>
<td>Alfar Aesar, Karlsruhe, Germany</td>
<td>C_{18}H_{34}O_2</td>
<td>0.89 g/cm^3</td>
<td>33.7 mPa s</td>
</tr>
<tr>
<td>Octanol</td>
<td>Merck KGaA, Darmstadt, Germany</td>
<td>C_{8}H_{18}O</td>
<td>0.83 g/cm^3</td>
<td>85.0 mPa s</td>
</tr>
<tr>
<td>Octanoic acid</td>
<td>Alfar Aesar, Karlsruhe, Germany</td>
<td>C_{8}H_{16}O_2</td>
<td>0.91 g/cm^3</td>
<td>58.8 mPa s</td>
</tr>
<tr>
<td>Octyl acetate</td>
<td>Alfar Aesar, Karlsruhe, Germany</td>
<td>C_{10}H_{20}O_2</td>
<td>0.87 g/cm^3</td>
<td>1.8 mPa s</td>
</tr>
<tr>
<td>Silicon oil AK 10</td>
<td>Wacker Chemie AG, München, Germany</td>
<td>Si (CH_{3})<em>3 Si (CH</em>{3})<em>n Si (CH</em>{3})_3</td>
<td>0.93 g/cm^3</td>
<td>9.3 mPa s</td>
</tr>
</tbody>
</table>

3.1.3 Hydrophilic substances

As hydrophilic liquids water and aqueous solutions of glycerol, fructose and PEO were used. Glycerol is an alcohol with three hydroxyl groups and the chemical name propanetriol. It is widely used in food and pharmaceutical industry as a humectant, lubricant or solvent. It is completely miscible with water. Viscosity and density data for various mixtures with water are given in Table 4.
Fructose is a monosaccharide with the systematic name 1,3,4,5,6-Pentahydroxy-2-hexanone. It is found in many plants and is widely used as a sweetener in food products. The crystalline substance is soluble in water up to 78 wt% and increases the viscosity of the mixture as can be found in Table 4.

Polyethylene oxide (PEO) is a water soluble polymer with the chemical formula C_{2n}H_{4n}O_{n+1}. In this study PEO with a molecular mass of 10^6 g/mol was used with different concentrations dissolved in water. PEO was purchased from Merck KGaA, Darmstadt, Germany.

**Table 4: Viscosity at $\dot{\gamma} = 100$ s$^{-1}$ and density of aqueous mixtures from glycerol, fructose and PEO.**

<table>
<thead>
<tr>
<th>Weight fraction</th>
<th>Viscosity ($\dot{\gamma} = 100$ s$^{-1}$)</th>
<th>Density</th>
</tr>
</thead>
<tbody>
<tr>
<td>Water</td>
<td>1 mPa s</td>
<td>0.997 ± 0.0005 g/cm³</td>
</tr>
<tr>
<td>25%</td>
<td>2.5 mPa s</td>
<td>1.071 ± 0.0006 g/cm³</td>
</tr>
<tr>
<td>50%</td>
<td>8.5 mPa s</td>
<td>1.124 ± 0.0015 g/cm³</td>
</tr>
<tr>
<td>75%</td>
<td>53.3 mPa s</td>
<td>1.205 ± 0.0003 g/cm³</td>
</tr>
<tr>
<td>100%</td>
<td>1100 mPa s</td>
<td>1.259 ± 0.0001 g/cm³</td>
</tr>
<tr>
<td>Glycerol</td>
<td></td>
<td></td>
</tr>
<tr>
<td>25%</td>
<td>2.2 mPa s</td>
<td>1.103 ± 0.0003 g/cm³</td>
</tr>
<tr>
<td>50%</td>
<td>10.9 mPa s</td>
<td>1.231 ± 0.0007 g/cm³</td>
</tr>
<tr>
<td>75%</td>
<td>870 mPa s</td>
<td>1.379 ± 0.0032 g/cm³</td>
</tr>
<tr>
<td>Fructose</td>
<td></td>
<td></td>
</tr>
<tr>
<td>0.1%</td>
<td>2 mPa s</td>
<td>0.997 ± 0.006 g/cm³</td>
</tr>
<tr>
<td>0.5%</td>
<td>9.6 mPa s</td>
<td>0.998 ± 0.0002 g/cm³</td>
</tr>
<tr>
<td>1.0%</td>
<td>42 mPa s</td>
<td>0.999 ± 0.0002 g/cm³</td>
</tr>
<tr>
<td>1.5%</td>
<td>130 mPa s</td>
<td>1.000 ± 0.001 g/cm³</td>
</tr>
<tr>
<td>2.5%</td>
<td>990 mPa s</td>
<td>1.001 ± 0.001 g/cm³</td>
</tr>
<tr>
<td>3.0%</td>
<td>2100 mPa s</td>
<td>1.001 ± 0.005 g/cm³</td>
</tr>
</tbody>
</table>
3.2 Methods

3.2.1 Rheological measurements

All measurements were conducted using a rotational rheometer (MARS II, Thermo Fisher Scientific, Karlsruhe, Germany) with a sandblasted rough plate-plate geometry (35 mm diameter) and a gap size of 1 mm if not stated otherwise. Measurements where done in triplicate and the calculated standard deviations are shown as error bars in the diagrams. For water continuous samples, the environment of the probes filled in the rheometer gap was water saturated to prevent drying of the sample while measuring.

Rheological measurements are based on the plate-plate model pictured in Figure 8. In stress controlled rheometers one moving plate with radius \( R \) is turning with an applied torque around its axis over a second plate with a gap size \( h \). This torque is inducing the angular frequency \( \omega \). The sample is filled in the gap and adhesion at the plate wall is assumed and mandatory. The shear rate \( \dot{\gamma} \) is defined as follows:

\[
\dot{\gamma}(x) = \frac{\omega x}{h} \quad 0 < x < R
\]

The rheometer used here is a stress-controlled measurement device where the deformation is detected at given torque, which is proportional to the shear stress.

![Figure 8: Schematic diagram of a plate-plate setup of a rotational rheometer with the gap size \( h \) and radius \( R \).](image)

The strength of capillary suspensions is characterized here through the apparent yield stress (for brevity subsequently termed yield stress). The yield stress is defined as the stress at which the sample begins to flow irreversibly. These measurements were conducted using stress-steps, where the yield stress was found as the point at which the slope of the logarithmic deformation as a function of the logarithmic shear stress changes from a low value \( \approx 1 \) to a high value \( >>1 \). For this the deformation \( \gamma \) was plotted over the shear stress \( \sigma \) and the point of intersection between two adjusted tangents was determined using the RheoWin Software from Thermo Fischer. Karlsruhe, Germany (see Figure 9).
The sample viscosity was measured using shear rate steps. The shear rate was increased incrementally and held at a constant value, for at least 10 s and for a maximum of 100 s, until the stress reached a steady state, \((d\sigma/\sigma)/dt < 0.01\) s\(^{-1}\), before recording the shear stress and corresponding viscosity.

**Figure 9:** Example for yield stress evaluation: Socal particles \(\phi = 10\%\) dispersed in sunflower oil.

For small amplitude oscillatory measurements a sinusoidal stress signal was applied to the sample and the responding deformation was detected. The linear viscoelastic behavior of the sample is then characterized in terms of the storage modulus

\[
G' = \frac{\tilde{\sigma}}{\tilde{\gamma}} \cos \delta
\]  
(3.2)

and the loss modulus

\[
G'' = \frac{\tilde{\sigma}}{\tilde{\gamma}} \sin \delta
\]  
(3.3)

where \(\tilde{\gamma}\) is the strain amplitude, \(\tilde{\sigma}\) the stress amplitude and \(\delta\) the phase shift between applied and responded signal. In the linear-viscoelastic regime (LVE) \(G'\) and \(G''\) are independent of applied deformation or stress and for the suspensions with dominating attractive particle interactions \(G'\), which describes the elastic part of the deformation, is dominating over the viscous dissipation (\(G''\)). The LVE regime is determined via amplitude sweeps. The deformation or the shear stress is increased at constant frequency over time and the critical point (\(\gamma_{\text{crit}}\) and \(\sigma_{\text{crit}}\)) where \(G'\) is not constant anymore (with a deviation of 10%) is determined as the end of the LVE regime and the response signal becomes non-linear. An example is given in Figure 10. Frequency sweeps were always performed in the LVE regime. Viscoelastic properties are measured over a wide frequency range to characterize the gel-like behavior of the suspension.
Materials and methods

Figure 10: Example for critical shear stress evaluation: cocoa particles dispersed in water and stored at 30°C, \( \phi = 10\% \).

Vane geometry

For highly filled yield stress fluids with an oil continuous phase the plate-plate geometry is not appropriate due to wall slip. In order to measure the yield stress for such suspensions, the vane geometry has been used. When using the vane setup not viscometric flow can be assumed. Only a “tearing stress” can be measured which is put on a level with the yield stress of the suspension. The vane used in this study has a diameter \( d_{vane} = 10\text{mm} \) with 4 blades and a blade height \( h = 20\text{ mm} \) (see Figure 11).

Figure 11: Vane used for yield stress measurements for oil continuous suspensions.
3.2.2 Microscopy

An inverse Leica TCS SP8 confocal laser scanning microscope (Leica Microsystems, Mannheim, Germany) was used for microscopic images. The microscope was supplied with lasers of 488 nm and 552 nm wavelength. Figure 12 shows a schematic image of the optical path of the microscope.

The advantage of using a laser scanning microscope is given through a specific signal, high contrast and no particles out of focus light. The fluorescent substance or dye absorbs a photon of the particular excitation wavelength and emits at different specific wavelength. Since the emitted photon has less energy, a longer wavelength is generated; this is described in Planck’s law [70]. The oil phase in this study was labeled using Nile Red (purchased from Sigma Aldrich). The absorption and emission maxima are highly dependent on the solvent and vary with the different hydrophobic phases used in this study. The water phase was stained with Promofluor-488 Carboxylic acid (Promokine, Heidelberg, Germany) which inhibits an absorption maximum of 490 nm when used with the 488 nm laser and Rhodamine B (purchased from Carl Roth, Germany), where the absorption maximum in aqueous solution is between 542 nm and 554 nm.

![Figure 12: Schematic image of the laser path through the microscope [71].](image)

The inverse fluorescence microscope Axio Observer D1 by Zeiss (Carl Zeiss AG, Oberkochen, Germany) was also used in this study. It was set up with a 365 nm laser and also a white light camera was attached, allowing the creation of overlaid images from laser and white light signals.

3.2.3 Contact angle and interfacial tension

Contact angle

The contact angle one fluid forms a particle surface in the presence of a secondary fluid is calculated by using the sessile drop method. Contact angle was measured directly in the three phase system [72],
Powder was pressed to a tablet using a hand press machine and soaked with the bulk fluid to ensure that no open pores are left in the pellet.

Figure 13: Schematic images of contact angle measurements when the high density fluid is the bulk phase (left) and when the low density fluid is the bulk phase (right).

Due to density differences, two setups were used for contact angle measurements. For oil continuous systems, the pressed pellet was placed in a glass container filled with oil and a drop of secondary fluid was placed onto the particle surface. If water is the continuous phase, the tablet was mounted in the glass container as described in Figure 13 and secondary fluid was injected under the pellet with a needle [74]. Contact angle was measured immediately through the secondary phase droplet and calculated via image analysis (Drop shape analysis software by Krüss GmbH, Hamburg). The droplet shape was fitted using a Laplace-Young approach and the angle between the manually set baseline was displayed. Given value have been corrected by taking surface roughness into account (see section 2.3).

**Interfacial tension**

Interfacial tension $\gamma_{ow}$ between the two fluid phases was measured using the pendant drop method. The droplet is hanging on a needle which was placed in the other fluid. The fluid with the lower density is used as the surrounding fluid and the droplet is formed using the higher density fluid. The droplet shape is captured via image analysis [75]. The correlation between the geometry of the formed droplet and the interfacial tension is based on the Laplace equation (see eq. (2.9)) and the calculation is performed using a nonlinear differential equation which refers to Bashforth and Adam [76]:

$$\frac{1}{r_{dr}/h_{dr}} + \frac{\sin \Theta_{dr}}{x/h_{dr}} = -B \frac{z}{h_{dr}} + 2$$

with

$$B = \frac{h_{dr}^2 g \Delta \rho}{B}$$

where $h_{dr}$ is a geometrical parameter and $r_{dr}$ the radius of curvature at coordinates $x,z$ and $\Theta_{dr}$, which are depicted in Figure 14, g is the gravitational constant and $\Delta \rho$ the density difference between the droplet and the surrounding fluid.
3.2.4 Standard sample preparation

Capillary suspensions were prepared as follows: particles were mixed into the bulk fluid using a turbulent beater blade (at 500–2000 rpm for 20 min) until a uniform suspension was created. This mixture was then held under vacuum (100 mbar) for a few minutes to remove air bubbles. The secondary fluid was added to the suspension and thoroughly mixed using a dispenser stirrer (at 1700 rpm for 5 min). The samples were held in airtight containers until the measurements were completed. All samples were measured after 24 h of storage if not stated otherwise.

3.2.5 Particle treatment

Pre-conditioning of particles

Hygroscopic particles can absorb water when stored in a humid environment. Sorption kinetics are dependent on the vapor pressure around the particles and particle surface chemistry. In this setup, particles are stored in a closed container over water at 20°C so the system is in an equilibrium state. The saturation vapor pressure \( p_{w, \text{max}} \) over water can be calculated using the Magnus- Equation:

\[
p_{w, \text{max}}(T) = 6.112 \ hPa \exp \left( \frac{17.62 T}{243.12^2 + T} \right) \quad (3.6)
\]

For 20°C a value of \( p_{w, \text{max}} = 2.3 \text{ kPa} = 0.023 \text{ bar} \) is calculated. So the maximum absolute humidity of \( \varphi_{w, \text{max}} \) can be calculated using following equation:

\[
\varphi_{w, \text{max}} = \frac{p_{w, \text{max}}}{R_w T} \quad (3.7)
\]

A maximum absolute humidity of 17.2 g/m³ for 20°C is determined. Vapor pressure \( p_w \) and therefore humidity can be varied by replacing the free water surface with saturated salt solutions. With
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decreased vapor pressure, water sorption of the particles for a given time is reduced compared to particles stored over pure water. Relative humidity

\[ \varphi_{rel} = \frac{p_w}{p_{w,\text{max}}} \]  

(3.8)

at 20°C for saturated solutions as well as solubility of salts in water are listed in Table 5.

Sorption at a particle surface is described using a sorption isotherm, which describes the equilibrium state of a solid surface with the environment. At constant temperature molecules of water adsorb and desorb in the same amount. Different models can be used to describe the sorption isotherms of surfaces depending on multilayer or monolayer absorption. The most common model used is the BET (Brunauer–Emmett–Teller) model since it takes mono- as well as multilayer absorption into account. It has been adjusted and improved to fit for food systems [78], for example the equation given by Kats [79]. The Guggenheim, Anderson, de Boer (GAB) model [80] is also widely used in literature and has been modified to give a versatile fit to measured data [81]–[84].

Table 5: Relative humidity of saturated salt solutions and the solubility of salt in water.

<table>
<thead>
<tr>
<th>Type of salt</th>
<th>Relative humidity ( \varphi_{rel} ) at 20°C for saturated solutions</th>
<th>solubility</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lithium chloride (LiCl)</td>
<td>12 %</td>
<td>832 g/l</td>
</tr>
<tr>
<td>Sodium chloride (NaCl)</td>
<td>76 %</td>
<td>358 g/l</td>
</tr>
<tr>
<td>Magnesium chloride (MgCl2)</td>
<td>33%</td>
<td>108 g/l</td>
</tr>
</tbody>
</table>

For this study particles were stored over distilled water in a closed vessel for 2 to 21 days at 20°C. The volume of the particles (21 ml) was kept constant, since the real surface of the particles could not be detected. Water uptake of conditioned granules was determined by the weight gain of granules during storage. So the amount of water was controlled by time and not via vapor pressure variation at equilibrium state. Water sorption over time for different particles is shown in Figure 15. For Socal U1S1 particles which adsorb water quickly over time, saturated salt solutions are used to allow a more precise water sorption. All samples with saturated salt solutions were stored for two days and the amount of water was therefore controlled by the vapor pressure in the closed vessel.
Figure 15: Absorbed amount of water for different particles over time. Particles had been stored over distilled water at 20°C.

Hydrophobisation of starch particles

For experiments regarding contact angle variation, the starch granules were treated with octenyl succinic anhydride (OSA) bought from Carl Roth to make the particles more hydrophobic [85]. For hydrophobization, the starch granules were dispersed in water and OSA was added in different concentrations (0%, 1%, 2% and 3 wt.% of the particle mass). Mixtures were stirred for 3 h and then centrifuged for 20 min. at 5000 min⁻¹. The starch was washed three times by dispersing the particles in water and repeating the centrifugation step. Particles were then dried at 40°C for 72 h. Before dispersing the treated starch granules in oil for sample preparation, the powder was ground using a mortar and pestle to carefully break agglomerated formed during the drying process.

Particle extraction and dying

Cocoa particles contain water soluble, oil soluble (cocoa butter) and non soluble substances as mentioned before in section 3.1.1. For capillary wetting the contact angle is important and this is influenced by the soluble substances in the particle. The particles have been extracted via cross-flow liquid-liquid extraction using a centrifuge. As solvents water, ethanol and n-hexane were used. 27.5 g of solvent was mixed with 6 g of particles and placed on a shaking table for 30 min. The mixture was then centrifuged (Hettlich Lab Technology, Universal 320) at 8980 g for 15 min. The solvent was disposed and replaced with new solvent. This procedure was repeated 3 times. Particles were dried over silica for at least 2 days and were then milled using a mortar. Particle size distribution of natural and extracted particles (see section 3.2.5) is given in Figure 16.

For dying of particles the procedure was similar: particles were dispersed in water and the dye was added. For each experiment 0.5 g of dye and 30 g of particles were added in water. After reaction time, the particles were washed and dried as written above. Cocoa particles were dyed with Toluidine blue, starch and Socal particles with Rhodamine B.
3.2.6 Small angle neutron scattering

Small angle neutron scattering (SANS) experiments were performed at the KWS-1 instrument of the Jülich center for neutron sciences (JCNS) outstation at the FRM II, Garching, Germany. For the experiments, a wavelength of $\lambda = 0.7$ nm ($\Delta \lambda / \lambda = 20\%$) and sample-detector distances of 1.4, 8 and 20 m were used thus covering the q-range from $q = 0.0012$ Å$^{-1}$ to $q = 0.23$ Å$^{-1}$, with $q$ being defined as,

$$q = \frac{4\pi \sin \Theta_{\text{scat}}}{\lambda} \quad (3.9)$$

where $\Theta_{\text{scat}}$ is the scattering angle. The samples were mounted in standard glass cuvettes with a light path of 0.5 mm in a double-walled aluminum sample holder. Data reduction was done using the software QtiKWS provided by JCNS. SANS curves were fitted with a Gaussian model to analyze the peak shift at large q values. Suspensions were prepared in D$_2$O instead of H$_2$O for these experiments in order to increase the scattering contrast between the organic particles and the surrounding fluid. For the scattering length density calculation, the cocoa particles are treated as cellulose.

3.2.7 Heat stability

Sample preparation

The model chocolate was prepared as follows: Cocoa butter was melted in a water bath at 50°C and cocoa solids were added under stirring. Water or other secondary fluids were heated separately and added to the bulk suspension also under stirring at 800 rps for 5 minutes. So prepared model chocolate was poured into a cylindrical silicon form with a diameter and height of 25 mm. Samples were stored to cool for 24 h at 20°C. The so formed blocks were analyzed using optical and rheological methods.
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Optical melting experiments

For optical measurements the model chocolate block was placed in an oven at 50°C and pictures were taken every 5 minutes. The height of the chocolate block was determined using the open source software image j. Height was then normalized by the initial height and plotted over time for varied water contents.

Rheological melting experiments

Rheological measurements were carried out on a Haake Rheometer RS 150 by varying the temperature using a Peltier element in the bottom plate. Samples were melted on the bottom plate at 40°C and then sheared in sinusoidal oscillation mode for 25 minutes at 25°C and \( f = 1 \) Hz to let the sample cool down. Subsequently, temperature was increased linearly from 25°C to 45°C in 46 min and \( G' \) and \( G'' \) were detected at a constant frequency of \( f = 1 \) Hz and a shear stress of \( \tau = 0.1 \) Pa.
4 Results and discussion

4.1 Oil continuous suspensions

The following sections focus on capillary suspensions that were prepared and investigated by dispersing starch and cocoa particles in oil. A closer look will be taken on the rheological behaviour and what factors are influencing suspension stability. Starch particles can serve as a well defined food model system for capillary suspensions. They exhibit properties typical for food particles, as they are hygroscopic, irregular shaped and have a wide particle size distribution. Also suspensions from cocoa particles have been investigated. These particles are additionally porous, exhibit various form factors in one batch and consist of different components (see section 3.1.1) which results in heterogeneous surface properties. These organic suspensions are also compared to model systems made from calcium carbonate, as well as PVC and glass particles. Parts of the results described in the following sections have been published in Food Hydrocolloid [86].

4.1.1 Wetting behavior of starch and cocoa particles in oil

Capillary bridging in suspension as well as wetting behavior of the particles has been imaged using two different microscopes as described in section 3.2.2.

A starch suspension was prepared by dispersing starch granules in sunflower oil at a volume fraction of $\phi = 0.33$. The contact angle of added water in the three phase system was measured to be $\theta = 129 \pm 14^\circ$ via the sessile drop method. Although starch granules do have a hydrophilic nature, the direct measurement of the contact angle in the three-phase system shows a wetting angel greater than 90°. Since the angle is not measured using a single particle, we have to consider that tablet surface roughness and porosity could play a role. Corresponding calculations have been performed using a modified Cassie equation (see equ. (2.14)) with a area fraction of $\zeta = 0.5$ and the contact angle reduces to $\theta = 116^\circ$. Still, contact angle measurements using the sessile drop method can be defective due to water sorption and tablet porosity.

![Figure 17: CLSM images of starch particles dispersed in oil and brought in contact to a water interface. The oil phase is displayed in red while water is colored in green.](image-url)
Therefore, confocal microscopic (CLSM) images have been taken to have a closer look at the wetting and adsorption behavior of the starch particles. For this, the granules have been dispersed in oil and an interface with water was created by adding a droplet of the oil/starch mixture next to a water droplet on an object slide. The oil phase was dyed with Nile Red and the water phase with Promofluor-488. Particles were one time dispersed in oil (appears red in the image) and the other time in water (appears green in the image) to see how the initial wetting is influencing the re-wetting and adsorption behavior at the interface. In Figure 17 CLMS images of starch in water and oil are displayed. Particles have first been suspended in oil and then brought in contact to the water interface. One can see that the starch particles are re-wetted by the water phase and diffuse to the water sided interface. But oil is surrounding the particles and clusters are created which can be small or even larger agglomerates. It has to be mentioned that the images taken here are done in quiescent and no mechanical or thermal force is applied. Obviously starch particles can be rewetted by the water phase but an exact wetting angle could not be determined using this technique. Since wetting behavior is not clear defined, no statement is made whether the suspension is in the pendular or capillary state. Therefore for further discussion the amount of secondary fluid for starch suspensions is given via a secondary fluid saturation $S_{sec}$ which does not indicate a statement about wettability:

$$S_{sec} = \frac{V_{sec}}{V_{bulk} + V_{sec}} \quad (4.1)$$

The second system investigated here are cocoa particles suspended in vegetable oil with water as secondary fluid. The suspension is presumably in the capillary state since oil as the bulk fluid is the better wetting liquid as has been reported by Galet [39]. The three phase contact angle was measured to be $\theta = 148 \pm 4.5^\circ$ using the sessile drop method in the three phase system. To further analyze the contact angle and the re-wetting behavior of the particles, microscopic images from particles at the oil-water interface were taken. The results are shown in Figure 18.

![Figure 18: Cocoa particles at the oil/water interface. Particles have been first dispersed in oil (left) and in water (right) before creating the interface.](image)

In the images where particles initially were dispersed in oil, it can be clearly seen how the particles stick to the interface. It cannot be clearly distinguished if the particles prefer to be in the water or oil phase. Depending on particle nature we can see both preferences. If we take a closer look to the
pictures were the particles have first been dispersed in water, it looks like the cocoa is more hydrophilic and does not want to enter the oil phase. In both cases we can see that the oil/water interface is not a straight line, but a diffusive movement around the particles is happening. Oil is forming clusters with the particles in the water phase, as well as clusters of water covered particles are formed in the oil phase. We can conclude that a defined statement about the wettability of cocoa particles is not possible at this state. Cocoa particles are a natural product and are produced from the cocoa bean containing fatty hydrophobic parts as well as a hydrophilic fiber and protein content. This is causing the non defined contact angles. For the capillary suspensions investigated here particles were first dispersed in oil and water was added afterwards. With the microscopic images (Figure 18 left) it is shown that the particles diffuse to the interface and therefore can be seen as surface active. A clear statement if capillary suspensions made from cocoa particles in oil are in the capillary or pendular state cannot be given, but due to sessile drop images we assume the suspension to be in the capillary state for further discussions.

4.1.2 Network formation in suspensions from cocoa and starch particles

Suspensions from starch granules in oil are prepared via standard sample preparation method described in section 3.2.4. In Figure 19 suspensions with (right) and without (left) added water are shown. One can see a drastic change in texture attributed to the formation of a capillary when water is added; this is shown by the transition from a fluid-like behavior on the left to a gel-like, solid looking paste on the right. The network formation is schematically sketched below the photos of the suspensions.

![Figure 19: Suspension of \( \phi=0.33 \) starch granules in oil without added water \((S_{\text{sec}} = 0.0, \text{left})\) and with added water \((S_{\text{sec}} = 0.10, \text{right})\). Pictures below the starch samples schematically show the network formation.](image-url)

In Figure 20 a diluted suspension of starch granules in oil is shown. Water was added as secondary fluid. The image was taken with the fluorescence microscope from Zeiss and has been created by
overlaying the white light image with the fluorescence intensity image, for the latter the low intensity scattering has been removed and the higher intensity has been colored in yellow. The water is located between the particles, which indicates that a strong cohesion is created due to capillary forces.

Figure 20: Microscopic image of a capillary suspension made from starch granules in oil with water as secondary phase. Water was colored with a fluorescent dye (PromoFluor-488 Premium, carboxylic acid) which appears yellow in the picture.

The transition of a cocoa suspension from a weakly viscoelastic fluid to a gel upon addition of water is shown in Figure 21. These images demonstrate that capillary suspensions can be created in cocoa suspensions even when no sugar is present. Cocoa particles form networks and maintain a smooth texture but do not cause large agglomerates as it has been reported for sugar particle suspensions [13]. Stability against sedimentation of cocoa particles suspended in oil at a volume fraction of $\phi = 0.35$ is also pictured in Figure 21. The pure suspension shows clear phase separation after 5 days resting (Figure 21, right A), whereas the suspension with water added as secondary fluid ($S = 0.90$) shows no settlement at all (Figure 21, right B). This indicates that a sample spanning network is created, which is preventing the particles from sedimentation.

Figure 21: Suspension of $\phi = 0.35$ cocoa solids in oil suspension without added water ($S = 1.0$, left) and with added water ($S = 0.90$, middle). Right image: sedimentation behavior after a storage time of 5 days, (A) no water added, (B) $S = 0.90$.

Cocoa particles are known to exhibit a strong autofluorescence [87], [88] in all available wavelength excitations of optical microscopes. This makes it hard to detect capillary bridges since it cannot be distinguished between particle autofluorescence and the dyed secondary fluid. Fluorescence of the particles is due to phenolic substances in the cocoa that cannot be removed easily. An attempt has been made to suppress the auto-fluorescence by dying the cocoa particles with toluidine blue. This dye is docking on the phenolic groups in the cocoa particles resulting in a reduced fluorescent signal [89]. This is pictured in Figure 22. It can be clearly seen that on the left picture the particles show
fluorescence for both laser extinctions, the 488 nm laser signal is pictured in green, the 552 nm in red and overlaid fluorescence appears yellow. In the middle image the fluorescent signal is highly suppressed by the toluidine blue. Only a weak fluorescent signal occurs for the 488 nm laser and some yellow spots refer to an overlaid signal. In the left image water that been colored with a carboxylic acid (promofluor-488) is added to the sample via the regular suspension preparation method. Cocoa particles appear red or yellow, whereas the water phase appears pure green in the image. Apparently, the water phase is homogeneously distributed between the particles. However, a network structure cannot be visualized.

Figure 22: Confocal images of cocoa particles in oil with and without added water. The system was excited with both laser lines (488 nm and 552 nm). In the left image cocoa particles are used as provided, in the middle they have been dyed with toluidine blue and the right image shows a dyed particles suspension with water as secondary fluid (green). The microscope setup was kept constant.

4.1.3 Rheological behavior of cocoa and starch suspensions

Rotational shear measurements were performed for oil based systems made with cocoa and starch particles. In the following section the basic rheological parameters like yield stress and viscosity are discussed as a function of volume fraction and saturation. Also steady shear data is compared with oscillatory shear measurements performed on the same rotational rheometer. In Figure 23 A the yield stress of starch granules in oil is shown as a function of saturation varied between $S_{sec} = 0.0$ to $S_{sec} = 0.10$ for volume fractions of $\phi = 0.29$ and $\phi = 0.33$. Yield stress data are normalized to the respective value of the suspension where no water is added. The suspensions with water as secondary phase show a linear increase of yield stress in a semilogarithmic plot and the absolute value of the slope is $|\alpha| = 30.3 \pm 2.8$ for both concentrations. This results in an increase of $\sigma$ by three orders of magnitude when $S_{sec}$ is increased from 0.0 to 0.1. This confirms the reinforcement of a sample spanning network when water is added to the starch in oil suspension since the yield stress is characteristic of the network strength. Starch suspensions with higher saturation exhibit a strong, inhomogeneous agglomeration, so reliable measurements could not be performed at saturations $S_{sec} > 0.10$. 
Results and discussion

Figure 23: Normalized yield stress as a function of saturation of the preferentially wetting fluid for (A) corn starch and (B) cocoa particles.

In Figure 23B the yield stress for cocoa suspensions with different volume fractions of cocoa is plotted over a wide range of saturation S. Again, yield stress data are normalized to the corresponding value where no water is present (S = 1.00). The normalized yield stress for different volume fractions follows a single master curve. Similar behavior has been observed for PVC capillary suspensions as shown in Figure 24.

Figure 24: Yield stress of a PVC suspension (capillary state) and a suspension made from Hermatite particles (pendular state) in DINP/water mixtures [1].

The yield stress of the cocoa suspension increases until a saturation of 0.7 is reached and then drops until a value close to the initial stress is obtained. This latter decrease is presumably due to spherical agglomeration [10], [11]. In previous studies spherical agglomeration within capillary suspensions at elevated degrees of saturation was reported for oxide powders in oil [50]. The process of wet granulation with water as bridging fluid is used to agglomerate dry cocoa powder [90]. The same granulation can also occur in suspension at high secondary liquid fractions during mixing. When the saturation exceeds 0.5, phase inversion is visible: water acts as the continuous phase and the suspension begins to phase separate. Again, a linear increase of the reduced yield stress as a function
of saturation is found in semilogarithmic representation for \(0.9 < S < 1.0\) with a slope of \([\alpha] = 14.4 \pm 1.0\), which is half the value found for starch suspensions. This difference may be caused by several factors such as wetting angle, interfacial tension and, most relevant here, the fact that cocoa particles have a higher porosity. This may lead to an uptake of water into the particles, which is then not available for capillary bridging. Differences in wetting behavior and the resulting effect on the rheology of the admixture are discussed in the following sections.

Viscosity curves for starch (A) and cocoa (B) suspensions in oil at different saturations are shown in Figure 25. In both cases, the low shear viscosity increases with the addition of water by several orders of magnitude. At shear rates \(\dot{\gamma} > 500 \text{ s}^{-1}\) the curves approach each other, since capillary bridges are breaking and viscosity is then just a function of particle volume fraction. This strong degree of shear-thinning guarantees that capillary forces do not affect processing or application where high shear rates are involved [91]. For spherical hard sphere suspensions with particle loadings between \(\phi = 0.33\) and \(\phi = 0.35\) and \(\eta_b = 34 \text{ mPas}\) we expect a high shear viscosity of \(180 - 220 \text{ mPas}\) [40], [92]. The absolute values observed here for the cocoa and starch suspensions are about a factor of 2-3 times higher. This is partly due to the non-spherical shape of the particles but may also indicate residual agglomerates in the suspension. For the starch suspensions, the high shear rate viscosity perfectly matches irrespect of the saturation. This confirms that granule swelling does not play a significant role in these capillary suspensions and bridges can be destroyed completely under high shear conditions.

![Figure 25: Viscosity vs. shear rate for suspension of starch granules (\(\phi=33\%\), A) and cocoa particles (\(\phi=35\%\), B) suspended in oil, at different saturation levels.](image)

For the cocoa particles (Figure 25B), the viscosity curve with no added water \((S = 1.0)\) has a lower high shear viscosity value than samples were water was incorporated. This could be an evidence for water absorption by the particles [93]. Since the amount of water, that we add as the secondary liquid is small compared to the whole particle volume, we assume residual agglomerates to be responsible for the increased high shear viscosity and not only particle swelling. At intermediate shear rates, all viscosity curves for starch as well as cocoa suspensions exhibit a short plateau region where we
assume structural changes to occur. Flow-induced break-up or reorganization of agglomerates might
be involved there [21].

Comparison of rotational and oscillatory measurements

Oscillatory shear tests can be used to describe the viscous and elastic behavior of a suspension. In the
following, oscillatory amplitude sweeps and frequency sweeps are discussed. In Figure 26 a frequency
sweep, operated at $\sigma = 5$ Pa of a corn starch suspension ($\phi = 33\%$) with different amounts of water is
depicted. The complex shear modulus

$$|G^*| = G' + iG''$$

is plotted over the frequency. A clear transition from a fluid like behavior, for the sample without
added water ($S_{sec} = 0.00$), to a gel like and highly elastic behavior, for samples with added water, can
be seen. The more water is added to the suspension, the higher the complex modulus.

![Figure 26: Frequency sweep of corn starch suspensions ($\phi = 33\%$) with varied saturations.](image)

Measurements have been performed at $\sigma = 5$ Pa.

In the previous section yield stress is measured via a continuous shear test in stress controlled mode.
But the strength of the network of a capillary suspension can also be characterized by performing
oscillatory shear tests [1], [21], [86]. The advantage of oscillatory amplitude tests lies in a reduced
sensitivity for wall slip. This has been explained in more detail in chapter 3.2.1. For suspensions
correlations between yield stress values obtained from steady shear and oscillatory shear
measurements have been widely studied [94]–[96] and for different fluids various correlations can be
found. For capillary suspensions no general approach has been done yet. Two characteristic data
points can be calculated from oscillatory data which can be correlated to the yield stress: The
crossover point of $G'$ and $G''$ called $\sigma_{cross\,over}$ and the critical stress where $G'$ is reduced by 10% of its
value from linear viscoelastic region written as $\sigma_{crit}$.
In Figure 27 the yield stresses obtained from steady shear measurements are plotted over the critical shear stress from oscillatory measurements (A) and over the crossover point (B) for starch and cocoa suspensions, which have been prepared with varied secondary fluids (aqueous fructose and glycerol mixtures) and secondary fluid contents. For the suspensions shown here the yield stress from steady shear data exhibit a linear relationship with the critical shear stress calculated from oscillatory data, since the slope is measured to be $|\alpha|=1.07$. Critical stress data $\sigma_{\text{crit}}$ can therefore be calculated from $\sigma_y$ by applying the equation $\sigma_{\text{crit}} = 0.14 \sigma_y^{1.07}$. In contrast for the cross over point $\sigma_{\text{cross over}}$, the slopes is $|\alpha|=1.2$ indicating a weak non-linear relationship, which can be describes as $\sigma_{\text{cross over}} = 0.21 \sigma_y^{1.2}$.

These results are not in line with data found in literature for silica dispersions [97], where it is said that the cross over tends to overestimate the yield stress of suspensions.

4.1.4 Influence of contact angle and viscosity ratio

For a basic characterization of the capillary suspensions of particles dispersed in oil, measurements were done using starch particles dispersed in sunflower oil with water as secondary fluid. To investigate the influence of wetting angle, interfacial tension and viscosity ratio, the particle surface was treated and various aqueous glycerol and poly ethylene oxide (PEO) mixtures have been used as secondary fluid. All yield stress measurements have been carried out using the vane geometry to avoid wall slip. The suspensions were all prepared with a volume fraction of starch particles of $\phi = 0.33$.

Secondary phase viscosity

If we have a look at the force of capillary bridges acting between two particles, a direct influence of secondary phase viscosity on capillary strength and therefore flow behavior of capillary suspensions is not expected [44]. But the preparation step of capillary suspensions may be seen as an emulsification of the secondary fluid in the bulk fluid. For emulsion preparation, it is known that laminar droplet breakup is a function of the ratio of dispersed phase and continuous phase viscosity $\eta = \eta_d/\eta_c$ [61].
Therefore, we have investigated the influence of the viscosity ratio on secondary phase distribution and flow behavior. To modify the secondary phase viscosity, water was thickened with polyethylene oxide (PEO) varying in concentration from 0.1 wt.% to 2.5 wt.%. This increases the viscosity of the secondary phase by three orders of magnitude. But within experimental uncertainty all solutions show the same value of interfacial tension $\gamma = 22 \pm 2$ mN/m.

Figure 28 shows corresponding yield stress for a cocoa and a starch suspension. The yield stress does not change significantly with increasing secondary phase viscosity, i.e. the droplet breakup conditions do not have an influence on network strength. The capillary bridging even occurs at ratios $\eta_d/\eta_c >> 1$ where shear induced droplet breakup should not appear assuming laminar shear flow and quasi-static conditions [61]. This suggests that the bridging phenomenon is not a just function of droplet breakup and emulsification of the secondary phase. Instead we propose that the network state is energetically more favorable and the network formation also depends on the diffusion of the secondary fluid. This energetic favorability was previously demonstrated for the formation of small particle number clusters induced by a non-wetting secondary fluid [21].

![Graph showing yield stress of corn starch ($\phi = 0.33$) and cocoa ($\phi = 0.35$) particles suspended in oil with aqueous PEO solutions of different viscosity as secondary phase versus the viscosity ratio of water to oil $\eta_d/\eta_c$.](image)

**Figure 28:** Yield stress of corn starch ($\phi = 0.33$) and cocoa ($\phi = 0.35$) particles suspended in oil with aqueous PEO solutions of different viscosity as secondary phase versus the viscosity ratio of water to oil $\eta_d/\eta_c$.

Assuming droplet breakup occurs at a Capillary Number $Ca \approx 1$ a minimal attainable droplet size due to mixing can be estimated assuming $\dot{\gamma} = 500 \text{ s}^{-1}$ and $\eta = 0.5 \text{ Pas}$, which refers to the suspension where no water was added. Calculation results in a minimum droplet radius of $r \approx 96 \mu m$, which is much larger than the typical size ($<< 5 \mu m$) of a capillary bridge for the particle sizes used here. This is further evidence for a diffusion driven, energetically favored network formation.

**Contact angle**

Changing the contact angle of the two fluids with the particle without influencing the interfacial tension can be done by treating the particle surface. This can be achieved easily for starch particles.
Their surface can be covered with octenyl succinic anhydride (OSA), thus making the particle more hydrophobic [85]. Accordingly, the wetting angle in the three phase system starch/vegetable oil/water should increase. Nevertheless, the three phase contact angles measured via sessile drop method for all treatment levels investigated here were found to be $\theta = 130 \pm 7^\circ$ and no difference was detected with our setup. On the other hand, an increase in OSA concentration results in a reduction in the yield stress as shown in Figure 29 for two saturation levels.

It seems that subtle differences in surface properties not resolvable with conventional analytical methods still have a remarkable impact on the rheology of these capillary suspensions. At constant saturation a linear decrease of the yield value is detected. This demonstrates that particle surface treatment is an option to tune the flow behavior of capillary suspension.

Figure 30 shows the yield stress of a corn starch in vegetable oil suspension ($\phi = 0.33$) with water or glycerol/water mixtures added as secondary liquid at various degrees of saturation $S_{\text{sec}}$. At given $S_{\text{sec}}$ the yield stress increases with increasing glycerol content. Samples with pure glycerol as the secondary liquid show yield stresses up to one order of magnitude higher than samples made with pure water. The yield stress at $S_{\text{sec}} = 0.06$, for example, is $\sigma_y = 39 \pm 4$ Pa with water as the secondary phase, whereas with glycerol the yield stress is measured to be $\sigma_y = 405 \pm 22$ Pa at the same saturation. This behavior fits the capillary bridging theory, whether the suspension is in the capillary or pendular state. Assuming capillary state, the Theory of Binks (section 2.2) can be applied. Particles that are placed at an interface of two fluids are held with the energy $E$ given by \( E = \frac{1}{2} \gamma AB \) \( \theta \). The maximum energy $E_{\text{max}}$ is therefore given at a contact angle of 90° and approaches zero for contact angles close to 180°. The energy gain coming with the formation of clusters varies with contact angle and degree of saturation [21]. In the aforementioned paper, it is described, that strongest clusters i.e. cluster structures with the lowest energy are formed at contact angles close to 90° and high saturations. Since glycerol and
sunflower oil form a contact angle with starch granules closer to 90° than for water/sunflower oil (Table 6), the force for cluster breakup is higher and the corresponding clusters are more stable. This results in a more stable network and finally in a higher yield stress if glycerol is used as secondary fluid. Even when assuming that the results of sessile drop measurements are not true equilibrium values, a trend in wetting can be seen: with more glycerol in the mixture better wettability of the starch by the secondary fluid can be detected. In pendular state, when applying equation (2.4) it can be stated that for lower contact angles the capillary force between two particles becomes stronger. In conclusion, whether the system is in pendular or in capillary state, lower contact angles result in stronger particle bridging and an increased yield stress of the suspension.

Table 6: Contact angles measured via sessile drop method between starch and various glycerol-water mixtures in the presence of sunflower oil as well as viscosity of these mixtures at 20°C.

<table>
<thead>
<tr>
<th></th>
<th>viscosity in mPa s</th>
<th>contact angle in °</th>
</tr>
</thead>
<tbody>
<tr>
<td>water</td>
<td>1</td>
<td>126 ± 13.0</td>
</tr>
<tr>
<td>25% glycerol</td>
<td>2.5</td>
<td>104 ± 17.1</td>
</tr>
<tr>
<td>50% glycerol</td>
<td>8.5</td>
<td>101 ± 2.7</td>
</tr>
<tr>
<td>75% glycerol</td>
<td>53.3</td>
<td>93 ± 7.9</td>
</tr>
<tr>
<td>100% glycerol</td>
<td>1100</td>
<td>92 ± 9.0</td>
</tr>
</tbody>
</table>

Uptake of secondary fluid by the starch granules could also be a reason for the variation of the yield stress with glycerol/water mixing ratio at constant $S$. Starch granules absorb water which is incorporated into their amorphous parts [67]. Water that is trapped within the granules is not available to act as a bridging fluid. The strength of the network should decrease with increasing fraction of water in the secondary liquid since the effective saturation $S_{eff}$ is lower than the nominal saturation $S_{sec}$ calculated from the amount of added fluid. In contrast, glycerol is not absorbed by the particles due to its higher molecular weight and therefore more secondary fluid is available for capillary bridging. For the measured difference in yield stress due to water incorporation into the granules, about 0.8g of water per 10g of starch must be trapped in the granules. This is a reasonable number [67], so water uptake as well as variation of contact angle might contribute to the variation of yield stress in capillary starch suspension. For non absorbing glass particles atomic force measurements performed by Negreiros [22] also have revealed that glycerol forms stronger capillary bridges in oil than water. They claim viscous forces to be the reason for this difference, but an influence on viscosity cannot be confirmed by rheological measurements in this study (see Figure 29).

Swelling of starch granules through the uptake of water increases the granule volume by about 12% for the maximum water uptake calculated above [67]. This small difference in volume fraction does
not account for the overall increase in yield stress in capillary suspensions and it would imply that the difference in $\sigma_y$ between suspensions prepared with glycerol or water should monotonically increase with $S_{sec}$.

![Diagram showing yield stress $\sigma_y$ vs. saturation $S_{sec}$ for various glycerol/water mixtures.](image)

**Figure 30:** Yield stress of a starch suspension in oil ($\phi = 0.33$) vs. saturation for various glycerol/water mixtures as secondary fluid.

### 4.1.5 Mixtures of hydrophilic and hydrophobic particles

In food products suspensions do rarely consist of just one particle type, but are mixtures of particles of different shape and surface properties (roughness, wetting). In this section a closer look is taken on mixtures of hydrophilic and hydrophobic particles and how they behave rheologically. Pure Suspensions of hydrophilic or hydrophobic particles already can show different flow behaviour [99]. For mixtures of hydrophilic and hydrophobic particles in an water/oil emulsion it has been found that adding hydrophilic particles to a Pickering stabilized emulsion system with hydrophobic particles results in an phase transition of the emulsion when more than twice the mass of hydrophilic particles compared to the hydrophobic once are added to the system [100]. The aim of this experiment is to find out if we can create pendular state suspensions as well as capillary state suspension by changing particle surface properties while keeping the bulk and secondary phase constant.

Two particle combinations have been analyzed in order to identify a general behavior. First, mixtures of hydrophilic Socal particles (Socal 31, $x_{50} = 70$ nm) and hydrophobic Socal particles (Socal U1S1, $x_{50} = 80$ nm) in sunflower oil with a volume fraction of $\phi = 10\%$ were investigated. Water was added as secondary fluid at two saturation levels. Hydrophobized Socal U1S1 suspensions ($\phi = 10\%$) without secondary fluid exhibit a yield stress of $\sigma_y = 8.5 \pm 0.5$ Pa. Yield stress is linearly decreasing with increasing volume fraction of hydrophilic Socal until a yield value of $\sigma_y = 1.4 \pm 0.1$ Pa is reached for pure Socal 31 suspensions.
Results and discussion

Figure 31: Normalized yield stress over hydrophilic fraction for Socal particle mixtures in oil with a volume fraction of $\phi = 10\%$ and saturations of $S_{sec} = 0.03$ and $S_{sec} = 0.05$.

The results of the measurements for saturations of $S_{sec} = 0.03$ and $S_{sec} = 0.05$ are shown in Figure 31. For hydrophilic volume fractions $H = V_{\text{hydrophilic particles}} / V_{\text{hydrophobic particles}} < 0.6$ the yield stress is increasing with increasing $H$ for both saturations. As has been reported before, hydrophobic Socal particles in silicon oil can form capillary suspensions with the addition of small amounts of water [1]. At the given saturations we have already exceeded the plateau region [1], so no different behavior is expected between the two saturations. Yield stress of the suspensions with added water is constant and increase in yield stress is due to decreasing zero value $\sigma_{y, S=0}$. For higher fractions of hydrophilic particles ($H = 0.7$) the normalized yield stress first drops and then increases again.

Figure 32: Confocal images of Socal particle mixtures in oil with water ($S_{sec} = 0.05$) as secondary fluid. From left to right: $H = 0$; $H = 0.3$; $H = 0.7$; $H = 1$.

The structural changes occurring in the analyzed system here can be visualized using confocal microscopy. Hydrophilic particles have been dyed using Rhodamine B, water was colored with Promofluor-488, the oil phase and the hydrophobic particles were kept uncolored. Images of different hydrophilic fractions of Socal at $S_{sec} = 0.05$ are pictured in Figure 32. One can see that for $H = 0$ the water (appears green in the images) is distributed through small droplets in the sample. For $H = 1$, hardly any water can be detected. This is due to the large amount of dyed particles covering the water signal. With a hydrophilic fraction of $H = 0.3$, the distribution of the water is fairly homogeneous and
appears more floc like. At $H = 0.7$ a clear change is visible, the water appears as large droplets, covered by particles. These larger droplets reduce the water that is available for particle capillary bridging and therefore the yield stress is decreased as shown in Figure 31. From the confocal images it cannot be clear said if the structure is mainly build by hydrophilic or hydrophobic particles.

As a second system starch particles have been hydrophobized and mixtures from natural and treated particles were investigated. The hydrophobization of the particles is described in Section 3.2.5. In contrast to the Socal suspension this system shows a monotonic variation of $\sigma_y$. The yield stress decreases with increasing amount of hydrophobized particles and at $H = 0$ $\sigma_y$ equals the value $\sigma_{y,S=0}$ of the pure suspension. This is demonstrated in Figure 33. The yield stress first drops slowly, since there are still hydrophilic particles in the system which can form a sample spanning network. Then $\sigma_y$ decreases due to a lower “capillary active” particle volume fraction, indicating a weaker network [86]. If no natural starch particles are left in the system no sample spanning network can be created and the yield stress drops.

![Graph](image)

**Figure 33:** Yield stress of a starch in oil suspension ($\phi = 35\%$) with a water saturation of $S_{sec} = 0.07$ plotted over hydrophilic fraction $H$ of starch particles.

### 4.1.6 Particle preconditioning

From wet granular material science it is well known that particles form capillary bridging networks not only if water is added directly to the granular substance which is a common approach in wet granulation processes [101], [102] but also if the particles themselves contain water that is adsorbed on the particle surface as a water layer due to capillary condensation. This is intensively discussed in reference [103]. Capillary bridges form if the wet particles come into contact. The system is then not in an energetically favorable state since a sharp edge is created between the water layers [104]. The wetting liquid induces the formation of a bridge which is energetically favorable and are held together due to the negative Laplace pressure across the curved surface [104].
Capillary forces can also form in ternary mixtures where one fluid is adsorbed to the particle surface and the bridges are formed upon contact of two particles [105]. It has been reported that capillary bridges are formed between particles dispersed in partially miscible liquids due to an absorbed fluid layer when stepping into the two phase region [106], [107]. This aggregation can be formed spontaneously [4], [108] or can be induced by shear [109].

In the following section experiments are discussed revealing how initially wet particles behave when dispersed into a dry, unpolar bulk phase. We compare the rheological behavior between two preparation methods of the suspensions. These preparation methods are schematically depicted in Figure 34. On the one hand, the particles were preconditioned with a specific amount of water as has been described in section 3.2.5 and these wet particles were directly dispersed in sunflower oil. On the other hand the same amount of water that has been soaked by the particles in the first preparation method is added drop wise under shear after dispersing the dry particles in oil. The setup for particle preconditioning as well as amount of absorbed water by different particles are described in section 3.2.5.

First measurements were performed using three different model systems: the PVC particle Vinnolit C 12/62 V and two different calcium carbonate particles Socal U1S1 (hydrophobic) and Socal 31 (hydrophilic). In Figure 35 the data obtained from Cavalier [110] is compared to own data from conditioned Socal 31 particles. The water content related to the bulk phase from Cavalier’s data has been calculated from given water activity (a\textsubscript{w}-value) using Herny’s law, which leads to m\textsubscript{w} = a\textsubscript{w} / 606 [110]. The plot shows that the yield stresses show a similar dependence on m\textsubscript{w} but the Cavalier data exhibit much higher absolute values. This can be attributed to the different hydrophobic bulk phase substances and a slightly higher volume fraction of particles in the Cavalier study.
Results and discussion

Figure 35: Yield stress of a suspension from Socal 31 in dioctylphthalate ($\phi=10.8\%$) with different water content in the oil phase [110] compared to a suspension from preconditioned Socal 31 particles in sunflower oil ($\phi=10\%$) plotted over the water content (calculated to the oil phase).

In Figure 36 the yield stress data from hydrophobic and hydrophilic Socal suspensions are plotted over saturation $S$. One can see that for hydrophilic calcium carbonate particles (Socal 31) the yield stress is higher for direct addition of water. So it seems that a small amount of water is trapped in the conditioned particles that cannot contribute to network formation. Also due to the very high sensitivity of the systems to small volumes of water, humidity while sample preparation can also have a high impact even though using dried particles. For the hydrophobic Socal (Socal U1S1) particles the measured data points show a wider deviation than for the hydrophilic ones. The experiments reveal that suspensions prepared with conditioned particles tend to have a higher yield value than suspensions made from dry particles with subsequent water addition. First, it seems unlikely that water absorbs to hydrophobic particles during the preconditioning step, but as has been reported before even fully hydrophobic particles can adsorb water due to defect sites at the surface [111]. It has also been shown that the greater the hydrogen bonding capacity and the smaller the size, the higher the water adsorption on particle surfaces [112]–[114]. So for Socal U1S1 particles water is adsorbed at the surface and water incorporated into the suspension this way is available to form a capillary suspension. It has been reported earlier that hydrophobically modified calcium carbonate particles in oil form a capillary state suspension when water is added [1], [3]. So we propose that the water that is bond around the particle surface is displaced by the oil and thus available to form small Socal particle cluster, further aggregating into a sample spanning capillary network. The deviation between the suspensions from conditioned particles and the direct addition of water can be explained as for Socal 31 particles by humidity or irreversible bonding of water into the particles.
Figure 36: Comparison of yield values of suspensions made from pre-conditioned and dry Socal particles (A: Socal 31, $\phi = 10\%$; B: Socal U1S1, $\phi = 14\%$) plotted over the saturation $S$.

The different suspension preparation methods described above have also been used for Vinnolit particles. The results are depicted in Figure 37. Yield stress of suspensions prepared with the two preparation methods nicely coincide within measurement error. The yield stress does not increase drastically upon the addition of water, but an increase of the factor of three can be measured and considered to be due to a weak network formation. The results discussed above all clearly show that capillary suspensions can be formed using both preparation methods for pendular and capillary state as long as particles exhibit a slight hygroscopic nature. It also suggests that theory from wet granular material discussed in the section above can be applied to the three-phase systems considered here.

Figure 37: Comparison of yield values of suspensions made from pre-conditioned and dry Vinnolit particles ($\phi=35\%$) plotted over the saturation $S$.

Food particles like cocoa and starch tend to absorb water as well as oil. For this it is interesting how the two preparation methods compare. In Figure 38A the yield stress data from cocoa particle suspensions prepared with the two methods are shown. For saturation values $S < 0.90$ direct water addition reveals higher values for the yield stress than particle preconditioning. Is the particle soaked with water before coming in contact with the oil phase, parts of the soaked water might be irreversibly
incorporated in the inhomogeneous particles especially at high water content, resulting in a reduced yield value compared to suspensions prepared via standard procedure. The bridge formation will be induced by the energetically driven curvature formation described for wet granular material in the section above. If the particles however are soaked with oil and the added water needs to remove the oil from the surface afterwards in order to build a capillary bridge, less water can be soaked by the particles and more fluid is available for the network formation.

In contrast, for starch suspensions yield stress values from both preparation methods match perfectly as illustrated in Figure 38B. It has been investigated before that starch granules do adsorb water in a single layer and, with higher relative pressure, multiple layers. The uptake can exceed a few molecular layers if the granule swells and thus incorporates water molecules [67], [81], [115]. Water added to the suspension is always utilized in the same way for both methods, whether all water is used for the formation of bridges or if parts of the secondary fluid are still incorporated into the particles and not available for network formation. The difference in behavior of starch and cocoa suspensions may be due to the fact that cocoa particles exhibit a more porous and inhomogeneous structure, as well as a wider particle size distribution with highly anisotropic form factors (see section 3.1.1) compared to starch particles.

The findings for model starch and cocoa suspensions clearly demonstrate that water which is ad- or absorbed by the particles is not trapped in the particle structure but obviously can diffuse from an adsorption layer into capillary bridges, no additional mechanical energy input is required to create capillary suspensions. The phenomenon suggests a new preparation method for all hygroscopic particles that form capillary suspensions when oil is the bulk phase. So the described method may be a valuable route for processing of new food products, but also points at a complication in existing processes where the product consistency may dramatically depend on the storage conditions for the solid particles.

![Figure 38](image-url)

**Figure 38**: Comparison of yield values of suspensions made from preconditioned and dry starch (A, $\phi=33\%$) and cocoa (B, $\phi=35\%$) particles plotted over the saturation $S_{sec}$ and $S$. 
4.1.7 Thermal stability

For temperature stability measurements samples were stored at different temperatures (-18°C, 4°C, 20°C and 40°C) in a closed vessel for 19h and yield stress was measured after letting the samples cool/warm up to 20°C. At 40°C samples were also stored in an open vessel to see the influence of evaporation at elevated temperatures. Yield stress values after 19h of storage are plotted in Figure 39. Frozen sample as well as the sample stored in an open vessel at 40°C show a distinct lower yield value than samples stored at 4°C, 20°C and 40°C, closed.

Since $\sigma_y$ for the sample stored at 40°C in the closed vessel is significantly higher than that of the sample stored in the open vessel, we can conclude that water evaporates from the samples at 40°C thus lowering the saturation and accordingly the yield value. This also indicates that water does diffuse through the oil phase, but influences the stability only by evaporation since the closed sample does only show a minor yield stress reduction compared to 4°C and no change compared to 20°C. Freezing the capillary suspension clearly destroys parts of the capillary network. Ice crystals formed during this slow freezing process may detach from the particle surface during re-heating to 20°C prior to the yield stress measurement. It is reported that Pickering stabilized emulsions exhibit a good freeze-thaw stability [116], but fat and ice crystals can also induce a loss in stability during freeze-thaw cycles due to collision-mediated destabilization [117], [118]. This might explain the destabilized network in the frozen capillary suspension. But ice crystals formed during the slow freezing process may just detach from the particle surface and phase separate thus not being available for capillary bridging after re-heating.

![Figure 39: Yield stress of cocoa suspensions ($\phi = 30\%$, $S = 0.9$) stored at different temperatures for 19h.](image-url)
4.1.8 Resistance against melting

If chocolate is exposed to heat like on a warm sunny day, it will melt and lose its favorable snap and handling behavior. One attempt to increase the stability against melting is to incorporate water into the chocolate as has been reviewed in chapter 1. The stability of the chocolate product is said to be due to capillary bridges formed between the sugar crystals. Here the melting behavior of a model chocolate that consists only of cocoa particles dispersed in cocoa butter without the addition of sugar has been investigated. Different amounts of water and other aqueous solutions like glycerol and fructose mixtures have been added as secondary phase. The melting behavior of the chocolate was measured via optical and rheological measurements as described in section 3.2.7.

4.1.8.1 Meltability of model chocolate with water as secondary fluid

In Figure 40 model chocolate blocks with and without added water are pictured. Here an example where a saturation of $S = 0.90$ was incorporated into the sample is shown. Blocks prepared with water as the secondary fluid show a strong increase in stability against melting compared to blocks where no water is added. One can clearly see that the block formed without water is melted after 40 minutes whereas the sample with water remains its shape. After what we have shown in the previous chapters we claim that capillary bridges are formed between the cocoa particles, which results in a particle network that prevents the melted cocoa butter from spreading. The heights of the samples were measured over time to compare value for different mixtures, quantitatively.

**Figure 40: model chocolate with a volume fraction of cocoa particles of 35% stored at 50°C with (left) and without (right) added water.**

Normalized height curve of a sample made from $\phi = 30\%$ of cocoa particles in cocoa butter is shown in the left image of Figure 41 for various water saturations. For samples with saturation $S_{sec} \geq 0.90$ a fast decrease in sample height can be detected, whereas samples with $S_{sec} \leq 0.90$ show an almost constant shape. Samples were water was added clearly melt slower than the sample were no water was added. Increased resistance against melting is a function of particle network stability. For higher secondary fluid volumes the network strength is enhanced. This can be compared to the yield stress measurements performed with the cocoa in sunflower oil suspensions (see Figure 23): Increasing the amount of secondary fluid leads to a more stable network.
In the right image of Figure 41 the apparent viscoelastic moduli for the same sample as shown in the left image is displayed. $G'$ and $G''$ is plotted over temperature, which was linearly increased from 32°C to 40°C within 46 minutes. The sample with a saturation of $S = 0.85$ is in the linear viscoelastic region during the whole measurement. The storage modulus $G'$ is dominating over the loss modulus $G''$, i.e. the chocolate is not in a fluid like state, but behaves like a solid throughout the whole temperature range. For a saturation of $S = 0.95$ a stronger decrease of $G'$ and $G''$ can be detected over temperature, but still elastic behavior is dominating. Samples with no water added show a crossover of $G'$ and $G''$ at around 36°C. Here the sample is melted and the suspension is in a liquid like state. This temperature is slightly higher than reported for commercial products, where the melting temperature is said to be around 30°C [66]. This might be due to the different processing like tempering and conching of the commercially produced chocolate [119], [120] and the sample shown here.

Figure 41: Sample height $h$ normalized to the height at the beginning of the test $h_0$ plotted over storage time at 50°C for various saturations at a volume fraction of cocoa particles of 30% (A). $G'$ (closed symbol) and $G''$ (open symbols) is plotted over temperature $T$ for the same samples as seen in (A) at $f=1$Hz and $\tau=0.1$ Pa (B).

Besides water content, the volume fraction of the particles in the suspension does also play a role in melting behavior. In Figure 42 this is pictured for a suspension made without (A) and with water as secondary fluid (B) with cocoa particle volume fractions of 25%, 30% and 35%. Figure (A) shows that the volume fraction has no influence on the melting behavior when water is added. Only a very instable van-der-Waals network exists between the cocoa particles. When the fat phase is melted, the network collapses independent of particle volume fraction. If water is added to the suspension, a different behavior is detected. The higher the volume fraction, the higher is the plateau that remains after the block is melted. More particles are available to provide a stronger network. This network resists shrinkage which results in a less collapsed looking chocolate block. Therefore, at constant saturation and wetting behavior the created network is more heat stable. With higher volume fractions the melting is also slower than for samples with lower volume fraction. Since without added water the volume fraction of cocoa particles does not play a role in melting behavior, this is attributed to the
flow of melted cocoa butter through the porous structure provided by the cocoa particle network. In previous work it has been shown that with higher volume fraction and constant saturation the porosity of the capillary network is slightly decreasing [50]. But phase transport though porous media cannot be described by the porosity alone, but is generally expressed by the specific permeability. Permeability of a system is a function of many variables like size and shape of pores and particles, as well as ratio of open and closed pores. Many different models have been published to describe the theory of fluid flow through porous media, a summary can be found in reference [121]. For the systems shown here no further statement can be made besides that permeability decreases with increasing volume fraction.

Figure 42: Comparison of melting behavior of suspensions with cocoa particle volume fractions of 25%, 30% and 35% for saturations $S = 1.0$ (A) and $S = 0.90$ (B).

Model system

As a non food model system calcium carbonate particles have been dispersed in melted paraffin wax at a volume fraction of $\phi = 8\%$ and cooled over night at 20°C in a round form as described in section 3.2.7. In one sample water was added to a saturation of $S = 0.05$ (we assume pendular state) and one was left without further treatment. In Figure 43 the two blocks are shown before and after exposure to 50°C for 110 minutes.

Figure 43: Calcium carbonate in paraffin wax ($\phi=8\%$) with (left) and without (right) added water, before (upper image) and after (lower image) exposure to 50°C for 110 min.
The model system behaves similar to the cocoa suspensions: The sample where water has been added melts slower than the sample without secondary fluid. It also retains its shape, whereas samples without water collapse. The melting characteristics of these samples are depicted in Figure 44 confirming results obtained for cocoa particles suspended in cocoa butter. The increased resistance against heat with addition of secondary fluid to a particle suspension seems to be a universal phenomenon that may be used for various industrial applications.

**Figure 44:** Height of the melting samples at 50°C shown in Figure 43 plotted over time.

4.1.8.2 Influence of varied secondary phase

The melting behavior of the model chocolate is strongly influenced by the kind of secondary fluid added to the particle suspension. The type of secondary fluid has an impact on wetting behavior between the particles and the two fluids. This wetting behavior is mainly influencing the network strength [3]. Viscosity of the secondary fluid is contributing to the droplet size distribution, since viscosity ratio is controlling mixing conditions in the two fluid suspension [61]. Glycerol and aqueous fructose solution (75 wt.%) have been used as secondary fluid to investigate their impact on melting behavior. We have shown before, that viscosity ratio in the here examined range does not have an impact on yield value of the resulting capillary suspension for cocoa in sunflower oil suspensions (see Figure 30). This leads to the assumption that in the samples shown here this is also a valid statement.

Figure 45 shows the melting behavior of the chocolate with the two different secondary fluids. In the left figure (A) data are shown for suspensions where glycerol with saturations of $S = 0.05$, $S = 0.10$ and $S = 0.15$ was used as secondary fluid. It can be seen that with increasing saturation melting is delayed and a higher plateau value is reached. In contrast to water (see Figure 41), all saturations shown here are inducing a melding resistance, whereas with water stability is only reached for saturations $S > 0.10$. The right figure (B) shows the melting behavior when an aqueous fructose solution, prepared with 75wt% fructose, was added as secondary fluid. Melting behavior varies strongly compared to water or glycerol. The sample does not show increased heat stability compared
to the pure model chocolate where no secondary fluid was added. The difference in melting behavior indicates a stronger particle network for glycerol stabilized suspensions compared to suspensions stabilized with aqueous fructose solution.

Figure 45: Melting behavior of the model chocolate with glycerol (A) and 75wt% fructose (B) as secondary fluids, with varied saturation.

To confirm that changes in particle network stability are due to different wetting behavior of the two fluids, the contact angle the fluids form with the model chocolate block is investigated. The contact angle measurements for the heat stability experiments have been performed similar to the method described in section 3.2.3, but with a simpler setup. Model chocolate blocks were prepared as written above and the secondary fluid droplet was placed on the surface without further treatment. This measurement was done in air, since tempering and covering with cocoa butter was not possible with the available instrument. We assume that results will reflect similar trends as direct measurements in the three phase system. In Figure 46 the contact angles of glycerol (left) and 75% fructose solution (right) on the model chocolate block with $\phi = 30\%$ are shown. Glycerol forms a contact angle of $\Theta_{\text{glycerol}} = 94 \pm 4.5^\circ$ and fructose of $\Theta_{\text{fructose}} = 110^\circ \pm 1^\circ$. The contact angle of glycerol is closer to 90° which indicates a stronger attachment of the particles at a surface. [47], [122]. The difference between the two angles might seem not to be very significant but applying it to Binks’ theory for particles attached at a surface (see eq. (2.6)) detachment energy for every particle is increased by a factor of two for glycerol compared to the fructose solution.

Figure 46: Contact angle of glycerol (left) and 75% fructose solution (right) in model chocolate plate.
Since particle network stability is the decisive factor for the maintenance of the block shape, melting behavior of the model chocolate can also be correlated with the rheological behavior. To compare rheological parameters like yield stress, capillary suspensions have been produced with sunflower oil instead of cocoa butter. This makes the measurements easier and can be directly compared to the cocoa butter suspensions. In Figure 47 the yield stress of cocoa suspensions in sunflower oil at a volume fraction of $\phi = 30\%$ with glycerol, water and 75% fructose is shown.

![Graph showing yield stress vs. saturation range](image)

**Figure 47: Yield value of a cocoa in sunflower oil suspension ($\phi=30\%$) with water, glycerol and 75% fructose solution added as secondary fluid.**

The trend is obvious; the lower yield value for the fructose solution throughout the measured saturation range results in a weaker resistance against heat, whereas glycerol samples with higher yield values throughout the investigated S-range exhibit higher heat stability. For water as the secondary fluid, $\sigma_y$ is lower than for the glycerol sample at high S-values close to one, which results in a lower heat stability. For $S = 0.80$ the yield value of the suspension with water is higher than for the glycerol sample and this again corresponds to a stronger resistance against melting. So we can conclude that the yield stress can be directly correlated to the melting behavior of the suspensions. The yield stress reflects the strength of the capillary bridges between the particles. The more stable the capillary bridge, the more cocoa butter can be melted before gravity is forcing it to break. The more strong bridges and clusters are formed in the suspension the more melted butter can be hold within the pores of the network before the structure breaks down. Similar observations have been made for suspensions used in ceramic injection molding. Ceramic green parts with higher yield stress are more stable during debinding [123].
4.2 Water continuous suspensions

Cocoa particles have been used to prepare water continuous suspensions. First, the effect of temperature on the flow behavior of cocoa in water suspensions ($\phi = 15$ vol%) will be discussed, including the influence of cocoa butter on this effect.

Secondly, various secondary fluids have been added to the cocoa suspensions to identify if capillary suspension can be build with these water continuous suspensions. Influences of secondary phase chain length, interfacial tension and solubility on flow behavior have been investigated. Also building mechanisms for particle networks in these cocoa suspensions will be discussed.

4.2.1 Effect of temperature on flow behavior of pure aqueous cocoa suspensions

Suspensions of $\phi = 15$ vol% cocoa particles in water were stored at three different temperatures $T_s$ for 24h in sealed containers. After that, the viscosity function was immediately measured at a temperature $T_m = 20^\circ$C. Samples were placed into the rheometer and rested for 1 minute to assure constant temperature. Corresponding viscosity functions are plotted in Figure 48. Storage at 30°C results in a three decades higher low shear viscosity than storage at 20°C, whereas the suspension stored at 40°C exhibits a viscosity level even lower than that of the sample stored at 20°C. The high shear data for 20°C and 30°C samples was taken using a capillary rheometer as described in [124]. At shear rates $\dot{\gamma} > 10000$ s$^{-1}$, the viscosity data of samples stored at different temperatures superimpose.

Figure 48: Viscosity function of cocoa samples ($\phi$=15%) stored at 20°C, 30°C and 40°C for 24 h. Measurements plotted with an open symbol were completed using a capillary rheometer. Stirred sample was prepared by mixing the sample during storage using a magnetic stirrer.

Obviously, storing water continuous suspensions of cocoa powder at different temperatures has an enormous effect on the flow behavior of these suspensions. One might assume that the viscosity increase results from a swelling of the particles, however, the strong increase observed here cannot be
due to swelling and must be due to the formation of a particle network. This is confirmed by the convergence of the high shear viscosity data. Inter-particle bonds are broken and viscosity is just a function of particle volume fraction under the influence of high shear rates. Particle swelling would result in an increased effective solid phase volume fraction and correspondingly in an increase of high shear viscosity. Network formation can be deduced from comparing sedimentation behavior of samples stored at different temperatures. Those stored at 20°C and 40°C show sedimentation as a supernatant can be detected after 24 h storage in quiescent. Samples stored at 30°C do not show sedimentation behavior on that time scale. In all cases sedimentation is so slow that it does not affect rheological measurements on a time scale of several minutes. Bridging between the particles at elevated temperatures can be induced by molten or softened cocoa butter on the particle surfaces. Crystal forms I-V of the cocoa butter start to melt at 30°C [31], [125], but the cocoa butter is not in a completely liquid form. To confirm this, cocoa butter flakes were placed in water and stored for 24 h at 20°C, 30°C and 40°C. At 20°C and 30°C the cocoa butter flakes maintained their shape, whereas the cocoa butter stored at 40°C became liquid and completely phase separated from the water. The melting behavior is also documented in Figure 49, where the frequency dependent storage and loss moduli $G'$ and $G''$ of pure cocoa butter are plotted for different temperatures. At temperatures $T < 35°C$, the cocoa butter behaves like a predominantly elastic solid where $G'$ dominates over $G''$ in the whole frequency range. At 40°C, the cocoa butter is in a fluid-like weakly viscoelastic state with $G'' > G'$ at low frequencies.

Two different mechanisms that can account for the temperature sensitive behavior of the cocoa suspensions can be proposed. One hypothesis is for a sintering process to occur. Sintering typically sets in when the material is heated above 60% of its melting point and solid bridges are formed via diffusion of the particle material. Higher temperatures increase the speed of the diffusion process. Sintering reduces the internal surface of the system and leads to an energetically more favorable state.

**Figure 49:** $G'$ (closed symbols) and $G''$ (open symbols) vs. frequency for cocoa butter at 20°C, 30°C, 35°C and 40°C.
For cocoa particles, cocoa butter would be the diffusing material and particle bridging could therefore be a function of the diffusion properties of cocoa butter when the particles are in contact. The other option is for the cocoa butter to be in a semi-crystalline state at 30°C. This may correspond to an increased tackiness compared to the liquid state at 40°C or the solid, fully crystalline state at 20°C [127], [128].

The stickiness, or self-adhesion, of the cocoa butter can be correlated to the storage modulus $G'$ of the butter at given temperature. Modulus values $G' \approx 10^5$ Pa correspond to a pronounced tackiness of soft polymers as pointed out by Dahlquist [129] who quoted that good tack for pressure sensitive adhesives is achieved for $G' < 3.3 \times 10^5$ Pa at low frequency. According to Figure 49, the modulus $G'$ of cocoa butter at 30°C and 35°C is in the range of $10^5$ Pa, so a good tack or stickiness should be provided. Values for cocoa butter at 20°C exceed this range ($G' \approx 10^8$ Pa) and no stickiness can be expected anymore. At 40°C, cocoa butter is liquid and accordingly does not provide any tackiness. Therefore, particles that come into contact at 30°C will stick together due to adhesive forces when cocoa butter is located at the surface. Moreover, I like to emphasize that the standard system is in quiescence during the tempering process, so shear-induced agglomeration can be excluded.

![Photographic images of diluted cocoa suspensions stored at 20°C (A), 30°C (B) and 40°C (C).](image)

As proposed earlier [86], capillary bridges may form from liquid cocoa butter spread over the particles surface, when particles come in contact, but this is not evident here. At 40°C, the suspension behaves similarly to the suspension stored at 20°C. For visual analysis of the samples at different temperatures, cocoa suspensions where diluted and corresponding images are shown in Figure 50 confirming that no agglomerates are formed at 40°C as well as at 20°C, whereas at 30°C a clear floc formation is visible in the diluted samples. It has been reported earlier that the addition or the presence of too much secondary fluid may lead to spherical agglomeration [6], but as shown here this is not the reason for the reduced viscosity at 40°C.

In order to probe the resistance of the network against shear, one sample stored at 30°C was continuously stirred during storage time using a magnetic stirrer bar at 200 rpm. In Figure 48, the viscosity of the stirred sample is plotted in comparison to the sample left in quiescence at the same temperature. The low shear viscosity of the stirred sample is clearly lower than that of the sample
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stored at rest. Continuous shearing either diminishes the formation of bridges between the particles or larger capillary agglomerates are formed resulting in a weaker network [3], [130].

![Graph showing viscosity and temperature changes](image)

**Figure 51: Continuous shearing of a cocoa suspension (ϕ = 15%) at ˙\( \gamma \) = 0.1 s\(^{-1}\) during cooling from 40°C to 20°C.**

Figure 51 shows the viscosity of a cocoa sample measured at ˙\( \gamma \) = 0.1 s\(^{-1}\) during a temperature ramp from 40°C down to 20°C. It is demonstrated that the viscosity of the sample once heated to 40°C will not go to the level found for the sample stored at 20°C for 24h. Instead, the viscosity increases to the value found for the sample stirred during storage at 30°C (30 Pa s) and remains at this high level even upon further cooling to 20°C. Once the cocoa butter is released from the particle interior, bridges will be liquid and unstable at 40°C, stable and viscoelastic at 30°C and solid at 20°C. As we have a continuous shearing process while measuring, the viscosity is close to the “30°C stirred” level shown in Figure 48 and not the level of the standard 30°C sample stored at rest.

### 4.2.2 Extraction of cocoa particles

The influence of cocoa butter included in the particles was further investigated by preparing suspensions of particles with the cocoa butter extracted. Particle extraction was performed using ethanol and n-hexane. N-hexane removes the fatty phase whereas ethanol is also capable of removing polar substances from the particles. Extracted particles have been evaluated considering particle size as described in the materials and methods section and wetting behavior. The wetting behavior has been analyzed via the sessile drop method. In the following diagrams (Figure 52) contact angle and change in droplet volume trends are shown for natural and extracted cocoa powders.
Results and discussion

Figure 52: Contact angle of a sessile drop of water on a tablet pressed from natural cocoa and cocoa extracted with different solvents (left). On the right droplet volume is compared to the contact angle for natural cocoa (■) and n-hexane extracted particles (▲). Open symbols represent the drop volume over time.

In the left diagram it is shown that the contact angles are changing when the particles have been treated with an extracting solvent. Water extracted particles show a similar trend as natural particles, whereas particles treated with ethanol show a decreased contact angle, but similar gradient. Alcohol soluble substances have been removed but the particle is not entirely non-polar. Particles extracted with n-hexane show the lowest contact angle since all non-polar substances have been removed from the particles. Since the contact angle drops very fast over time compared to the other cases, it has to be assumed that the n-hexane extracted tablet is more porous. To have a closer look, the volume of the contact angle has been calculated and also plotted over time. Drop volume for n-hexane extracted particles decreases faster than for natural cocoa, whereas it is almost constant. Since the slope of the contact angle over time is steeper than the decreasing drop volume, spreading of the droplet is dominating over the subsidence. This confirms that the contact angle of n-hexane extracted particles is clearly smaller than that of natural cocoa as well as water and ethanol extracted particles.

Figure 53: Viscosity curves of suspensions (φ=15%) prepared with untreated particles and particles extracted with n-hexane, water and ethanol stored at 20°C (left), 30°C (middle) and 40°C (right).
In Figure 53, the flow behavior of suspensions made from differently treated particles at different temperatures is displayed. Particles extracted with n-hexane and ethanol do not show an increase in low shear viscosity when stored at 30°C while the untreated and the water extracted system clearly exhibit an increased low shear viscosity as was previously discussed. These findings demonstrate that the residual cocoa butter is critical for the structure formation and unusually high viscosity of the suspension stored at 30°C. At 20°C and 40°C storage temperature, viscosity curves for suspensions of extracted and non-extracted particles superpose. In summary, the storage temperature does not affect the flow behavior of suspensions made from n-hexane or ethanol extracted cocoa particles and this is attributed to the absence of the cocoa butter. Cocoa particles that have been extracted with water form suspensions that inhibit even at 20°C a higher viscosity than untreated particle suspensions. This may be due to the extraction of water soluble components (e.g. phenolic substances, organic acids (lactic-, acetic- and citric acid), proteins and free amino acids as well as carbohydrates (saccharose)) that may lower the low shear viscosity of the suspension by stabilizing the particles. Also, as can be seen in Figure 16, the extraction process breaks the cocoa agglomerates, which is reducing the average particle size. Smaller particle sizes result in a higher low shear viscosity if attractive particle interactions dominate.

Figure 54: Yield stress plotted over particle volume fraction of water continuous cocoa suspensions stored at 30°C for 12 h. Particles with a cocoa butter content of 11% and 22% were used.

Cocoa particles with different amounts of remaining cocoa butter are commercially used. Here I have investigated suspensions of cocoa particles with 11% residual cocoa butter as standard and also cocoa particles with 22% remaining butter for comparison. In Figure 54 the yield stresses of cocoa suspensions with varied particle volume fraction and the two different cocoa butter contents stored at 30°C are plotted. The yield stress characterizes the strength of the particle network within the suspension similar as the low shear viscosity of the samples. The yield stress increases with increasing particle loading, but below \( \phi = 5\% \), no yield stress can be detected. Samples made from cocoa with 22%
of residual cocoa butter exhibit a higher yield value throughout the whole concentration range. More cocoa butter obviously leads to a stronger network and we attribute this to the formation of more and larger cocoa butter bridges between the particles.

4.2.3 Effect of secondary fluids on flow behavior of aqueous cocoa suspensions

As written in section 4.1 for oil continuous suspensions, gelling can be achieved by adding a secondary immiscible fluid to the suspension. In this section the influence of added secondary fluids is investigated for water continuous suspensions.

Figure 55: Suspension of cocoa particles ($\phi=0.15$) in water without (A) and with (B) linoleic acid ($S=0.02$) added as secondary liquid.

For network formation in water continuous suspensions, the secondary liquid must be able to rewet the particles, meaning it has to remove water from the particle surface. Thus, a slightly polar fluid that is immiscible with water must be used. It has been found that fatty acids like linoleic acid or oleic acid are able to induce a network in cocoa suspensions. In Figure 55 the transition from a fluid-like suspension of cocoa in water (A) to a gel-like behavior (B) is seen, when linoleic acid is added as secondary liquid.

Figure 56: Yield stress over saturation of suspensions of cocoa particles in water for cocoa volume fractions of 15%, 20% and 25%. The saturation of secondary fluid (oleic acid) was varied from $S=0.0$ to $S=0.2$. The dotted line represents the yield value of a commercial cocoa spread (Rewe “Feine Welt”).
The increase in the yield stress due to the addition of oleic acid to the suspension of cocoa particles in water is shown in Figure 56 for different volume fractions of cocoa particles. Suspensions formed with linoleic acid and oleic acid show the same rheological behavior. An increase in the yield stress as high as in the oil continuous suspensions presented earlier is not observed. But up to an order of magnitude change can be measured and the attainable yield stress level is similar to that of a commercial fat continuous chocolate spread. It should be noted, that in contrast to the oil continuous suspensions, the yield stress of water continuous cocoa suspensions reaches a maximum value at lower fractions of secondary liquid.

Figure 57: REM images from dried cocoa samples without (left) and with saturations of $S=0.05$ (middle) and $S=0.10$ (right).

In Figure 57 REM images of dried cocoa particles (20°C, vacuum) are shown. The left picture shows the pure cocoa powder, whereas in the middle and right picture linoleic acid was added to the suspension prior to drying. For a saturation of $S=0.05$ a more agglomerated structure is detected. Particles are not assembled flat as can be seen in the left picture, but show a more round structure with more homogenous pores. On the right picture it can be seen that the surface properties have changed. The lighter spots in the image may be interpreted as fat that is located at the particle surface. This fat might be a mixture of linoleic acid and cocoa butter and could be contributing to the sticky particle surface as has already been described in section 4.2.1.

4.2.3.1 Influence of increased temperature

In Figure 58 the viscosity curves of suspensions with and without added oleic acid obtained at different storage temperatures are shown. A strong effect of oleic acid added to the suspension is visible for the suspension stored at 20°C. The suspension with added oleic acid shows a low shear viscosity almost two orders of magnitude higher than that of the pure suspension. In contrast, for 30°C the curves superpose but at a higher level than the 20°C curve with secondary fluid. Suspensions stored at 40°C do not show an increase of low shear viscosity upon the addition of oleic acid.
Different hypotheses can now be proposed regarding the effect of adding oleic acid to a cocoa suspension. First, the adsorption of oleic acid to the particle surface may render the surface of the cocoa particles more hydrophobic as reported for other particle systems [37], [131], [132]. According to Koh et al. [37], oleic acid added to a water based sheelite suspension absorbs to the particle surface inducing a hydrophobic attraction that causes particle agglomeration. Furthermore, they propose that simultaneously oleic acid can form liquid bridges between particles, holding them together. These two, probably interfering processes, may also occur for the cocoa particle system described here. The second possible mechanism is the dissolution and extraction of the cocoa butter distributed inside the cocoa particles by the oleic acid. Solving cocoa butter in oleic acid could result in a waxy, sticky surface around the particles gluing them together upon contact. Oleic acid may act as a plasticizer for cocoa butter having a similar effect on mobility and stickiness as increasing the temperature. This phenomenon has been investigated by mixing oleic acid and cocoa butter on a ratio that occurs in a cocoa particle (residual cocoa butter of 11%) suspension with saturation $S = 0.02$ and $\phi = 15\%$ ($80 \text{ ml of oleic acid and } 110 \text{ g cocoa butter}$) on a shaking table for 24 h. $G'$ and $G''$ data for this mixture are compared to values of cocoa butter at different temperatures. This is plotted in Figure 59. The moduli of the cocoa butter/oleic acid mixture coincide with cocoa butter data at 30°C. So the stickiness of the mixture is confirmed. This supports the thesis that a sticky layer is causing the particles to form a network upon contact whether cocoa butter is softened by appropriate heating or by mixing with oleic acid.
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Figure 59: $G'$ (closed symbols) and $G''$ (open symbols) vs. frequency for cocoa butter at 20°C, 30°C and 40°C as well as a mixture from cocoa butter and oleic acid. The mixture was created using volume fraction as would occur in suspensions with a secondary fluid saturation of $S = 0.02$.

At 40°C cocoa butter is in a fluid state and the confocal microscope images (Figure 60, right) show that the liquid cocoa butter is at least partly removed from the particles surface appearing as separate emulsion droplets. These droplets apparently cannot form capillary bridges to create a sample-spanning network and rather remain emulsified in the suspension, which explains its fluid-like flow behavior. Images of the 30°C sample, however, show a blurry distribution of the cocoa butter which is presumably at the particles surfaces or located between them thus enabling a network formation but with a bridge size too small to be distinguished individually. At 20°C, the signal of the cocoa butter is much weaker than the residual signal of the particles indicating that the butter remains inside.

Figure 60: Confocal microscopy images of cocoa particles suspended in water $\phi = 15\%$, for storage temperatures of 20°C (left), 30°C (middle) and 40°C (right). The yellow signal is generated by the cocoa particles and the red channel denotes to the cocoa butter. The fluorescence of cocoa particles has been suppressed by washing them in Toluidene Blue and the cocoa butter has been stained with Nile Red.

The effect of added oleic acid on suspensions of cocoa particles from which cocoa butter was extracted using n-hexane has also been investigated. The corresponding viscosity data are also shown in Figure 58, clearly demonstrating that the addition of oleic acid does not change the rheology of
cocoa suspensions when cocoa butter is removed. This result leads to the conclusion that the high low shear viscosity achieved upon adding oleic acid is a result of particle bridges formed from cocoa butter/oleic acid mixtures and cocoa butter is the key parameter controlling the flow behavior of aqueous cocoa suspensions. To further understand the network formation process, the viscosity of suspensions with and without added oleic acid stored at 20°C was measured over time of storage, starting right after sample preparation and continuing with discrete steps using a new sample for every measurement point. Corresponding viscosity data taken at $\dot{\gamma}=0.1\text{s}^{-1}$ is plotted in Figure 61. Viscosity increases in all cases but with different slopes. Samples with added secondary fluid exhibit higher initial viscosities, but the viscosity values match after 7h for the samples stored at 30°C as already shown in Figure 58. The sample with oleic acid stored at 20°C needs longer time for network formation, but after 10h it reaches a similar viscosity level as those stored at 30°C. This further supports the hypothesis that cocoa butter is dissolved by the secondary fluid as already described in the previous section. Obviously, the process of melting the cocoa butter at 30°C is faster than dissolving it in oleic acid at 20°C.

Figure 61: Time evolution of network formation for pure cocoa samples ($\phi=15\%$) stored at 20°C and 30°C with and without added secondary fluid. Viscosity data was measured at a shear rate of $\dot{\gamma}=0.1\text{s}^{-1}$. The rheometer was filled with a fresh sample for each measurement.

### 4.2.3.2 Variation of secondary and bulk fluids

The effect of adding oleic acid to cocoa suspensions has been discussed so far. Now, different fluids immiscible with water but comprising different functional groups, varied hydrocarbon chain lengths and entirely unpolar oils will be compared. These substances and their characteristic features are listed in Table 7. In contrast of what has been found for oil-based capillary food suspensions [86] interfacial tension is not the decisive factor for network formation and thickening, since sunflower oil and oleic acid show similar equilibrium interfacial tensions against water at 20°C, but a change in viscosity can only be detected when oleic acid is added.
Table 7: Characteristics and solubility of immiscible liquids added as secondary fluids to cocoa suspensions

<table>
<thead>
<tr>
<th></th>
<th>octyl acetate</th>
<th>octanoic acid</th>
<th>oleic acid</th>
<th>octanol</th>
<th>sunflower oil</th>
<th>silicon oil AK10</th>
</tr>
</thead>
<tbody>
<tr>
<td>functional group</td>
<td>-OCOOH</td>
<td>-COOH</td>
<td>-COOH</td>
<td>-OH</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>interfacial tension</td>
<td>20 mN/m</td>
<td>8 mN/m</td>
<td>14 mN/m</td>
<td>8 mN/m</td>
<td>22 mN/m</td>
<td>39 mN/m</td>
</tr>
<tr>
<td>chain length</td>
<td>8 C</td>
<td>8 C</td>
<td>18 C</td>
<td>8 C</td>
<td>16 C-20 C</td>
<td>-</td>
</tr>
<tr>
<td>capillary effect</td>
<td>✓</td>
<td>✓</td>
<td>✓</td>
<td>✓</td>
<td>X</td>
<td>X</td>
</tr>
<tr>
<td>solubility of cocoa butter</td>
<td>&lt; 1h</td>
<td>1:42 h</td>
<td>8 h</td>
<td>8:15 h</td>
<td>&gt;58 h</td>
<td>insoluble</td>
</tr>
</tbody>
</table>

Yield stress values obtained at different times after preparation of the two-fluid cocoa suspensions ($S = 0.01$) are plotted in Figure 62. Obviously, secondary fluids exhibiting a shorter chain length (C8 vs. C18) induce faster increase of yield stress than fluids with longer chains like oleic acid. After a storage time of 100h suspensions prepared with secondary fluids of different chain length show the same yield value. This leads to the conclusion that the chain length of the secondary fluid determines the kinetics of network formation in these suspensions, but does not affect the final network strength despite the different interfacial tension.

Figure 62: Yield stress over time for aqueous cocoa suspensions ($\phi = 15\%$) with different secondary fluids ($S=0.01$). The rheometer was filled with a fresh sample for every measurement point.

Different functional endgroups obviously do not result in a measurable difference with respect to the yield stress or gel strength of the suspension. However, suspensions with added sunflower oil or silicon oil do not show any measurable effect on flow behavior compared to the regular bulk suspension. We attribute these findings to a different solubility of cocoa butter in the different
secondary fluids. Cocoa butter did not completely dissolve in sunflower oil or silicone oil even after 58 h. In contrast, cocoa butter dissolved completely in octanoic acid within 2 h and in oleic acid within 8 h. Mixtures of cocoa butter with these organic acids form wax-like, soft, sticky fluids at the concentration ratios present in the cocoa suspensions ($\phi = 15\%$, residual cocoa butter within particles 11%, $S = 0.02$) and this seems to be the prerequisite for the formation of network structures similar to those developing when pure aqueous cocoa suspensions are heated to $T = 30^\circ C$.

Mixtures from oleic acid and sunflower oil have been prepared and added to the cocoa suspension as secondary fluid. The yield stress of the resulting suspensions is plotted in Figure 63. The higher the amount of oleic acid in the mixture, the stronger the suspension gels. Samples with only sunflower oil as secondary fluid hardly show an increase in yield stress, which is $\sigma = 40.0 \pm 2.1$ Pa with added sunflower oil compared to $\sigma = 23.2 \pm 6.5$ Pa without added secondary fluid. This increase may be ascribed to the low fraction of free fatty acids that are regularly present in the commercial oil. This further confirms that only free fatty acids or other slightly polar substances can cause a sample spanning network in aqueous cocoa suspensions, whereas entirely non-polar triglycerides do not contribute to particle clustering.

![Figure 63: Yield stress over fraction of oleic acid in secondary fluid for cocoa suspensions ($\phi = 20\%, S = 0.01$).](image)

Bulk fluid viscosity has been varied by adding different concentrations of fructose and to the water phase. Adding glycerol and fructose is not only changing the viscosity of the bulk phase, but also influences the three phase wetting behavior. In Figure 64 the normalized yield stress is plotted over the bulk viscosity (left) and over the interfacial tension between bulk and secondary phase (right).
Results and discussion

Figure 64: Normalized yield stress of cocoa suspensions with aqueous mixtures from fructose and glycerol as bulk fluids plotted over the bulk viscosity (left) and the interfacial tension (right).

Secondary fluid (linoleic acid) saturation was kept constant at $S = 0.02$.

A weak linear dependency of normalized yield stress on bulk viscosity, but not on interfacial tension is detected. In oil continuous capillary suspensions we did find that the formation of capillary suspensions is not a function of droplet breakup, but a diffusion driven process (see section 4.1.4). Since the network formation in water continuous suspension is a slow process (see Figure 62) compared to network build-up in oil continuous suspension the influence of diffusion might be more distinct and observable here. The diffusion coefficient $D$ is not a function of interfacial tension but inversely proportional to the bulk viscosity,

$$D = \frac{k_B T}{6 \pi \eta R_0}$$  \hspace{1cm} (4.3)

with $k_B$ being the Boltzmann constant and $R_0$ the hydrodynamic radius of the diffusing molecule. In contrast, droplet breakup is proportional to bulk viscosity $\eta$ and inversely proportional to interfacial tension $\Gamma$ assuming laminar mixing conditions and droplet breakup to occur for $Ca > 1$. But since turbulent conditions can be assumed when using a dispenser mixer blade, interfacial tension does account for droplet size $d_{dr}$ (2.16). It cannot be clear said, but it is assumed that the network formation is a combination of mechanisms, droplet breakup and diffusion.

4.2.3.3 Structural information about cocoa suspensions from SANS and micro computed tomography

Small angle neutron scattering experiments were used to identify structural changes upon heating or addition of secondary fluid to water based cocoa suspensions. In Figure 65, the scattering curves of samples made from $\phi=15\%$ cocoa powder in water at 20°C with and without secondary fluid and at 30°C as well as 40°C without secondary fluid are shown. The scattering intensity strongly decreases with increasing q-value in all cases. The samples stored at 20°C and 30°C exhibit a characteristic peak in the high q-range. For the pure cocoa suspensions this peak occurs at $q = 0.173 \text{ Å}^{-1}$ corresponding to
a characteristic length of 36 Å. We can interpret this as a bilayer of triglyceride molecules. For the samples with added oleic acid this length increases to 51 Å and a characteristic length is 53 Å is found for the pure suspension heated to 30°C. This would then correspond to a triple layer of triglyceride. No peak is found for the sample stored at 40°C.

The characteristic length in the suspensions revealed by the peak in the scattering intensity is interpreted as a layer of cocoa butter covering the cocoa particles. The thickness of this layer increases when cocoa butter leaks to the particle surface at elevated temperatures or when oleic acid is added. No surface layer of cocoa butter exists at 40°C. As already shown in Figure 60, the cocoa butter separates from the particles at this temperature and forms large droplets which do not contribute to the scattering intensity in the q-range investigated here. The surface layer observed in SANS experiments may not be sufficient to facilitate bridging between particles considering their irregular shape but its change is another indication for the leaking of the butter from the particle interior at elevated temperatures or upon addition of a secondary fluid. The stickiness of the surface layer provided either by the mixture of cocoa butter and oleic acid or by the pure cocoa butter at elevated temperature is decisive for the formation of a sample spanning network structure and a distinct rheological behavior corresponding to a creamy texture.

Figure 65: Scattering intensity I vs. scattering vector q for cocoa in water samples ($\phi=15\%$) at temperatures of 20°C, 30°C and 40°C and 20°C with added oleic acid ($S=0.02$).

Structural changes in aqueous cocoa suspensions can also be detected using micro tomography. Suspensions with and without added linoleic acid have been dried in vacuum (20°C) and carefully put in a phoenix nanotom (GE measurements & control, located at Max Planck Institute, Göttingen). Images resulting from these measurements are shown in Figure 66. Samples without secondary fluid appear dark and unstructured since detection of individual cocoa particles is below the resolution limit of the device (1µm). In contrast, samples with secondary fluid show a more structured image. Cocoa particle agglomerates are seen to be uniform in size and a uniform network structure is visible throughout the sample. Due to the limited solution of the microtomography device individual liquid
bridges between particles cannot be resolved. For the same reason it is not possible to determine coordination numbers or the fractal dimension of the network.

Figure 66: microtome images of dried cocoa suspensions without (left) and with (right) added secondary fluid.
5 Conclusions

Capillary suspensions offer new opportunities for the formulation of food products. We can see a clear transition from a liquid or very weak gel with no secondary phase added, to a strong gel-like state when water is added to the oil continuous suspensions made from cocoa or starch granules. This indicates that strong capillary forces act in this food model systems as has been reported earlier for suspensions of inorganic or synthetic polymer particles [1]. Stability against sedimentation can be dramatically increased, texture can be modified and flow properties can be easily adjusted according to the demands of a broad variety of processing and application techniques and perception demands.

The wetting behavior of the particles with water and oil has been documented using confocal microscopy as well as contact angle measurements using the sessile drop method. Various rheological parameters of capillary suspensions have been documented and compared, revealing that oscillatory shear measurements underestimate the yield stress obtained from rotational shear data. Texture modifications can be done mainly by adapting the amount of the secondary fluid, but also by changing the wetting properties like the contact angle the two fluids form with the particles. On the other hand, the secondary fluid viscosity has no influence on rheological quantities like yield stress or low shear viscosity of the investigated capillary suspensions. Furthermore, the adsorption of water by particles from environmental sources during storage can dramatically influence the properties of oil continuous suspensions since capillary bridges form spontaneously from adsorbed water layers. Suspension rheology is similar if the water is absorbed prior to suspension preparation or added to the suspension afterwards. These findings lead to the conclusion that the formation of these capillary suspensions is energetically favorable and not controlled by mechanical energy input. This aspect might be of special relevance from a technical or processing perspective.

Capillary bridges offer an additional advantage to chocolate or other temperature sensitive systems by preserving shape stability even if the continuous phase of the suspension is molten. This stability has been documented via optical experiments and also was measured using rheological setups. Increased volume of secondary fluid as well as higher volume fractions of particles lead to a more stable product.

The rheological behavior of aqueous cocoa particle suspensions stored at different temperatures or modified by the addition of a secondary fluid has also been investigated in this study. When cocoa particles are dispersed in water and stored at a temperature of 30°C, a network of particles is formed which leads to an increase in low shear viscosity and a gel-like, creamy texture. Heating the sample further up to 40°C or above results in a network collapse and the suspension behaves fluid-like again. The critical factor controlling this behavior is the residual cocoa butter in the particles. At 30°C, the cocoa butter is in a semicrystalline state and bridges of this cocoa butter between the particles are formed via a diffusion process (similar to sintering) or just by sticking together upon contact due to cocoa butter that has leaked to the surface. The cocoa butter is completely liquid at 40°C and partly
forms emulsion droplets in the water phase that do not contribute to the particle network as shown via confocal imaging. The key role of cocoa butter for network formation, texture and flow of the aqueous suspensions has been proven by extracting it from the particles as well as increasing the amount of cocoa butter. Particles, where the fat has been removed show no temperature dependent change in flow behavior. Particles with higher butter content form stronger networks as proven for a wide range of particle loadings. Samples, once heated above 30°C, will not go back to their initial state at 20°C and remain in a gelled state. High network strength is obtained upon storage over extended periods of time at rest (>24h).

Adding a secondary fluid to the cocoa suspension results also in an increased yield stress attributed to the formation of a network of particles even at room temperature. This bridging phenomenon only appears for particles with residual cocoa butter content. This leads to the assumption that the secondary fluid dissolves the cocoa butter and similar to the heated particle suspensions a soft and sticky semi-crystalline fluid is created that glues the particles together. This process is faster for short chain solvents with a polar end-group compared to longer chain fatty acids and can be correlated to the speed at which the respective fluid dissolves the cocoa butter. Less polar substances like silicone oil or sunflower oil do not induce a particle network formation, which is attributed to the poor and slow solubility of cocoa butter in these fluids.

SANS experiments also revealed structural differences between samples stored at different temperatures. Samples stored at 30°C or at 20°C, but with added oleic acid exhibit a scattering peak corresponding to a characteristic length scale of about 50 Å whereas the characteristic length reduces to about 35 Å for a pure cocoa suspension stored at 20°C. This length scale is attributed to the cocoa butter layer on the particle surface and completely vanishes upon storage at 40°C.

Finally, we have shown how the flow behavior of oil and water continuous food suspensions and correspondingly their creaminess and texture can be tuned in a wide range via appropriate thermal treatment or addition of an adequate, immiscible secondary fluid. This may open up a new pathway for the formulation of novel food products based not only on cocoa but any other finely dispersed particle species.
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