Thymosin β4 Improves Differentiation and Vascularization of EHTs

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Abstract

Induced pluripotent stem cells (iPSC) constitute a powerful tool to study cardiac physiology and represent a promising treatment strategy to tackle cardiac disease. However, iPSCs remain relatively immature after differentiation. Additionally, engineered heart tissue (EHT) has been investigated as a therapy option in preclinical disease models with promising results, although their vascularization and functionality leave room for improvement. Thymosin β4 (Tβ4) has been shown to promote the differentiation of progenitor cell lines to cardiomyocytes while it also induces angiogenic sprouting and vascular maturation. We examined the potential impact of Tβ4 to enhance maturation of cardiomyocytes from iPSC. Assessing the expression of transcription factors associated with cardiac differentiation, we were able to demonstrate the increased generation of cells displaying cardiomyocyte characteristics in vitro. Furthermore, in a zebrafish model of embryonic vascular development, Tβ4 is crucial for the proper execution of lymphatic and angiogenic vessel sprouting. Finally, utilizing Tβ4-transduced EHTs generated from mice genetically engineered to label endothelial cells in vitro, we show that treatment with Tβ4 promotes vascularization and contractility in EHTs, highlighting Tβ4 as a growth factor improving the formation of cardiomyocytes from iPSC and enhancing the performance of EHTs generated from neonatal cardiomyocytes.

1. Introduction

Engineered heart tissue generated from neonatal cardiomyocytes has been a useful tool to study cardiac physiology and presents a promising treatment option for patients with heart failure or excessive scar tissue formation after myocardial infarction, which has been demonstrated in preclinical animal models of myocardial replacement therapy [1, 2]. However, major obstacles in generating functioning EHT-grafts of sufficient size still remain. For instance, while grafts can generate primitive vascular networks [3], those vessels typically are unable to support EHTs of increasing thickness [4]. Additionally, myocardial-grafts derived from neonatal cardiomyocytes lack the level of maturity seen in adult myocardium. On the one hand, those EHTs do display features of fully matured, adult cardiomyocytes, such as the expression of contractile proteins, their organization into sarcomeric structures, and the capability to mimic basic cardiomyocyte physiology [5–7]. However, thusly generated engineered heart tissue grafts still have a higher ratio of fetal myosin heavy chain and are unable to generate comparable force [8], which reflects their relative immaturity.
Thymosin β4 (Tβ4), although first described as a G-actin sequestering peptide regulating cytoskeletal rearrangement and cellular motility [9], has recently been shown to possess additional functions during cell fate determination and angiogenesis. Thus, it was demonstrated that Tβ4 in mouse models of myocardial ischemia promotes the proliferation of mesenchymal stem cells, improving cardiac function [9], and advances the transdifferentiation of epicardial progenitors into integrated cardiomyocytes [10]. Furthermore, Tβ4 was shown to facilitate angiogenic sprouting, by activating SRF target genes via the nuclear translocation of MRTF-A [11], and to foster the attachment of mural cells to endothelial cell tubes, improving the functionality of vessels [12, 13].

On account of Thymosin β4 promoting cardiomyocyte differentiation and improving tissue supply with functioning vasculature, we investigated the effect of Tβ4 on the differentiation of iPSCs to myocardial cells and its impact on the quality of engineered heart tissues.

2. Materials and Methods

2.1. Generation of Induced Pluripotent Stem Cells. Induced pluripotent stem cells derived from murine embryonic fibroblasts were kindly provided by Ulrich Martin [14]. Cells were cultured on mitotically inactivated murine embryonic fibroblasts (MEFs; 50,000 cells/cm²) in 6-well plates. The culture medium was composed of Dulbecco’s modified Eagle’s medium (Invitrogen, Carlsbad, Calif) with 15% fetal calf serum (Thermo Fisher Scientific, Germany), 0.2 mmol/L L-glutamine (Invitrogen), 0.1 mmol/L β-mercaptoethanol (Invitrogen), 0.1 mmol/L nonessential amino acid stock (Invitrogen), and 0.1% human leukemia inhibitory factor-conditioned medium, which had been produced by transient transfection of a human leukemia inhibitory factor plasmid into human embryonic kidney 293 cells. To initiate embryoid body formation, cells were cultured in the “hanging drop” technique with 600 cells/20 µL of differentiation medium, consisting of Iscove’s modified Dulbecco’s medium with 15% FCS, 0.2 mmol/L L-glutamine, 0.1 mmol/L β-mercaptoethanol, and 0.1 mmol/L nonessential amino acid stock. After 3 days of differentiation EBs were transferred onto nonadherent 1% agarose-coated 6-well plates. The culture medium was composed of Dulbecco’s modified Eagle’s medium (Invitrogen, Carlsbad, Calif) with 15% fetal calf serum (Thermo Fisher Scientific, Germany), 0.2 mmol/L L-glutamine (Invitrogen), 0.1 mmol/L β-mercaptoethanol, and 0.1 mmol/L nonessential amino acid stock (Invitrogen), and 0.1% human leukemia inhibitory factor-conditioned medium, which had been produced by transient transfection of a human leukemia inhibitory factor plasmid into human embryonic kidney 293 cells. To initiate embryoid body formation, cells were cultured in the “hanging drop” technique with 600 cells/20 µL of differentiation medium, consisting of Iscove’s modified Dulbecco’s medium with 15% FCS, 0.2 mmol/L L-glutamine, 0.1 mmol/L β-mercaptoethanol, and 0.1 mmol/L nonessential amino acid stock. After 3 days of differentiation EBs were transferred onto nonadherent 1% agarose-coated 6-well plates, followed by a transfer of 10 EBs per well on a 0.1% gelatin-coated 6-well culture dish.

2.2. Vascular Patterning in Zebrafish. Embryos and adult zebrafish were raised and maintained under standard conditions. The transgenic zebrafish line Tg(fli1a:EGFP)13 was used as previously described [15]. Morpholinos, used to knockdown tmsb-like expression and control morpholinos, were injected into embryos at the 1 to 2 cell stage (15 ng/embryo). The developing vasculature was examined using the Leica SP5 confocal microscope.

2.3. Flow Cytometry. Induced pluripotent stem cells were trypsinized, washed in cold PBS, and fixed for 20 minutes at RT in 3.7% formaldehyde. Cells were permeabilized with 0.4% Triton X-100 in PBS and subsequently incubated with antibodies as indicated for 2 hours at 37°C in 0.4% Triton X-100 in PBS. Stainings were performed with antibodies against cardiac Troponin I (BD Pharamingen, San Diego, US), αMHC (BD Pharamingen, San Diego, US), and α-actinin (Sigma Aldrich, St. Louis, US). A FITC conjugated IgG antibody served as a control. After washing, cells were analyzed on a Coulter Epics XL-MCLTM flow cytometer (Beckman Coulter, Krefeld, Germany).

2.4. Quantitative Real-Time PCR. RNA from iPSCs was isolated at timepoints indicated and reverse-transcribed into cDNA. qPCR was performed on a q-i-cycler (Bio-Rad, Munich, Germany) by SYBR-Green Supermix (Bio-Rad, Munich, Germany). For qPCR, the following primers were used: Brachyury (T): upper: 5’ ACCCAGCTCTAAGGA-ACCAC 3’; lower: 5’ ACTCCGAGGCTAGACCATT 3’; Eomes: upper: 5’ CTCTAGACTCCAGGACTCC 3’; lower: 5’ GCCTTTGAGGTTGCTTTTAC 3’; Mesp: upper: 5’ CAGTACGGAACACAGCATC 3’; lower: 5’ GGTTCCT- TAGAAGGACCAGCA 3’; NKX-2.5: upper: 5’ CATTATT-CCGGGAGCCTACG 3’; lower: 5’ GCTTTGCTGC CGCCGTGC 3’; ANF: upper: 5’ GCAAATCCCTGTGAC-AGTG 3’; lower: 5’ CAGGTGTGGTACAGGTCTCT 3’; CX43: upper: 5’ CTTCACTGGTCTCTT 3’; CX43: lower: 5’ GTCGACACGACAGAATG 3’.

2.5. Generation of EHTs. Neonatal mouse heart cells were isolated from Cdh5-CreERT2 x Rosa26-LacZ mice, expressing LacZ in endothelial cells as described previously [16]. Cells were mixed with fibrinogen (1 mg/mL, Sigma, F4753), medium (DMEM), Matrigel (100 µL/mL, BD Bioscience, 356235), and thrombin (100 U/mL, Sigma, T7513) and subsequently casted into strip-format molds in agarose (dimensions 12 x 3 x 3 mm) with pairs of elastic silicone posts placed from above. After fibrinogen polymerization (2 hours, 37°C, 7% CO2), mold racks were transferred to a new 24-well plate and EHTs were cultured with DMEM (Biochrom F0415), supplemented with 10% horse serum (Gibco 26050), 2% chick embryo extract, 1% penicillin/streptomycin (Gibco 15140), insulin (10 µg/mL, Sigma I9278), tranexamic acid (400 µmol/L, Sigma 857653), and aprotinin (33 µg/mL, Sigma A153).

2.6. Skinfold Chamber. BALB/c nude mice were anaesthetized with an intraperitoneal injection of midazolam, medetomidine, and fentanyl and placed on a warming platform. After hair removal and fixation of the back skin via silk sutures, two openings at the base of the skinfold were prepared through which the connecting screws of the chamber were passed. The observation window was marked and one skin layer was removed. Subsequently, the previously generated EHT-grafts were sutured into the observation window and sealed with a cover glass kept in place by a snap ring. After fixing the second half of the chamber, anaesthesia was antagonized with atipamezole, flumazenil, and naloxone and mice were placed in individual cages until further use. For imaging of angiogenic sprouting derived from the EHT-grafts, whole mount X-Gal staining from Cdh5-CreERT2 x
Rosa26-LacZ derived EHTs was performed and subsequently imaged via light microscopy.

2.7. Statistical Analysis. Data are given as mean ± SEM. Differences among several groups were tested using ANOVA and the Student Newman Keuls post hoc analysis. A p value of < 0.05 was considered statistically significant. All data were assessed using the SPSS software package (version 20.0; http://www.spss.com). Sample sizes are provided in the figure legends.

3. Results and Discussion

3.1. Thymosin β4 Drives Cardiac Differentiation in Induced Pluripotent Stem Cells. To investigate the potential of Thymosin β4 (Tβ4) to induce cardiac differentiation in murine induced pluripotent stem cells (iPSC), we analyzed differences in the expression of a variety of transcription factors and cardiac markers at different timepoints during treatment of iPSCs with Tβ4 (Figure 1(a)). We found that during early stages of differentiation (day 4) Tβ4 significantly increased the expression of mRNA encoding the T-Bos-transcription factors Eomesodermin (Eomes) and Brachyury (T), which are indispensable for mesoderm formation in general and are known to be early mesoderm markers [17, 18] (Figures 1(b) and 1(c)). Apart from promoting the formation of early mesoderm, a treatment with Tβ4 also enhanced the expression of MesPi, the earliest marker of cardiovascular progenitors [19, 20], and NKX-2.5, which has been shown to be crucial for cardiac development [21] (Figures 1(d) and 1(e)). Similar results for the expression of MesPi and NKX-2.5 were observed in Tβ4 treated GSES embryonic stem cells (data not shown). Finally, Tβ4 treatment after differentiation for 6 days was able to significantly increase the expression of the myocardial proteins atrial natriuretic factor (ANF) and connexin 43 (CX43) and the production of the cardiac specific proteins alpha myosin heavy chain (αMHC), Troponin, and α-actinin (Figures 1(f)–1(h)). The increased expression of contractile cardiac proteins in iPSCs via the treatment of those cells with Tβ4 also enhanced the differentiation of iPSCs with Tβ4 (Figure 1(i)).

The mode of action of Tβ4 comprises two separate physiological processes: first, Tβ4 binds monomeric G-actin, preventing it from forming or joining actin filaments via profilin, which, upon binding, favors the incorporation of actin into filaments [22]. Secondly, Tβ4 can regulate the expression of a number of serum response factor (SRF) target genes. This effect depends on the nuclear translocation of the myocardin-related transcription factor A (MRTF-A), also a G-actin binding protein [23]. An increase in actin binding to Tβ4 frees actin-bound MRTF-A leading to its nuclear translocation and SRF-binding [24, 25]. Inside the nucleus, MRTF-A acts as a transcriptional coactivator physically associating with SRF [26]. Interestingly, SRF signaling appears to be indispensable for mesoderm formation. SRF-deficient mice die during early embryonic development starting from E9.5 with impaired gastrulation and a lack of mesodermal cells [27]. Furthermore, embryonic stem cells lacking SRF are incapable of driving Brachyury (T) expression upon induction of differentiation [28]. Additionally, Nam et al. demonstrated that MRTF-A is a potent inducer of cardiac protein expression during the reprogramming of human foreskin fibroblasts, increasing the amount of fibroblasts expressing cardiac troponin from 0.2% (2%) to 17% (13%) during GMT (or GHMT) induced cardiac differentiation [29]. Additionally, Thymosin β4 is well known to promote the differentiation of embryonic stem cells [30] and drive fibroblast to cardiomyocyte differentiation in models of myocardial ischemia [31]. Furthermore, Thymosin β4 has been shown to promote proliferation and reduce apoptosis under stress-conditions in mesenchymal stem cells [9], highlighting the possibility that Tβ4 might increase the pool of stem cells differentiating via proliferation and by shielding them from apoptosis. These findings point to the possibility that Tβ4 facilitates the differentiation of iPSCs to cardiomyocytes either by directly inducing differentiation, by increasing the abundance of mesodermal progenitors, which subsequently differentiate independent of Tβ4, or by protecting iPSCs during differentiation.

3.2. Inhibition of Thymosin β4 Induces Lymphatic Abnormalities in Zebrafish. To investigate the role of Tβ4 during development in zebrafish, we first sought to identify which possible Tβ4 isoform in zebrafish corresponds to the human and mouse isoforms. To this end, the amino acid sequences of Tβ4 from human and mouse were aligned with the zebrafish orthologs beta Thymosin and beta Thymosin-like (Figure 2(a)). Aligning these sequences demonstrated a closer homology of the consensus actin binding sequence [32] of beta Thymosin-like to the mammalian Thymosins than that of zebrafish beta Thymosin. Additionally, the genomic localization in regard to its proximity to tlr7 and egfl6 of tmsb-like more closely resembles that of murine and human Thymosin β4, as demonstrated in Figure 2(b). For these reasons, we assessed the effect of a morpholino mediated knockdown of beta Thymosin-like rather than beta Thymosin. Before evaluating the effect of tmsb-like morpholino mediated knockdown on vascular development, zebrafish embryos were microinjected with GFP-tagged tmsb-like in the presence or absence of tmsb-like morpholinos (tmsb-like-MO, Figure 2(c)) to investigate knockdown efficiency. This led to a clear expression of the GFP-tagged protein, while additional injection with tmsb-like-MO blunted the expression of GFP-tmsb-like, indicating the efficiency of our morpholino mediated Thymosin beta like knockdown (Figure 2(d)). The tmsb-like morpholinos and control morpholinos were subsequently injected into Tg(fli1.egfp)31 transgenic zebrafish, which express enhanced GFP under the control of the fli1 promoter in the entire vasculature to assess the embryonic development of the vasculature and lymphatic system [33].

In investigating morpholino treated zebrafish at day 5.5 postfertilization (dpf), varying degrees of edema formation were apparent in tmsb-like-MO treated animals while formation of edema was virtually absent in control-MO treated zebrafish (tmsb-like-MO with moderate edema: 40%, sever edema: 37%, Figures 3(a) and 3(b)). Since excessive
undiff. miPSC   d0   d4   d6   d8   undiff. miPSC

Tβ4   Beachedy/ Eomes

Differentiation   d6   d6 + 8

Flow cytometry

Beating areas

ANF/CX43

(a)

Figure 1: Continued.
Figure 1: Thymosin β4 promotes the generation of cardiomyocytes from murine induced pluripotent stem cells. (a) Undifferentiated iPSCs were treated with Tβ4-peptide and expression of transcription factor encoding mRNAs was measured 4 (Eomes, Brachyury (T)), 6 (Mespl), or 8 (NKX-2.5) days after Tβ4 treatment (red). In another approach, murine iPSCs were differentiated for 6 days and treated with Tβ4-peptide. 8 days after Tβ4 treatment, expression levels of cardiomyocyte markers (ANF, CX43) were measured and flow cytometric assessment of cardiac specific proteins (αMHC, Troponin, and α-actinin) was performed. Furthermore, the amount of beating areas was measured (blue). (b–g) Relative expression levels of transcripts as indicated ((b–e) without prior differentiation of iPSCs, (f and g) with 6 days of prior differentiation), measured via quantitative real-time PCR, reveal that Tβ4 induces the expression of transcription factors regulating the differentiation of stem cells to cardiomyocytes via mesenchymal and cardiac progenitor stages and the expression of cardiac specific transcripts. (h) Flow cytometry after intracellular staining for αMHC, Troponin, and α-actinin reveals the increased presence of these cardiac specific proteins after Tβ4 stimulation. (i) Tβ4 treated iPSCs show a high rate of spontaneous beating compared to untreated iPSCs in which beating was virtually absent. (*P < 0.05).

edema formation is associated with malformation of the lymphatic system and defective thoracic duct formation (TD) [34–36], we investigated if a tmsb-like knockdown impairs the thoracic duct development by assessing its presence in 10 somites. These results revealed that, in tmsb-like-MO treated animals, TD formation was severely impaired at 5.5 dpf with no animals displaying TD formation in all somites and 68% showing no duct formation at all, whereas in control-MO treated animals 96% showed proper TD formation (Figures 3(c) and 3(d)). Since the thoracic duct is preceded by the emergence of parachordal lymphatic vessels (PC) that start to sprout from the embryonic posterior cardinal vein (PCV) at around 32 hours postfertilization (hpf) [37, 38], we investigated the influence of a tmsb-like KD on the pattern of PC-formation at 60 hpf by in vivo confocal imaging of Tg(fli1-egfp)y1 zebrafish. At this stage, the already formed main vessels (dorsal aorta (DA) and posterior cardinal vein (PCV)), which form approximately at 1 dpf [39], displayed no apparent phenotype. Intersegmental vessels (ISV) and the dorsal longitudinal anastomotic vessels (DLAV), however, seemed disorganized and displayed a hypersprouting phenotype in tmsb-like-MO treated animals compared to control (Figures 3(e) and 3(g)). Furthermore, PC-development was severely impaired as demonstrated by a reduction in parachordal lymphangioblast carrying somites (Figures 3(e) and 3(f)). The role of Tβ4 in lymphatic vessel development remains poorly understood, while the effect of Tβ4 on blood vessel maturation has been studied intensively in recent years, demonstrating the potent stabilizing effect of Tβ4 on newly formed vessels. To this end it has been shown that Tβ4, apart from inducing angiogenic sprouting via the activation of CCN1 [40], promotes the recruitment of mural cells (pericytes in particular) to the vasculature via the activation of MRTF-A and the downstream induction of the SRF target gene CCN2 [41]. Thus, the disorganization of the intersegmental vessels and the dorsal longitudinal vessels may be based on the absence of stabilizing mural cells. The hypothesis of increasing vascular destabilization in the absence of tmsb-like was further supported by the observation that tmsb-like-MO treated animals suffered from a high rate of cerebral vascular hemorrhages, a hallmark of unstable vessels due to the lack of mural cells [42], while hemorrhage in control-MO treated animals was rarely seen, pointing at the presence of unstable vessels in tmsb-like-MO animals (Figures 3(h) and 3(i)). Similar phenotypic changes, an increase in vascular hemorrhage due to a reduction in mural cell coverage, have been demonstrated in mouse models of Tβ4 deficiency, indicating the importance of Tβ4 in the proper vascular development throughout vertebrate species [13]. In the case of the lymphatic system, the incompetence in lymphatic vessel sprouting seen in tmsb-like-MO animals (Figures 3(h) and 3(i)). Similar phenotypic changes, an increase in vascular hemorrhage due to a reduction in mural cell coverage, have been demonstrated in mouse models of Tβ4 deficiency, indicating the importance of Tβ4 in the proper vascular development throughout vertebrate species [13].
Figure 2: Identification of a Thymosin β4 ortholog in zebrafish. (a) Amino acid sequence alignment of zebrafish beta Thymosin and beta Thymosin-like and mouse and human Thymosin β4. The yellow box indicates the consensus actin binding motif, indicating a closer degree of homology of zebrafish beta Thymosin-like to the mammalian Thymosins than beta Thymosin. (b) The comparison of the genomic localization of beta Thymosin and beta Thymosin-like in zebrafish, in relation to the murine and human localization, shows a closer proximity of beta Thymosin-like than beta Thymosin to the mammalian Thymosin β4. (c) Vector used to integrate pEGFP-N1-tmsb-like in zebrafish with indicated binding site of tmsb-like morpholinos (tmsb-like-MO) used to knockdown tmsb-like in zebrafish. (d) Treatment of pEGFP-N1-tmsb-like expressing zebrafish embryos with tmsb-like-MO demonstrates high knockdown efficiency.

for lymphangioblast migration from the posterior cardinal vein [37].

3.3. Treatment of Engineered Heart Tissue with Tß4 Improves Vascularization and Contractility. Having demonstrated that treatment of iPSCs with Tß4 increases the abundance of functional cardiomyocytes and knowing that Tß4 promotes vascular stability and is necessary for proper angiogenic sprouting, we sought to investigate if a treatment of engineered heart tissue (EHT) from murine heart cells with Tß4 promotes the vascularization of these tissue grafts. To this end, EHTs were generated from neonatal mouse heart cells of Cdh5-CreERT2 × Rosa26-LacZ mice, expressing LacZ in endothelial cells, and treated with Tß4-peptide. Initially, cross sections of EHTs revealed a significantly higher capillary density if treated with Tß4 (Figure 4(a)) and displayed...
an increase in contractile force (data not shown). Subsequently, EHTs were implanted into skinfold chambers in mice. Already macroscopically, Tβ4 treated, implanted EHTs showed a dramatic increase in vascularization (Figure 4(b)), which was confirmed by quantifying the vessels per quadrant of those EHTs 5 days after implantation (ctrl.: 8.4 ± 1.9, Tβ4 14.8 ± 1 vessels/quadrant) which became even more prominent 8 days after implantation (ctrl.: 6.4 ± 0.8, Tβ4 24.5 ± 2.4 vessels/quadrant, Figures 4(b) and 4(c)). In addition, intravenous injection of FITC-dextran demonstrated that these vessels are perfused, highlighting the functionality of these vessels (Supplementary Movie 1 in Supplementary Material available online at https://doi.org/10.1155/2017/6848271). To investigate the origin of these new vessels, we implanted EHTs from Cdh5-CreERT2 × Rosa26-LacZ mice after Tβ4 or control treatment and observed, after β-galactosidase
Figure 4: Increased vascularization and contractility in engineered heart tissue under Thymosin β4 treatment. (a) Cross sections of EHTs from reporter mice expressing LacZ in endothelial cells (Cdh5-CreERT2 × Rosa26-LacZ) reveal an increase in capillary density if treated with Tβ4. (b and c) AAV-mediated transfection with Tβ4 (rAAVβ4, versus rAAVGFP as control) of EHTs implanted into skinfold chambers leads to an increase of graft-vascularization, both 5 and 8 days after implantation (n = 4 per group, *p < 0.05; **p < 0.0008). (d) Images of whole skinfold chambers containing EHTs generated from LacZ reporter mice show little sprouting of graft resident endothelial cells into host tissue, while rAAVβ4 treatment drastically induces angiogenic sprouting into the host tissue of skinfold chambers (dotted lines indicate the graft/host border). (e) Treatment with rAAVβ4 leads to an increase of spontaneous contractions in implanted EHTs compared to rAAVGFP control (n = 4 per group, *p < 0.05).

staining, that the newly formed vasculature had its origin predominantly in the EHTs sprouting to the surrounding tissue of the skinfold chamber (Figure 4(d)). Lastly, to investigate the functionality of the implanted grafts, the amount of contractions was assessed and revealed a dramatic increase in the number of contractions per minute in Tβ4 treated EHTs (Figure 4(e)). Taken together, these findings implicate that the vascularization and performance of EHT can be improved by the treatment of EHTs with Tβ4. As with our results regarding iPSsCs, this effect might be attributed to an increase in the fidelity of neonatal mouse cardiomyocytes during EHT production, possibly due to a decrease in apoptosis and an increase in survival, a mechanism which has been demonstrated in previous work from our group in the context of neonatal rat cardiomyocytes [43].

Combining the findings from our experiments regarding induced pluripotent stem cells and zebrafish, our results implicate that Tβ4 promotes the expansion of mesodermal progenitors, which subsequently differentiate into blood and lymphatic vessels. Those, in turn, increase the perfusion and maintenance of our EHTs. Tβ4 additionally might stabilize the cardiomyocyte phenotype of neonatal cardiomyocytes during EHT-generation by promoting the expression of cardiac proteins. Another possible mechanism, by which Tβ4 enhances EHT functionality, might be by generally promoting proliferation and inhibiting apoptosis, rendering EHTs more stable during their assembly and subsequent implantation.

4. Conclusion

Here we demonstrate that Tβ4 significantly increases the amount of cardiomyocyte-like cells differentiated from
induced pluripotent stem cells and their respective progenitors (mesodermal cells, cardiogenic progenitors, and cardiac precursors) and that Tβ4 is required for proper embryonic development of lymphatic vessels. Furthermore, Tβ4 improves vascularization and beating capacity of EHT generated from neonatal heart cells, possibly by promoting overall proliferation, protecting from apoptosis, and inducing angiogenesis and lymphangiogenesis. These results introduce Tβ4 as a potential driver of iPSC-differentiation and EHT vascularization, improving the efficiency of generating EHTs for further preclinical and clinical use.

Competing Interests

The authors declare that they have no competing interests.

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