Net ecosystem fluxes and composition of biogenic volatile organic compounds over a maize field–interaction of meteorology and phenological stages

FELIX WIEß1,*, ANDREA GHIRARDO2,*, JÖRG-PETER SCHNITZLER2,†, CLAAS NENDEL3, JÜRGEN AUGUSTIN4,§, MATHIAS HOFFMANN5 and RÜDIGER GROTE1

1Karlsruhe Institute of Technology, Institute of Meteorology and Climate Research – Atmospheric Environmental Research (KIT/IMK-IFU), Garmisch-Partenkirchen, Germany, 2Helmholtz Zentrum München, Institute of Biochemical Plant Pathology, Research Unit Environmental Simulation (EUS), Neuherberg, Germany, 3Leibniz Centre for Agricultural Landscape Research (ZALF), Institute of Landscape Systems Analysis, Müncheberg, Germany, 4Leibniz Centre for Agricultural Landscape Research (ZALF), Institute of Soil Landscape Research, Müncheberg, Germany, 5Leibniz Centre for Agricultural Landscape Research (ZALF), Institute of Landscape Biogeochemistry, Müncheberg, Germany

Abstract

Bioenergy crop production is rapidly expanding in Europe, and the potential emissions of biogenic volatile organic compounds (BVOCs) might change the chemical composition of the atmosphere, influencing in turn air quality and regional climate. The environmental impacts of bioenergy crops on air chemistry are difficult to assess due to a lack of accurate field observations. Therefore, we studied BVOC fluxes from a bioenergy maize field in North-Eastern Germany throughout the entire reproductive growth stage of the plants. Combining automated large chambers and proton transfer reaction mass spectrometry (PTR-MS), we successfully measured fluxes of the highly reactive hydrocarbons monoterpenes (MTs) and sesquiterpenes (SQTs), together with several other BVOCs, including alcohols, aldehydes, ketones, benzenoids, and fatty acid derivatives. Emissions of MTs and SQTs were relatively high (17.0% and 3.6% of total mean molar BVOC emission, respectively) compared to methanol emissions (17.6%). Seasonal MT and SQT fluxes were clearly associated with the flowering phase, originating mainly from the flowering tissues as shown in additional laboratory experiments. From the observations of CO2 net ecosystem exchange and evapotranspiration rates, we could exclude heat and drought stress-induced BVOC emissions. Standard emission factors calculated for all compounds, chemical groups, and growth stages, showed that the temperature dependency of volatile terpenoid fluxes decreased distinctively with proceeding development stage. The results indicate that emissions from large-scale bioenergy maize fields should be better differentiated and considered in regional estimates of aerosol formation. For the implementation of such relation into biogeochemical modelling, it should be considered that not only seasonal weather development but also phenological growth stages are determining the BVOC patterns and emission potentials.

Keywords: biogenic emission, biogenic volatile organic compounds, emission factor, large-chamber measurements, phenology, seasonality, sesquiterpenes, Zea mays

Introduction

Biogenic volatile organic compounds (BVOCs) play a pivotal role for air chemistry processes and the formation of secondary organic aerosols (SOA). They increase CH4 life time by removing hydroxyl radicals, participate in the formation of SOA, and are essential for photochemical ozone production (Atkinson & Arey, 2003; Makkonen et al., 2012; Calfapietra et al., 2013; Schultz et al., 2015). Thus, BVOCs affect air quality and (by affecting radiation extinction and cloud formation) also regional climate. BVOC emissions strongly depend on plant species and on environmental conditions that are all subject to change, in particular temperature, radiation, and water availability (Kesselmeier & Staudt, 1999). Thus, the effects of a changed BVOCs emission regime related to major land-use changes should not be neglected (Rosenkranz et al., 2015).

In many parts of the world, especially in the tropics, bioenergy production is currently one of the largest drivers of land-use change (Rosenkranz et al.,...
Biomass is already the main contributor to total primary renewable energy supply. For Germany, this share is about 57% (in 2015), while it is even higher for whole Europe (62% in 2012) or at the global scale (72% in 2013) (European Union, 2014; International Energy Agency, 2015; Fachagentur für Nachwachsende Rohstoffe, 2016). Until 2050, projections for the EU show a further increase of biomass and waste usage for total primary energy production from currently 136 Mt oil equivalents to 164 Mt (Capros et al., 2014). Also Germany has set goals to increase the fraction of renewable energy sources from currently 13% up to 50% using 26% of total primary energy supply from biomass (Erneuerbare-Energien-Gesetz, 2014; Arbeitsgemeinschaft Energiebilanzen, 2015). Apart from direct biomass burning, fermentation into biogas is a main energy production pathway, with currently maize as a preferred substrate (Fachagentur für Nachwachsende Rohstoffe, 2016). Maize, along with rapeseed, energy grasses or poplar and willow coppices, is supposed to be the preferred substrate (Fachagentur für Nachwachsende Rohstoffe, 2016). Maize, along with rapeseed, energy grasses or poplar and willow coppices, is supposed to play an important role in future agricultural production of renewable energy (Bentsen & Felby, 2012). This development has raised environmental concerns, in particular related to groundwater pollution (Nikiema et al., 2012), additional greenhouse gas releases (Crutzen et al., 2008), or biodiversity issues (Immerzeel et al., 2014). However, changes in the emission regimes of BVOCs caused by land-use change have received not much attention so far (Rosenkranz et al., 2015).

In the US, the high-emitting giant cane (Arundo donax L.) has been evaluated having a negative impact on air quality when extensively planted (Porter et al., 2012), while a land-use change from natural to energy grasses, which are supposed to be low emitters, may have contrary impacts (Miresmailli et al., 2013). For the majority of bioenergy plants such as rapeseed, however, the impact of BVOCs on air quality is generally seen as smaller (Morrison et al., 2016) or indifferent from other agricultural crops (Renner & Münzenberg, 2003). In Europe, the air quality impacts of a changed BVOCs emission regime have only been considered yet in case of extended short rotation coppices with hybrid poplars (Ashworth et al., 2013; Beltman et al., 2013) which are well-known as high BVOC emitters, but do play only a minor role in current crop cultivation (~1 % of the total agricultural area in Germany). Nonetheless, the few studies addressing BVOC emissions from maize fields show large qualitatively and quantitatively discrepancies, due to different methodologies ranging from field leaf-level flux analysis to concentration gradient measurements and eddy covariance technique. Often, measurements were taken manually at midday for few days, carried out for leaves only or are concentrating on early growth stages (Das et al., 2003; Graus et al., 2013; Leppik et al., 2014). Only Bachy et al. (2016) reported field fluxes over an entire growing season by means of using eddy covariance. Thus, the BVOC fluxes known so far differ strongly, although the most abundant documented fluxes were methanol followed by monoterpenes (MTs), acetone, and green leaf volatiles (GLVs; for more details see Brilli et al., 2011). Remarkably, sesquiterpene (SQT) fluxes have not been described under field conditions so far, probably due to technical and methodological difficulties, although these highly reactive compounds are assumed playing an important role in air chemistry regarding aerosol formation (Bonn & Moortgat, 2003).

In this study, we analysed changes of BVOC emissions from a maize field as a function of environmental cofactors, phenological stages, and plant status (potential heat and drought stress effects). To achieve this, we applied large automatic chamber systems, initially used to capture net ecosystem exchange (NEE) of CO2 and evapotranspiration (Hoffmann et al., 2016). These systems allow measuring concentration differences of air chemical compounds under almost all weather conditions and the simultaneously derived carbon and water fluxes can be used to evaluate plant stress. In contrast to eddy systems, large chambers capture fluxes for a specific plot only enabling to define precise boundary conditions without advective influences. We used these chambers for the first time in combination with proton transfer reaction quadrupole mass spectrometry (PTR-QMS) and infrared spectroscopy. In addition, we performed laboratory and greenhouse measurements determining tissue-specific BVOC emission pattern under controlled environmental conditions.

Overall we aimed to (i) examine the BVOC composition and fluxes throughout the major part of maize development, (ii) determine tissue-specific emissions, and (iii) evaluate the impact of environmental and phenological conditions on the source strength of BVOC emissions. Based on these data we calculated standard emission factors for the different plant growth stages, as prerequisite for accurate BVOC emission modelling of bioenergy maize fields.

**Materials and methods**

**Description of the field site**

The field study was conducted from the end of July until mid-September 2015 at the CarboZALF-D experimental site located at a ground moraine landscape near Dedelow, NE Germany (N 53.3793, E 13.7856) (Sommer et al., 2016). Maize (Zea mays cv. Zoey, an energy and food cultivar from Advanta Seeds DMCC) was seeded at the end of April 2015 and harvested at 14
September 2015. The 690 m² parcel with 11 maize plants per m² and a row width of 0.75 m was neighboured by winter wheat and grassland. The site is equipped with instrumentation for greenhouse gas (GHG) flux analysis (i.e. CO₂, CH₄, and H₂O; Los Gatos Fast Greenhouse Gas Analyser; 9110010, Los Gatos), meteorological parameters (temperature, radiation, wind, relative humidity, atmospheric pressure) and soil temperature. The leaf area index (LAI) was measured at 5 cm height, six times for each of three directions to get one data point (SunScan Canopy Analysis Systm SS1-BF5, Delta-T, Cambridge, UK). LAI and aboveground biomass were determined at the beginning, mid, and end of the measurement period. The phenological growth stages of the maize plants were recorded each week and reported according to the Biologische Bundesanstalt, Bundessortenamt and Chemical industry (BBCH) identification keys (Meier, 2001).

Measurements of gas exchange and BVOCs in the field

BVOC fluxes were measured from 24 July to 15 September, including one day after the maize was harvested. The air was sampled from a closed system (flow-through non-steady-state) which consisted of two transparent automatic polycarbonate (thickness of 2 mm; light transmission ~70%) chambers of 1.5 m x 1.5 m x 2.5 m size each, with about 20 maize plants growing in each chamber. Chambers were built more than six months prior to the measurements to minimize sources of additional VOCs from the chamber material. A 3/2 way polytetrfluoretylen (PTFE) valve for each automatic chamber was switched between drawing air from the closed chamber to the BVOC measuring device and drawing air when the chamber was opened for flushing the tube. Around 10 L min⁻¹ air flow was drawn from both systems at the same time and, except for the ~70 mL min⁻¹ used for BVOC analysis, the air was redirected back into the chambers. Two fans per chamber ensured homogeneous air mixing. To minimize the impact of the chambers on the plant growth and to ensure ambient environmental conditions, the chambers were kept open for 80% of the time (i.e. 48 min h⁻¹). The median change rates and its standard deviation during a closed chamber period (i.e., within 12 min) were 0.02 and 1.43 K for the chamber temperature, 6.28 and 79.20 ppmv for CO₂, and 1750 and 6078 ppmv for H₂O, respectively.

BVOC concentrations were measured online using high-sensitivity PTR-QMS (Ionicon Analytik GmbH, Innsbruck, Austria) (Lindinger et al., 1998). Operation of the instrument and calculation of BVOC concentration have been previously described (Ghirardo et al., 2010, 2011; Kreuzwieser et al., 2014). For this study, we set the drift tube at pressure of 2.2 mbar, a voltage of 600 V and a temperature of 60 °C, resulting in an E/N of 134 Td (E = the electric field strength, N = the gas number density; 1 Td = 10⁻¹⁷ V cm⁻²). The ions from the first water cluster (i.e. H₂O(H₂O)⁺), the O₃⁺ and the NO⁺ were kept below 10%, 3%, and 0.2% of the primary hydronium ions (H₃O⁺), respectively.

We measured 18 different BVOCs (see Table 1) throughout approximately one minute. The PTR-QMS was connected to the chambers with approximately 20 m 1/4” PTFE tubes with

<table>
<thead>
<tr>
<th>m/z</th>
<th>Measured compound</th>
<th>Abbreviation</th>
<th>Formula</th>
<th>Dwell time (s)</th>
</tr>
</thead>
<tbody>
<tr>
<td>21</td>
<td>Hydronium ions</td>
<td>-</td>
<td>H₄¹⁸O⁺</td>
<td>0.2</td>
</tr>
<tr>
<td>33</td>
<td>Methanol</td>
<td>-</td>
<td>CH₃O⁺</td>
<td>2</td>
</tr>
<tr>
<td>39</td>
<td>Water cluster</td>
<td>-</td>
<td>H₂O(H¹⁸O)⁺</td>
<td>1</td>
</tr>
<tr>
<td>45</td>
<td>Acetaldehyde</td>
<td>-</td>
<td>C₇H₈O⁺</td>
<td>5</td>
</tr>
<tr>
<td>47</td>
<td>Ethanol</td>
<td>-</td>
<td>C₉H₁₀O⁺</td>
<td>1</td>
</tr>
<tr>
<td>59</td>
<td>Acetone</td>
<td>-</td>
<td>C₆H₈O⁺</td>
<td>1</td>
</tr>
<tr>
<td>69</td>
<td>Fragments of pentanal; Octanal; Nonanal; Decanal</td>
<td>m/z 69</td>
<td>GLV*</td>
<td>C₄H₁₀O⁺</td>
</tr>
<tr>
<td>71</td>
<td>Unspecified mass feature</td>
<td>m/z 71</td>
<td>GLV*</td>
<td>C₄H₁₀O⁺</td>
</tr>
<tr>
<td>99</td>
<td>(E)-2-hexenal; (Z)-3-hexenal</td>
<td>GLV*</td>
<td>C₄H₁₀O⁺</td>
<td>2</td>
</tr>
<tr>
<td>107</td>
<td>Xylenes; Benzene; 1,2,3-trimethyl (fragment)</td>
<td>Xylenes</td>
<td>C₆H₁₁⁺</td>
<td>1</td>
</tr>
<tr>
<td>137</td>
<td>Monoterpenes†</td>
<td>MT</td>
<td>C₁₉H₁₇⁺</td>
<td>5</td>
</tr>
<tr>
<td>151</td>
<td>(E)-4,8-dimethyl-1,3,7-nonatriene</td>
<td>DMNT</td>
<td>C₁₉H₁₉⁺</td>
<td>1</td>
</tr>
<tr>
<td>153</td>
<td>Camphor; (E)-2-caren-4-ol; Methyl salicylate</td>
<td>m/z 153</td>
<td>GLV*</td>
<td>C₁₉H₂₁O⁺</td>
</tr>
<tr>
<td>155</td>
<td>Oxygenated monoterpenes†</td>
<td>oMT</td>
<td>C₁₉H₂₁O⁺</td>
<td>1</td>
</tr>
<tr>
<td>205</td>
<td>Sesquiterpenes†</td>
<td>SQT</td>
<td>C₁₉H₂₅⁺</td>
<td>10</td>
</tr>
<tr>
<td>225</td>
<td>C₁₅H₂₀O₂/C₁₇H₂₀</td>
<td>m/z 225</td>
<td>C₁₅H₂₀O⁺; C₁₇H₂₁⁺</td>
<td>1</td>
</tr>
</tbody>
</table>

*These lipoxygenase (LOX) products, commonly called green leaf volatiles (GLV) are fatty acids derivatives of the octadecanoid pathway (Ghirardo et al., 2011; Portillo-Estrada et al., 2015).
†See Table S1 for the chemical identification.

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were sown three times with a time lag of 4 weeks to obtain three different phenological stages at the time of measurement. According to the BBCH identification keys for maize, these growth stages can be described as leaf development (LD; half of all number of leaves unfolded), as the mid and as the end of the flowering phase (FW_MID and FW_END, respectively). As flowering of the tassel and emerging of stigmata depends on the growth stage of the plant, BVOC analyses from those two compartments were carried out with plants in FW_MID. BVOC collections from roots were only measured in LD and FW_MID, as no significant changes appeared in the development of the root system between FW_MID and FW_END. BVOC emissions were collected and analysed by TD-GC-MS using the same headspace sorptive extraction approach described previously in Weikl et al. (2016).

The different plant tissues were enclosed into transparent glass vials of 2-mL (for roots, flowers, and stigmata) and 10-mL volume (for leaves) with non-polar, PDMS coated stir bars (Twister, film thickness 0.5 mm, Gerstel) and incubated for 65 h under light condition (photosynthetically active radiation (PAR) ~100 μmol m⁻² s⁻¹). The mass of analysed roots was 536 ± 39 mg, of the flower 75 ± 15 mg, of the stigmas 188 ± 15 mg. Leaf discs of 4.9 cm² were punched from a leaf at two of three of the leaf length in respect to the stem but out of the midvein (L_M), which represents the reference leaf among the different growth phases LD, FW_MID and FW_END. To investigate the leaf-age dependency, additional leaf discs were collected from the same growth stage FW_MID, an upper leaf (L_U) which is younger than the reference leaf L_M, a lower leaf (L_L) older than L_M, and from the same leaf of L_M, but at 1/3 of the leaf length (L_M2), therefore, closer to stem. Leaf discs were accommodated at the bottom of the 10 mL vial with 200 μL carbonated water (Aqua Culinaris Medium, Uurstromquelle GmbH, Baruth/Mark, Germany) to continuously provide CO₂ for leaf photosynthesis. Control measurements were carried out exactly in the same way, but in absence of the leaf material and the mass signals recorded (negligible) were used for background correction.

**Calculation of BVOC fluxes**

We calculated the BVOC fluxes as the slope of a linear regression (ordinary least squares), fitted to the concentration measurement points over time obtained during 12 min of sampling from each closed chamber. All fluxes from nonsignificant linear regressions (H0: slope of regression line is zero; P > 0.05; that is, r < 0.60 for a two-tailed t distribution test with 12 data points) were set to zero. We scaled all flux rates from ppb s⁻¹ (dVOC_ppb/dt) to nmol m⁻² ground area s⁻¹ (dVOC_mol/m²/dt) by

\[
\frac{dVOC_{\text{mol/m}^2}}{dt} = \frac{dVOC_{\text{ppb}}}{dt} \times \frac{V_C}{A_C \times V_m}
\]

with \(V_C\) (5.625 m³) is the volume, \(A_C\) (2.25 m²) is the base area of the chamber, and \(V_m\) is the molar volume calculated by the ideal gas law. From the resulting twofold 12-minute mean emission rates per hour, we calculated hourly mean and daily mean emission rates. Positive and negative fluxes indicate net ecosystem emissions and deposition, respectively.

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From hourly mean values (daytime and night-time), we calculated standard emission factors (SEF) indicating species-specific emissions scaled to the defined environmental standard conditions of PAR at 1000 μmol m$^{-2}$ s$^{-1}$ and 30 °C leaf temperature (Guenther et al., 1993). During the field study, we only found 20 data points per compound that complied with these conditions. Thus, similar to Tarvainen et al. (2005) we fitted the SEF with all observed hourly mean values for each compound to the equations proposed by Guenther (1997) (see Eqs (2) and (3)) by curve fitting via the Levenburg–Marquardt algorithm with nonlinear least squares. For all compounds we used the temperature-dependent emission function:

$$E_{obs} = SEF \times C_{TM}$$  

$$C_{TM} = e^{β(T−T_{s})}$$  

with $E_{obs}$ (μmol m$^{-2}$ h$^{-1}$) as the observed emission rate at temperature $T$ (K), $β$ (K$^{-1}$) as an empirical temperature coefficient which was either set to 0.09 (Guenther et al., 1993; Guenther, 1997; Tarvainen et al., 2005) or also statistically fitted, and $T_s$ being the standard temperature (303.15 K).

As some volatiles show strong dependencies on temperature as well as radiation (e.g. isoprene, and in some plant species also MTs (Loreto et al., 2000; Ghirardo et al., 2010)), we additionally applied the following function to determine SEFs:

$$E_{obs} = SEF \times \frac{a_{CL1} L}{\sqrt{1 + z^2 L^2}} = \frac{e^{\left(\frac{C_{T3} (T−T_{TM})}{\sigma}\right)}}{C_{T3} + e^{\left(\frac{C_{T3} (T−T_{TM})}{\sigma}\right)}}$$  

with $L$ (μmol m$^{-2}$ s$^{-1}$) as PAR, $R$ (8.314 J K$^{-1}$ mol$^{-1}$) is the ideal gas constant, and $x$ (0.0027), $C_{T1}$ (1.086), $C_{T2}$ (95 000 J mol$^{-1}$), $C_{T3}$ (230 000 J mol$^{-1}$), $C_{T4}$ (0.961), that is, depending on $z$ to set Eqn (3) at standard light and temperature conditions to the value of 1), and $T_{TM}$ (314 K) are empirical parameters given by Guenther (1997). For $m/z$ 80 and MTs, using Eqn (3) could describe the field observations better than using Eqn (2).

As SEFs may change with growth stages (Monson et al., 2012), we distinguished between the three temporal periods of flowering (FW; 27 July–16 August, $N = 495$), fruit development and ripening (FR; 18 August–13 September, $N = 591$), and the whole season (WS; 27 July–13 September, $N = 1110$). Additionally, we calculated a mean SEF for each group of compounds with same seasonal flux pattern by:

$$gSEF_{gram} = \sum_i SEF_{i,gram} \frac{SEF_{i,mod}}{\sum_i SEF_{i,mod}}$$  

with $gSEF_{gram}$ as the mean group SEF in μg m$^{-2}$ (ground) h$^{-1}$, $SEF_{i,gram}$ as the SEF of each component $i$ within a group in μg m$^{-2}$ h$^{-1}$, $SEF_{i,mod}$ as the SEF of each component $i$ within a group in nmol m$^{-2}$ s$^{-1}$, and $gSEF_{i,mod}$ as the summed group SEF in nmol m$^{-2}$ s$^{-1}$. This equation can be described as the summed up molar fraction of each compound’s SEF to the group SEF multiplied by its mass SEF.

To evaluate the development of BVOC fluxes that originate from cumulative temperature and radiation conditions, we also calculated the seasonal development (SD) based on the respective MEGAN algorithms (Guenther et al., 2006).

$$SD = C_{TM} + f_T C_{T1} \frac{C_{T2}}{C_{T3}}$$  

$$C_{T50} = C_{T72} \frac{e^{(C_{T50}/x_{30})}}{C_{T72} - C_{T71} \times [1 - e^{(C_{T50}/x_{30})}]}$$  

$$x_{30} = \frac{T_{opt} - T}{R}$$

with $f_T$, $C_{T1}$, $C_{T2}$, and $T_{opt}$ being the dynamically calculated modifiers for the emission potential that are calculated in dependence on light and temperature of the previous ten days as described in Guenther et al. (2006). SD is calculated in half-hourly time steps and averaged over the day for presentations. The normalized SD factor is daily SD divided by the largest daily SD obtained in the measured period.

**Calculation of net ecosystem CO₂ fluxes**

Fluxes for the net ecosystem CO₂ exchange (NEE; μmol C m$^{-2}$ s$^{-1}$) were calculated according to the ideal gas law, equivalent to the calculation of the BVOC fluxes (Eqn 1). Data noise that originated from either turbulence or pressure fluctuation caused by chamber deployment or from increasing saturation and canopy microclimate effects was excluded by the application of a death-band of 5% to each measurement (Davidson et al., 2002; Kutzbach et al., 2007; Langensiepen et al., 2012). Multiple data subsets based on a variable moving window with a minimum length of 4 min were generated for each measurement (Hoffmann et al., 2015) and linearly fitted (ordinary least squares; Leiber-Sauheitl et al., 2014; Leifeld et al., 2011; Pohl et al., 2015) to calculate the concentration change with time. Resulting multiple NEE fluxes per measurement (based on the moving window data subsets per measurement) were subsequently scrutinized by the following threshold criteria: (i) range of within-chamber air temperature less than ±1.5 K ($R_{exo}$ and NEE fluxes) and deviation of photosynthetic active radiation (PAR) less than ±20% of the average; (ii) significant regression slope ($P ≤ 0.1$); and (iii) nonsignificant tests ($P > 0.1$) for normality (Lillfor’s adaption of the Kolmogorov–Smirnov test), homoscedasticity (Breusch–Pagan test) and linearity of CO₂ concentration data. Calculated CO₂ fluxes that did not meet all threshold criteria were discarded. In cases of more than one flux per measurement meeting all exclusion criteria, the NEE flux with the steepest slope was chosen.

**Statistical analyses**

Principal component analyses (PCA) were performed using the software package SIMCA-P (v13.0.0.0, Umetrics, Umeå, Sweden) following established procedures (for details, see Ghirardo et al., 2012, 2016; Weikl et al., 2016). The results were validated by full cross-validation and if specified, significant at the 95–99.9% confidence interval. One-way ANOVA was carried out at significance levels of $α < 5\%$, using the software package SIGMAPLOT (v11.0, Systat Software Inc., San Jose, CA, USA) and the PYTHON library SCIPY (v0.14).
Results

Meteorology, plant growth stages, NEE, and evapotranspiration

The months of July, August, and September 2015 were generally drier (−24 mm, −13%) and warmer (+1.3 °C) compared to the long-term mean observations from 1992 to 2015 at the field site in Dedelow. While July was slightly warmer but wetter, August was notably warmer (+3 °C) and drier (−23 mm, −50%) than the long-term mean. September 2015 was generally in line with long-term observations. Based on the meteorological data, we distinguish five phases of measurement periods: (i) late July with low daily mean temperatures (around 15 °C) and frequent precipitation events, (ii) first half of August with oscillating but high temperatures (>20 °C) and mostly dry weather (only few precipitation events with maximum daily sums of 5 mm), (iii) second half of August with slightly cooler temperatures but higher radiation in the first week followed by days with lower radiation, (iv) a short period beginning of September when temperatures sharply rise and decline within few days, and (v) the rest of the measurement campaign with cooler temperatures (12 to 16 °C) and moist conditions. This sectioning of weather data enabled comparing the composition and quantity of BVOC fluxes from the two-week hot period of phase 2 with those from rather long-term average temperature conditions of phase 3 and 5.

During phase 1, maize plants were in the vegetative growth and the inflorescence emerged; in phase 2 the plants were flowering and reached highest NEE and evapotranspiration rates; in phase 3 and 4 the fruit developed and during phase 5 the plants were ripening—according to the phenological growth stages from the BBCH identification keys of maize (Meier, 2001). As the biomass of the maize plants inside the chambers could only be measured at the time of harvesting, the biomass development was determined from plants close to the chambers. Generally, only very low pest infestation by corn borer (*Ostrinia nubilalis*) has been reported during the year (data not shown).

Field BVOC fluxes

The PTR-QMS measured BVOC concentrations within the headspace of a chamber during the period when the chamber was closed (e.g. m/z 153, see Fig. 1). For many compounds, no significant concentration changes during night had been observed indicating neither net ecosystem emission nor net deposition of BVOCs. This further indicates that the production of these compounds was strongly light-dependent. In the morning hours, concentrations of all emitted compounds slowly rise with maximum increases generally appearing...
around early afternoon and then subsequently decreasing again. This diurnal cycle was temperature- and light-dependent, reaching emission maxima around early afternoon and minima in the night or during early morning hours (Fig. 2).

**Absolute fluxes**

Throughout the whole study, the highest daily mean BVOC emissions were observed within the 2nd phase (flowering and beginning of fruit development), when means of temperature and radiation were highest (Fig. 3c–e). Similar environmental conditions of temperature and radiation were observed in phase 4 ($P > 0.05$; ANOVA), but all BVOC emissions except for ethanol ($P = 0.051$) were significantly lower than in phase 2, suggesting that the plant phenology was a key factor in controlling BVOC emissions. We observed rather low BVOC fluxes during the end of the vegetative growth phase as well as during the end of the fruit development until plant ripening. The change in methanol flux from a net ecosystem emission to a net deposition was very likely caused by its high water solubility as it coincides with precipitation events (Niinemets et al., 2004).

Overall, highest molar emissions (>0.20 nmol m$^{-2}$ s$^{-1}$) were measured for GLVs, MTs, methanol, acetone, and acetaldehyde (arranged in decreasing order), with a corresponding mean portion to total molar BVOC emissions of 13.2% to 18.1% for the whole period (see Table 2). Lower molar emissions (<0.10 nmol m$^{-2}$ s$^{-1}$) were measured for $m/z$ 69, $m/z$ 153, SQT, ethanol, and $m/z$ 71 (arranged in decreasing order) with portions < 5.4% for the whole period. Very low but still significant emissions were detected for (E)-4,8-Dimethyl-1,3,7-nonatriene (DMNT), xylenes, oxygenated MTs (oMTs), and a C$_{17}$H$_{30}$/C$_{13}$H$_{20}$O$_{3}$ compound ($m/z$ 225) (not shown). Until harvesting (ripening growth stage) the emissions of most of the compounds gradually decreased to daily mean values below 0.03 nmol m$^{-2}$ s$^{-1}$. At harvesting as well as on the following day, all emissions were below 0.01 nmol m$^{-2}$ s$^{-1}$. We also observed net deposition fluxes of methanol and ethanol and to some degree also of acetaldehyde and acetone. Whereas mean compound emissions are decreasing with growth stage, the partitioning between the compounds is more complex (see Table 2), with some compounds increasing portions in later growth stages (GLVs, acetone, ethanol, xylenes, acetaldehyde, $m/z$ 71, and DMNT).

**Normalized fluxes**

By normalizing daily mean BVOC fluxes to each compound’s daily mean maximum of the whole season, we separated the compounds into three groups with distinct seasonal flux patterns. The first group mainly consists of volatile terpenoids (MTs, oMTs, SQTs) and their derivatives ($m/z$ 153, $m/z$ 225) (see Fig. 4a). The second group summarizes the C$_{1}$-C$_{2}$ alcohols methanol and ethanol (see Fig. 4b). Compounds from the third group are defined as other VOCs (see Fig. 4c) and include GLVs, acetaldehyde, $m/z$ 71, xylenes, acetone, and DMNT, which originates from the terpenoid biosynthetic pathway (Richter et al., 2016).

The volatile terpenoids show high fluxes particularly in phase 2 and then gradually decline. The methanol and ethanol fluxes showed emission and deposition rates with similar amplitudes and generally decreasing

---

**Fig. 2** Diurnal cycle of 10–12 August 2015. (a) Hourly mean temperature and photosynthetically active radiation (PAR); (b) hourly mean BVOC emission rates, with minimum and maximum values as error bars ($N = 2$).

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fluxes in phase 5. The group of other VOCs shows a similar flux pattern as terpenoids in phase 1 and 2 but did not decrease in phase 3. This group also showed a generally stronger oscillation of the emission rates until the end of phase 4. From the relation to the calculated SD development, it can be derived that terpenoid emissions considerably depend on past weather conditions (particularly apparent in phase 3), while the emissions of other VOCs strongly respond to instantaneous radiation and temperature. However, the emission increase in both groups at the start of phase 2 was generally more pronounced than indicated by the SD factor. Moreover, the responsiveness to the increase in temperature in phase 4 was much less than could be assumed from the cumulative weather conditions. Thus, cumulative weather as well as the development stage needs consideration to determine the correct emission pattern.

**Tissue-specific BVOC emissions**

The field measurements indicated that changes in terpenoid emissions could not be explained by environmental factors alone, suggesting that plant ontogenesis plays a crucial role for the pattern of overall plant emissions, especially for the steep increase of most compounds during the flowering phase. To determine the source of these emissions, we performed laboratory analyses under environmentally controlled conditions, measuring emission rates separately from different plant tissues and for different growth stages and leaf ages (Fig. 5b).

Overall, flowering tissues were the strongest source of terpenoids, in particular of oMTs, SQTs, and oxygenated SQTs (oSQTs), with up to 14-fold higher emission rates compared to leaves (see Fig. 5 and Table S1). Noteworthy, the BVOC emission rate of...
Table 2  Mean molar emission in pmol m$^{-2}$ s$^{-1}$ (negative fluxes set to zero) and fraction of each compound to the total BVOC emission in % in brackets for the two phenological growth stages of flowering and the fruit development and ripening as well as for the whole observed growth period. See Table S1 for a further identification of the groups of MTs, oMTs, and SQTs.

<table>
<thead>
<tr>
<th>Compound</th>
<th>Flowering (21 days)</th>
<th>Fruit dev. and ripening (27 days)</th>
<th>Whole period (49 days)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Methanol</td>
<td>137.8 (17.6)</td>
<td>70.23 (16.2)</td>
<td>99.57 (16.8)</td>
</tr>
<tr>
<td>Acetaldehyde</td>
<td>102.12 (13.0)</td>
<td>57.65 (13.3)</td>
<td>78.05 (13.2)</td>
</tr>
<tr>
<td>Ethanol</td>
<td>12.94 (1.7)</td>
<td>16.65 (3.8)</td>
<td>14.98 (2.5)</td>
</tr>
<tr>
<td>Acetone</td>
<td>100.22 (12.8)</td>
<td>72.27 (16.7)</td>
<td>85.55 (14.4)</td>
</tr>
<tr>
<td>m/z 69</td>
<td>47.44 (6.1)</td>
<td>18.85 (4.4)</td>
<td>31.96 (5.4)</td>
</tr>
<tr>
<td>m/z 71</td>
<td>24.63 (3.1)</td>
<td>14.32 (3.3)</td>
<td>19.03 (3.2)</td>
</tr>
<tr>
<td>GLVs</td>
<td>122.5 (15.6)</td>
<td>93.59 (21.6)</td>
<td>107.78 (18.1)</td>
</tr>
<tr>
<td>Xylenes</td>
<td>9.89 (1.3)</td>
<td>9.42 (2.2)</td>
<td>9.73 (1.6)</td>
</tr>
<tr>
<td>MTs</td>
<td>133.46 (17.0)</td>
<td>49.19 (11.4)</td>
<td>87.96 (14.8)</td>
</tr>
<tr>
<td>DMNT</td>
<td>17.29 (2.2)</td>
<td>10.39 (2.4)</td>
<td>13.65 (2.3)</td>
</tr>
<tr>
<td>m/z 153</td>
<td>36.43 (4.7)</td>
<td>9.71 (2.2)</td>
<td>21.9 (3.7)</td>
</tr>
<tr>
<td>oMTs</td>
<td>9.56 (1.2)</td>
<td>4.47 (1.0)</td>
<td>6.84 (1.2)</td>
</tr>
<tr>
<td>SQTs</td>
<td>28.57 (3.6)</td>
<td>5.79 (1.3)</td>
<td>16.29 (2.7)</td>
</tr>
<tr>
<td>m/z 225</td>
<td>0.36 (0.0)</td>
<td>0.05 (0.0)</td>
<td>0.19 (0.0)</td>
</tr>
<tr>
<td>Total</td>
<td>783.19 (100.0)</td>
<td>432.56 (100.0)</td>
<td>593.49 (100.0)</td>
</tr>
</tbody>
</table>

Total sums are indicated in bold.

Changing standard emission factors under changing growth stages

Because it is scaled to specific environmental conditions, the SEF is independent from temperature and radiation, and therefore can be used to examine seasonal and phenological dependencies of BVOC emissions. Accordingly, we calculated SEFs for the different growth stages flowering (FW), fruit development together with ripening (FR), and the whole season (WS) (see Table 3). Highest SEFs (>40 μg m$^{-2}$ h$^{-1}$) for WS were obtained for MT, GLVs, acetone, m/z 153, and SQT (arranged in decreasing order). The mean SEF for all terpenoids (from six BVOCs with 495, 591, and 1110 values each for FW, FR, and WS, respectively) decreases by half from FW to FR (114 to 60 μg m$^{-2}$ h$^{-1}$). In contrast, the mean SEF for all other (seven) VOCs remains approximately constant at an emission rate around 54 μg m$^{-2}$ h$^{-1}$ for all seasons. The different SEF development is reasonably resembled by each specific compound within these two groups, except if an additionally influence of radiation is considered in the calculation of the SEFs. In this case, SEFs for MTs and m/z 69 only differed little between growth phases. The mean SEF value for methanol and ethanol is hardly meaningful as the SEF fit the equation for temperature-dependent emissions with a constant β-coefficient of 0.09 was very poor for both alcohols ($R^2 < 0.2$).

When additionally fitting the β-coefficient, the standard error decreases with increasing $R^2$ for all compounds, meaning that the resulting function better represented the observed emission rates than only fitting the SEF value. For terpenoids and other VOCs this lead to a minimum β-coefficient of 0.131 K$^{-1}$ for acetaldehyde during flowering and a maximum value of 0.203 K$^{-1}$ for m/z 153 for the whole period.

Discussion

In the following, we discuss the changes and dependencies of BVOC fluxes from the investigated maize field throughout an almost entire growth period. In particular, we could show that flowering tissues are important additional sources of volatile terpenes. Nonterpene compounds did not show these phenological and seasonal dependencies but could be explained by instantaneous weather conditions. The application of a new technique that combines large automatic chambers with PTR-QMS technology showed the feasibility of measuring BVOC emissions at the field scale continuously over extended time periods. It also demonstrated that the total net carbon loss from all observed BVOCs during the field study (0.13 gC m$^{-2}$) was quite small in relation to the net carbon assimilation (561.1 gC m$^{-2}$). However,
the emission of highly reactive MTs and especially SQTs in significant amounts indicates that maize fields are likely to play an important role in regional aerosol formation and should thus be considered in regional biosphere-atmosphere feedback analysis, particularly regarding land-use change scenarios that evaluate bioenergy production impacts.

Seasonality and phenology impact

The BVOC fluxes from the maize canopy were highly influenced by immediate temperature and radiation conditions. However, we also demonstrate an important impact of plant ontogenesis which is apparent from: (i) the very steep increase in MT, m/z 153, and SQT emissions during the flowering period (transition from meteorological phase 1 to 2) that could not be fully explained by current environmental conditions, (ii) the short period of temperature increase in late August/ beginning of September (phase 4) which is not followed by an emission increase that would be expected from the early season responses (see Fig. 3), and (iii) the dramatic change in SEF of the mean of terpenoids and their derivatives (MTs, oMTs, m/z 153, SQTs, m/z 225), calculated for the flowering phase and fruit development/ripening.

Seasonality, defined as long-term (cumulative) environmental conditions, explained and described some, but not all, of the temporal dynamics of the BVOC fluxes. Seasonal changes of MT and SQT emissions reflect changes of transcript, protein levels, availability of substrates for their biosynthesis, and enzyme activities involved in their biosynthesis (Fischbach et al., 2002; Mayrhofer et al., 2005; Ghirardo et al., 2014; Wright et al., 2014). While the cellular biochemistry is affected by the past weather conditions (Monson et al., 2012), we could neither explain the large terpenoid emissions observed during the flowering phase, nor the sensitivity of other VOCs to high radiation phases during the ripening stage by means of the seasonal development factor.

One possible explanation is the presence of other influences that enhance BVOC emissions during the
flowering period. Our laboratory results indicate that flowering tissues are a strong source of terpenoids. This observation agrees well with previous studies on maize showing the tissue- and developmental stage-specific expression of terpene synthase genes in maize (Köllner et al., 2004a,b). In accordance with the observations of Köllner and colleagues, we argue that terpenoids originating from the husks of the maize kernels further contributed significantly to the high observed fluxes of MTs and SQTs.

By applying the two different methods of normalizing and calculating SEFs for different growth periods, we could differentiate the compounds according by their temporal flux pattern into three groups: (i) terpenoids (MT, oMT, SQT) and their derivatives (m/z 153, m/z 225) with strongly differing SEF between the growth stages and decreasing normalized emission rates from flowering to fruit development; (ii) methanol and ethanol showing an irregular SEF as well as emission and deposition flux pattern with strong relations to humidity; and (iii) other VOCs (fatty acids derivatives, ketones, aldehydes, and aromatics) with similar SEFs in all plant growth stages and decreasing normalized emission rates. Whereas the dynamics of terpenoids are explained by seasonal weather development and modified by phenological stages, the emissions of other VOCs strictly follow instantaneous temperature and radiation conditions. Moreover, the net fluxes for methanol and ethanol are difficult to reproduce without considering deposition fluxes that depend on air humidity or the occurrence of water on leaf surfaces. As methanol and ethanol fluxes changed from net

Fig. 5 Terpenoid emission rates from: (a) the plant tissues stigmata (S), male flower (F), leaf (L), and root (R); (c) young (L_U; upper leaf), mature (L_M1, L_M2), and old (L_L; lower leaf) leaves; (d) the phenological stage leaf development (L_D), mid (FW_MID) and end (FW_END) of the flowering phase (means of N = 4 ± standard error). The origin of the samples is schematically depicted in (b). Inserts: Principal component analysis (PCA) of the respective sample set. Compounds: 1 limonene; 2 1,8-cineole; 3 (E)-4,8-dimethyl-1,3,7-nona-triene; 4 camphor; 5 (E)-2-caren-4-ol; 6 decanal; 7 4-ethylguaiacol; 8 α-ylangene; 9 cyclosativene; 10 β-cubebene; 11 β-caryophyllene; 12 (E)-x-bergamotene; 13 (E)-β-farnesene; 14 α-humulene; 15 β-bisabolene; 16 γ-cadinene; 17 β-sesquiphellandrene; 18 (E)-nerolidol; 19 β-cadinol. Further information can be found in Table S1.
Table 3  Fitted standard emission factors (SEF) with corresponding standard errors of the estimate (SE) both in µg m$^{-2}$ h$^{-1}$, fitted and standard value of the empirical β-coefficient from the equation for temperature-dependent emissions in K$^{-1}$ (see Guenther, 1997) and coefficient of determination ($R^2$). SEF values are also calculated as a mean value for the three compound groups: terpenes, methanol, and ethanol, and other VOCs. All values are calculated for three growth periods flowering (FW, N = 495), fruit development and ripening (FR, N = 591), and the whole season (WS, N = 1110).

<table>
<thead>
<tr>
<th>Compound</th>
<th>Period</th>
<th>SEF (± SE)</th>
<th>β</th>
<th>$R^2$</th>
<th>SEF (± SE)</th>
<th>β</th>
<th>$R^2$</th>
</tr>
</thead>
<tbody>
<tr>
<td>m/z 69*</td>
<td>FW</td>
<td>39.24 (± 5.45)</td>
<td>-</td>
<td>0.82</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>FR</td>
<td>30.49 (± 2.96)</td>
<td>-</td>
<td>0.77</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>WS</td>
<td>36.73 (± 4.47)</td>
<td>-</td>
<td>0.82</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>MTs*</td>
<td>FW</td>
<td>254.2 (± 35.16)</td>
<td>-</td>
<td>0.85</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>FR</td>
<td>184.84 (± 15.02)</td>
<td>-</td>
<td>0.85</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>WS</td>
<td>235.36 (± 27.93)</td>
<td>-</td>
<td>0.85</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>m/z 69</td>
<td>FW</td>
<td>29.72 (± 6.52)</td>
<td>0.09</td>
<td>0.75</td>
<td>33.78 (± 4.98)</td>
<td>0.135</td>
<td>0.85</td>
</tr>
<tr>
<td></td>
<td>FR</td>
<td>16.79 (± 4.23)</td>
<td>0.09</td>
<td>0.53</td>
<td>31.21 (± 3.31)</td>
<td>0.171</td>
<td>0.71</td>
</tr>
<tr>
<td></td>
<td>WS</td>
<td>24.67 (± 6.15)</td>
<td>0.09</td>
<td>0.65</td>
<td>32.3 (± 4.41)</td>
<td>0.152</td>
<td>0.82</td>
</tr>
<tr>
<td>MTs</td>
<td>FW</td>
<td>177.74 (± 58.32)</td>
<td>0.09</td>
<td>0.6</td>
<td>207.09 (± 48.91)</td>
<td>0.146</td>
<td>0.72</td>
</tr>
<tr>
<td></td>
<td>FR</td>
<td>93.85 (± 28.12)</td>
<td>0.09</td>
<td>0.47</td>
<td>202.34 (± 20.66)</td>
<td>0.191</td>
<td>0.72</td>
</tr>
<tr>
<td></td>
<td>WS</td>
<td>145.43 (± 48.51)</td>
<td>0.09</td>
<td>0.54</td>
<td>198.66 (± 37.47)</td>
<td>0.162</td>
<td>0.72</td>
</tr>
<tr>
<td>m/z 153</td>
<td>FW</td>
<td>59.77 (± 23.07)</td>
<td>0.09</td>
<td>0.55</td>
<td>73.57 (± 15.89)</td>
<td>0.183</td>
<td>0.79</td>
</tr>
<tr>
<td></td>
<td>FR</td>
<td>20.35 (± 5.37)</td>
<td>0.09</td>
<td>0.53</td>
<td>44.05 (± 3.52)</td>
<td>0.193</td>
<td>0.8</td>
</tr>
<tr>
<td></td>
<td>WS</td>
<td>44.44 (± 18.27)</td>
<td>0.09</td>
<td>0.46</td>
<td>67.28 (± 11.69)</td>
<td>0.203</td>
<td>0.78</td>
</tr>
<tr>
<td>oMTs</td>
<td>FW</td>
<td>15.67 (± 5.33)</td>
<td>0.09</td>
<td>0.61</td>
<td>19.11 (± 3.49)</td>
<td>0.174</td>
<td>0.83</td>
</tr>
<tr>
<td></td>
<td>FR</td>
<td>9.95 (± 3.3)</td>
<td>0.09</td>
<td>0.44</td>
<td>22.43 (± 2.47)</td>
<td>0.197</td>
<td>0.68</td>
</tr>
<tr>
<td></td>
<td>WS</td>
<td>13.47 (± 4.54)</td>
<td>0.09</td>
<td>0.55</td>
<td>19.25 (± 3.01)</td>
<td>0.177</td>
<td>0.8</td>
</tr>
<tr>
<td>SQTs</td>
<td>FW</td>
<td>57.28 (± 16.61)</td>
<td>0.09</td>
<td>0.66</td>
<td>68.41 (± 10.81)</td>
<td>0.166</td>
<td>0.86</td>
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<td>16.66 (± 4.72)</td>
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<td>0.5</td>
<td>36.61 (± 3.22)</td>
<td>0.195</td>
<td>0.77</td>
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<tr>
<td></td>
<td>WS</td>
<td>41.86 (± 14.78)</td>
<td>0.09</td>
<td>0.52</td>
<td>61.46 (± 8.99)</td>
<td>0.193</td>
<td>0.82</td>
</tr>
<tr>
<td>m/z 225</td>
<td>FW</td>
<td>0.82 (± 0.29)</td>
<td>0.09</td>
<td>0.58</td>
<td>0.99 (± 0.22)</td>
<td>0.165</td>
<td>0.76</td>
</tr>
<tr>
<td></td>
<td>FR</td>
<td>0.14 (± 0.07)</td>
<td>0.09</td>
<td>0.22</td>
<td>0.27 (± 0.07)</td>
<td>0.17</td>
<td>0.3</td>
</tr>
<tr>
<td></td>
<td>WS</td>
<td>0.56 (± 0.25)</td>
<td>0.09</td>
<td>0.41</td>
<td>0.84 (± 0.18)</td>
<td>0.201</td>
<td>0.68</td>
</tr>
<tr>
<td>Mean</td>
<td>FW</td>
<td>113.52</td>
<td>0.09</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Terpenoids†</td>
<td>FR</td>
<td>60.47</td>
<td>0.09</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>WS</td>
<td>92.7</td>
<td>0.09</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
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<tr>
<td>Methanol</td>
<td>FW</td>
<td>32.65 (± 34.35)</td>
<td>0.09</td>
<td>0.2</td>
<td>45.88 (± 30.46)</td>
<td>0.215</td>
<td>0.37</td>
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<tr>
<td></td>
<td>FR</td>
<td>18.45 (± 19.5)</td>
<td>0.09</td>
<td>0.1</td>
<td>53.32 (± 18.4)</td>
<td>0.218</td>
<td>0.2</td>
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<tr>
<td></td>
<td>WS</td>
<td>25.14 (± 28.5)</td>
<td>0.09</td>
<td>0.14</td>
<td>44.96 (± 25.71)</td>
<td>0.221</td>
<td>0.3</td>
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<tr>
<td>Ethanol</td>
<td>FW</td>
<td>3.14 (± 7.01)</td>
<td>0.09</td>
<td>0.05</td>
<td>6.07 (± 6.1)</td>
<td>0.293</td>
<td>0.28</td>
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<tr>
<td></td>
<td>FR</td>
<td>5.38 (± 9.42)</td>
<td>0.09</td>
<td>0.05</td>
<td>18.82 (± 9.02)</td>
<td>0.215</td>
<td>0.13</td>
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<tr>
<td></td>
<td>WS</td>
<td>3.75 (± 8.45)</td>
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<td>0.04</td>
<td>8.18 (± 8.04)</td>
<td>0.23</td>
<td>0.13</td>
</tr>
<tr>
<td>Mean</td>
<td>FW</td>
<td>30.8</td>
<td>0.09</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
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<tr>
<td>Methanol and Ethanol</td>
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<td>16.25</td>
<td>0.09</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>WS</td>
<td>23.13</td>
<td>0.09</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Acetaldehyde</td>
<td>FW</td>
<td>41.05 (± 8.98)</td>
<td>0.09</td>
<td>0.71</td>
<td>46.22 (± 8.31)</td>
<td>0.131</td>
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<tr>
<td></td>
<td>FR</td>
<td>31.65 (± 10.26)</td>
<td>0.09</td>
<td>0.42</td>
<td>57.76 (± 8.92)</td>
<td>0.167</td>
<td>0.56</td>
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<tr>
<td></td>
<td>WS</td>
<td>37.19 (± 10.37)</td>
<td>0.09</td>
<td>0.6</td>
<td>46.78 (± 8.82)</td>
<td>0.14</td>
<td>0.71</td>
</tr>
<tr>
<td>Acetone</td>
<td>FW</td>
<td>57.29 (± 16.79)</td>
<td>0.09</td>
<td>0.66</td>
<td>67.37 (± 12.68)</td>
<td>0.152</td>
<td>0.8</td>
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<tr>
<td></td>
<td>FR</td>
<td>55.1 (± 15.44)</td>
<td>0.09</td>
<td>0.49</td>
<td>104.28 (± 12.45)</td>
<td>0.173</td>
<td>0.67</td>
</tr>
<tr>
<td></td>
<td>WS</td>
<td>56.23 (± 16.16)</td>
<td>0.09</td>
<td>0.59</td>
<td>72.45 (± 13.27)</td>
<td>0.146</td>
<td>0.72</td>
</tr>
<tr>
<td>m/z 71</td>
<td>FW</td>
<td>17.45 (± 6.82)</td>
<td>0.09</td>
<td>0.56</td>
<td>21.55 (± 4.69)</td>
<td>0.183</td>
<td>0.79</td>
</tr>
<tr>
<td></td>
<td>FR</td>
<td>12.38 (± 5.66)</td>
<td>0.09</td>
<td>0.29</td>
<td>26.16 (± 5.04)</td>
<td>0.186</td>
<td>0.44</td>
</tr>
<tr>
<td></td>
<td>WS</td>
<td>15.3 (± 6.39)</td>
<td>0.09</td>
<td>0.45</td>
<td>22.07 (± 4.99)</td>
<td>0.179</td>
<td>0.67</td>
</tr>
<tr>
<td>Hexenal</td>
<td>FW</td>
<td>36.47 (± 12.53)</td>
<td>0.09</td>
<td>0.61</td>
<td>44.7 (± 7.96)</td>
<td>0.176</td>
<td>0.84</td>
</tr>
<tr>
<td></td>
<td>FR</td>
<td>35.87 (± 12.52)</td>
<td>0.09</td>
<td>0.4</td>
<td>71.89 (± 10.58)</td>
<td>0.18</td>
<td>0.57</td>
</tr>
<tr>
<td></td>
<td>WS</td>
<td>36.13 (± 12.57)</td>
<td>0.09</td>
<td>0.52</td>
<td>49.16 (± 10.06)</td>
<td>0.161</td>
<td>0.69</td>
</tr>
<tr>
<td>Hexanal</td>
<td>FW</td>
<td>93.7 (± 34.41)</td>
<td>0.09</td>
<td>0.58</td>
<td>114.74 (± 24.26)</td>
<td>0.171</td>
<td>0.79</td>
</tr>
</tbody>
</table>

(continued)
ecosystem emission to net deposition during or after precipitation events, it is very likely that this behaviour is affected by the high water solubility of the compounds (Niinemets et al., 2004). In dry periods, however, methanol emissions seemed to develop similar to those of other VOCs, indicating a temperature dependency related to biological processes and their biosynthesis as has been recently pointed out by Mozaffar et al. (2017). Whereas ethanol emissions are the results of alcoholic sugar fermentation generally originating from roots (Kreuzwieser et al., 1999).

We also found a strong similarity ($R^2 = 0.94$) between normalized fluxes from $m/z$ 69 and MTs. The TD-GC-MS analysis could not identify $m/z$ 69 as isoprene, but traces of pentanal, octanal, nonanal, and decanal could be detected. These fatty aldehydes, which are typical natural products used in fragrance and flavour production, all fragment into $m/z$ 69 as isoprene, but those of other VOCs, indicating a temperature dependency related to biological processes and their biosynthesis (Niinemets et al., 2004). In dry periods, however, methanol emissions seemed to develop similar to those of other VOCs, indicating a temperature dependency related to biological processes and their biosynthesis (Niinemets et al., 2004). In dry periods, however, methanol emissions seemed to develop similar to those of other VOCs, indicating a temperature dependency related to biological processes and their biosynthesis (Niinemets et al., 2004). In dry periods, however, methanol emissions seemed to develop similar to those of other VOCs, indicating a temperature dependency related to biological processes and their biosynthesis (Niinemets et al., 2004).

Comparison with other BVOC emission studies on maize

So far only a few studies have analysed BVOC fluxes from maize canopies. These include some investigation with GC-MS analyses during a field campaign in Germany (Wedel et al., 1998) and some short-term investigations by Das et al. (2003) and Graus et al. (2013). The only extensive field study was performed by Bachy et al. (2016) followed up by the recent work of Mozaffar et al. (2017) who measured two cultivars for each phenological developmental stage. Generally, all studies have in common that BVOC emissions showed a large diurnal and daily variability similar to the present work. However, the distinct compound classes, in particular the group of SQTs has neither been depicted nor analysed before.

It should be noted, however, that comparison of SEFs depends on the collection of BVOC data as well as the parameter set for each phenological developmental stage. Generally, all studies have in common that BVOC emissions showed a large diurnal and daily variability similar to the present work. However, the distinct compound classes, in particular the group of SQTs has neither been depicted nor analysed before.

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during warm and dry weather periods alone, because the importance of deposition processes during wet periods has been neglected.

To compare our results with those from the different studies, we are concentrating on the flowering period and calculate all emission data as $\mu g \text{ m}^{-2} \text{ g}^{-1}$. We found that the most abundant emissions from this period were methanol, MTs, GLVs, acetaldehyde, and acetone ranging from 13\% to 18\% of the total molar BVOC fluxes. In absolute terms, the mean methanol emission rate is 4 $\mu g \text{ m}^{-2} \text{ h}^{-1}$ (mean of positive and negative fluxes), which is considerably smaller than what has been found by Bachy et al. (2016) ($\sim 50$ $\mu g \text{ m}^{-2} \text{ h}^{-1}$), Graus et al. (2013) (400 $\mu g \text{ m}^{-2} \text{ h}^{-1}$), or Das et al. (2003) (3450 $\mu g \text{ m}^{-2} \text{ h}^{-1}$). Also MT and acetone emissions (65 and 21 $\mu g \text{ m}^{-2} \text{ h}^{-1}$, respectively) are more than one magnitude lower compared to Das et al. (2003) (661 and 425 $\mu g \text{ m}^{-2} \text{ h}^{-1}$, respectively) but are higher than has been reported by Graus et al. (2013) ($\sim 50$ $\mu g \text{ m}^{-2} \text{ h}^{-1}$) and Bachy et al. (2016) ($\sim 2$ $\mu g \text{ m}^{-2} \text{ h}^{-1}$). GLVs were only detected by Graus et al. (2013) and Wedel et al. (1998). In summary, we observed higher emission rates than obtained by Bachy et al. (2016), for all compounds except for methanol, while they are general lower when compared to the results from Das et al. (2003) and Graus et al. (2013). However, it should be considered that at least the latter work reported only midday fluxes and fluxes under environmental standard conditions.

The SEF for MTs for the whole season is 145 $\mu g \text{ m}^{-2} \text{ h}^{-1}$ if we assume temperature dependence only and 235 $\mu g \text{ m}^{-2} \text{ h}^{-1}$ if the flux is assumed to be temperature and radiation dependent (see Table 3). Both values are of similar magnitude compared to other bioenergy crop systems such as oil palm (SEF of 102–113 $\mu g \text{ m}^{-2} \text{ h}^{-1}$) and eucalyptus (200–600 $\mu g \text{ m}^{-2} \text{ h}^{-1}$) (Rosenkranz et al., 2015). Additionally, we observed a whole array of substances including significant emissions of SQTs for which flux rates over maize fields were previously not detected. SQTs are considered to be a very important precursor for SOA, already at relatively low concentrations (Bonn & Moortgat, 2003; Mentel et al., 2013). Given the much higher abundance of maize fields than short rotation coppices in most parts of the world, the present data highlights the importance for estimates from agricultural sides to assess regional air quality and regional climate.

Overall, the investigation adds to the large variability found in different studies, which can be only partially explained with methodological differences. As maize fields are carried out with a lot of different cultivars, differences in genotypes may also explain part of the variation (Gouinguené et al., 2001), underlining the need for a systematic screening of cultivars.

Suitability of automatic chamber measurements

Using large automatic chambers for measuring maize canopy BVOC fluxes have shown several advantages. First, the closed chamber method enabled measuring absolute fluxes of many volatiles including SQTs, $m/z$ 153, and oMTs, which were not observed or at least not quantified in previous maize field experiments. In particular, the volatile SQTs were hardly detectable at canopy scale (Ciccioli et al., 1999; Jardine et al., 2011), although they are supposed to play an important role in aerosol formation (e.g. Hallquist et al., 2009; Li et al., 2011). The investigation thus confirms previous genetic and physiological investigation on tissue level which already indicated that both SQTs and oMTs are potentially emitted by maize in considerable quantities (Schnee et al., 2002, 2006; Köllner et al., 2004a,b; Fontana et al., 2011). A second advantage of the present chamber approach is the fact that a defined ecosystem patch is measured, which on the one hand can be assumed as representative and on the other can be easily defined (in terms of number of individuals, height, LAI, phenological stage).

Nevertheless, a number of precautions need to be carefully taken into account to avoid erroneous data collection and reduce uncertainties. In particular at days with strong radiation, the ‘greenhouse effect’ within the chamber may lead to rising temperatures and the strong assimilation decreases CO$_2$ concentrations and increases relative humidity, which both potentially affect BVOC emissions and deposition. For security reasons, chambers open automatically when a temperature threshold of about 40 $^\circ$C is exceeded, which has not been reached during the investigations (observed maximum of 38.7 $^\circ$C). Within this limit, we argue that not only temperature changes but also variations in CO$_2$ concentration are not affecting the de novo synthesis of BVOCs within 12 min (i.e. the period of chamber closure) (Noe et al., 2010). Thus, only emissions from storages that do not directly originate from de novo production but depend on storage size and evaporation resistance could still be influenced (Ghirardo et al., 2010; Peñuelas & Staude, 2010), which we checked using laboratory experiments. These exercises showed no BVOC emission increase within 12 min for changes in temperature and CO$_2$ concentrations that were similar to the highest ones observed during the field campaign (data not shown). As we could not detect strong nonlinearities of BVOC concentration changes within the chambers, we assume that the share of storage-based emission is either small (in agreement with Köllner et al., 2004a) or that the change in headspace temperature was unable to increase tissue temperature during chamber closure.

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Another source of uncertainty is the possible reaction of BVOCs with atmospheric NOx, OH, and O3 inside the chamber, which may occur particularly if radiation and temperature are high. We have no indication about OH (pptv) concentrations but assume that they are generally low at the remote site, which is not much affected by anthropogenic emissions. The remaining reactivity depends on the relation of BVOCs to NO and O3 concentrations. NO originates mainly from soil emissions (Pilegaard, 2013) which are continuously fuelling air concentrations and thus are likely to be relatively high in comparison with background levels of O3 (usually 10–40 ppb according to Atkinson & Arey, 2003). Therefore, chemical reactions of BVOCs are unlikely because NO molecules will react with O3 quite fast producing NO2 and depleting O3 quickly. This is supported by the linear increase of BVOC emissions, indicating no or minor secondary reactions between BVOC and inorganic components of the atmosphere. Thus, the simple linear regression approach seems suitable to retrieve reliable trace gas exchange rates for BVOCs.

Despite some uncertainties that remain to be investigated in future studies, the overall approach is likely to be applicable to other crop species as well. As the chambers can be temporarily removed from the ground, different kinds of management can also be applied and investigated. Together, this makes the method potentially interesting for comparative studies in preparation for extrapolating the results to the regional scale.

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Competing financial interests

The authors declare no competing financial interests exist.

References


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Supporting Information

Additional Supporting Information may be found online in the supporting information tab for this article:

Table S1. Chemical identification of BVOCs and calculated Kovats’ retention index.

Figure S1. Principal component analysis of all BVOC data from different maize tissues collected under controlled laboratory conditions.