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Hydraulic Water Redistribution by Silver Fir (*Abies alba* Mill.) Occurring under Severe Soil Drought

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Received: 20 December 2019; Accepted: 29 January 2020; Published: 31 January 2020



Abstract: Hydraulic redistribution (HR) of water from wet- to dry-soil zones is suggested as an important process in the resilience of forest ecosystems to drought stress in semiarid and tropical climates. Scenarios of future climate change predict an increase of severe drought conditions in temperate climate regions. This implies the need for adaptations of locally managed forest systems, such as European beech (*Fagus sylvatica* L.) monocultures, for instance, through the admixing of deep-rooting silver fir (*Abies alba* Mill.). We designed a stable-isotope-based split-root experiment under controlled conditions to test whether silver fir seedlings could perform HR and therefore reduce drought stress in neighboring beech seedlings. Our results showed that HR by silver fir does occur, but with a delayed onset of three weeks after isotopic labelling with ²H₂O (δ^2 H \approx +6000‰), and at low rates. On average, 0.2% of added ²H excess could be recovered via HR. Fir roots released water under dry-soil conditions that caused some European beech seedlings to permanently wilt. On the basis of these results, we concluded that HR by silver fir does occur, but the potential for mitigating drought stress in beech is limited. Admixing silver fir into beech stands as a climate change adaptation strategy needs to be assessed in field studies with sufficient monitoring time.

Keywords: hydraulic redistribution; drought; silver fir; European beech; mixed stand

1. Introduction

Hydraulic redistribution (HR) is the passive flux of water between wet- and dry-soil zones through plant roots as conduits. It is driven by soil-water potential gradients between dry- and wet-soil layers, and between roots and soil matrix [1,2]. Typically, HR occurs during the night, when transpiration has ceased [3–5]. Water can be redistributed in the upward (i.e., hydraulic lift [2,6,7]), downward (i.e., hydraulic descent [8–11]), and lateral directions [12–15]. Field observations showed that HR plays an important role in terrestrial ecohydrological cycles. Plants can benefit from HR through enhanced photosynthesis and transpiration [16], alleviated soil-moisture loss during the dry season [17], and a prolonged growing season [18,19]. These immediate benefits of HR consequently enhance nutrient acquisition [20], increase nutrient mobility, and facilitate root-litter decomposition [21,22].



HR has been documented in more than 100 species [23], including agricultural crops and grasses [24–26], and forest trees and shrubs [27–29]. Although HR has been observed in diverse climatic settings [16,23,30,31], it is most prevalent in arid and semiarid ecosystems, such as savannas [16,32,33], arid climates and semideserts [5,34–36], Mediterranean-type ecosystems [10,37–39], and tropical forests [40–42]. However, HR may become increasingly relevant in temperate ecosystems that are subject to intensified drying–wetting cycles due to extreme drought events in projected climate-change scenarios [27,43].

Several techniques can be used to identify HR under laboratory or field conditions. Reverse water flow in roots can be identified by sap-flow techniques like the heat-balance [29,44] or -ratio method [45,46]. However, quantification of reverse water flow in roots by sap-flow measurements can easily be disrupted by fluctuations in ambient temperature [46,47], or small-scale soil and geomorphic variability [10]. Furthermore, sap flow can only be measured in individual roots and upscaling to the root system is a significant source of error [48]. Contrastingly, measurements of soil-water potential near plant roots, accompanied by water stable isotope analyses, have been used to track water movement in soil [30,49,50]. In this context, stable isotopes, either at natural abundance or by using heavy isotope enriched water (i.e., $H_2^{18}O$, 2H_2O) as a tracer, are used as a novel technique to quantify water flow between dry- and wet-soil layers through plant roots and water uptake by adjacent plants [3,27,51].

In temperate forests, HR was only detected in a few tree species in the field, such as Norway spruce (*Picea abies* (L.) Karst.), Douglas fir (*Pseudotsuga menziesii*), ponderosa pine (*Pinus Ponderosa*), loblolly pine (*Pinus taeda*), and sessile oak (*Quercus petraea*) [51–55]. In an adult mixed oak/European beech forest, Zapater et al. [51] showed HR by oaks using an ¹⁸O-labelling approach but did not find any tracer material in European beech. However, both HR and the uptake of redistributed water by neighboring plants was detected in studies with seedlings of English oak, Norway spruce, and European beech under moderate drought in split-root systems in the greenhouse [27].

In Central Europe, European beech, being both an abundant natural tree species and a key species in forestry, was reported to be particularly vulnerable to drought [56,57]. As extreme drought events and intensified drying–wetting cycles are projected to become more prevalent [43,58,59], beech forests in Central Europe face consequences such as declining growth and drastic economic losses for forestry [60–62]. Admixing deep-rooting tree species could potentially increase the resilience of beech stands. In this context, silver fir was proposed due to its high productivity and presumably higher drought resistance [63,64]. Furthermore, recent studies indicate that water supply to European beech in mixed forest stands may be supported by the presence of silver-fir neighbors [65,66].

The aim of this study was to show if silver fir can perform HR under extreme drought conditions. For this purpose, we applied an improved split-root approach under controlled conditions, combined with the ${}^{2}\text{H}_{2}\text{O}$ labelling of water and in situ stable isotope analysis of soil moisture. We hypothesized that fir roots were able to allocate ${}^{2}\text{H}_{2}\text{O}$ from moist- to dry-soil zones by HR.

2. Materials and Methods

2.1. Mesocosm Setup

This study was conducted using plant-soil mesocosms under controlled conditions in the scientific greenhouse at the KIT Campus Alpin in Garmisch-Partenkirchen, Germany. Temperature (T) and relative humidity (rH) were controlled and underwent daily cycles (T = $20.5 \pm 4^{\circ}$ C, and rH = 58 ± 12 % on average). Ambient CO₂ concentration showed diurnal fluctuations between 380 and 450 ppm without long-term trends during the timespan of the experiment.

Six mesocosms, each comprising 2 nested polyvinylchloride (PVC) compartments (dimensions of inner and outer compartment: length × width × height = $90 \times 38 \times 40$ cm³ and $20 \times 20 \times 10$ cm³, respectively) were set up as shown in Figures 1 and 2. Soil for the mesocosms was collected in autumn 2015 in the Black Forest close to Emmendingen (SW Germany). The material was taken from the Ah horizon of a Dystric Cambisol that originated from Triassic sandstone and showed a sandy loam texture

(see [67] for site and soil details). The soil material was mixed with perlite at a volume ratio of 1:1. Perlite is a highly porous mineral that improves soil drainage and aeration properties while retaining moisture. These properties helped with the homogenization of soil moisture and isotope equilibration.



Figure 1. Experiment mesocosms. Two fir and 1 beech saplings were planted in polyvinylchloride (PVC) compartments, whereas a beech sapling was isolated within a smaller compartment in the center. A root strand of each fir was redirected into the beech compartment. Soil moisture was extracted via diffusion into dry air passing through gas-permeable tubing connected to a water isotope analyser (Picarro L 2130-i). Experiment comprised 6 such mesocosms; 3 were used as replicates and 3 as controls (see main text for more detail).



Figure 2. Photo of two mesocosms with silver-fir and European beech saplings in separate compartments.

Two silver-fir seedlings (3.5 years of age) were planted in the outer compartment (hereafter referred to as the fir compartment), and a single European beech seedling (2 years of age) in the inner

compartment (hereafter referred to as the beech compartment). Root length of the fir and beech seedlings was, on average, 30 and 15 cm, respectively. A first-order coarse root with intact second- and third order fine roots of each silver fir seedling was redirected to the beech compartment. The mesocosms were then wrapped with plastic film to avoid evaporation and consequent contamination of the lab environment with 2 H₂O vapor. The fir-root strands were the only hydraulic connection between beech and fir compartments. As experiment control, 1 mesocosm was left unplanted but otherwise received identical treatment (Figure 1).

2.2. Environmental Parameters

Volumetric soil moisture and temperature were measured using DECAGON EM50 loggers (Decagon Devices, Inc., Pullman, Washington, DC, USA). A soil-moisture sensor (Type GS1 and 5TM, Decagon Devices, Inc., Pullman, WA, USA) was vertically installed in each beech compartment at approximately 10 cm depth. Volumetric water content (VWC) was recorded every 2 hours. Reported soil-moisture data were calibrated against gravimetric measurements of the same soil material used in the HR experiment. We observed erratic readings from soil-moisture sensors at very low soil-water contents. This had implications on the calculation of soil-water potential, as is explained in the following section.

2.3. Soil-Water Potential

Soil-water potential was calculated for the beech compartments with a widely used model for water retention in soils [68]:

$$\theta = \theta_r + \frac{(\theta_s - \theta_r)}{\left(1 + (\alpha |\Psi|)^n\right)^m} \tag{1}$$

where $|\Psi| = \frac{1}{\alpha} \times \left\{ \left(\frac{(\theta - \theta_r)}{(\theta_s - \theta_r)} \right)^{-\frac{1}{m}} - 1 \right\}^{\frac{1}{m}}$, and Ψ is the absolute value of soil-water potential (hPa/cm water

column) at a specific volumetric water content θ (cm³ cm⁻³). θ_s and θ_r were the saturated and residual water content of the soil, respectively. Parameters α (hPa), n, and m are shape parameters. For fitting, θ_s and θ_r were determined from measurements, and α and n were estimated using the soil texture of the potting substrate (sandy loam texture) according to Hodnett et al. [69]. For calculation, we used gravimetrically calibrated volumetric soil moisture. We conducted predawn water potential ($\Psi_{predawn}$) measurements on 2 October to validate model results. $\Psi_{predawn}$ was measured on beech branches using a Scholander pressure chamber [70] (Model 1000 Pressure Chamber Instrument, PMS Instrument Company, Albany, Oregon, USA). Plant $\Psi_{predawn}$ could be used as an estimate for soil Ψ on the basis of the assumption that plant $\Psi_{predawn}$ was in equilibrium with soil Ψ adjacent to roots. Water potential is expressed as pF values, which is the logarithm of the absolute values of Ψ (hPa). Due to potential biases inherent to VWC measurements at very dry soil conditions, pF calculation can be subject to large uncertainty leading to pF values above the physical limit of ~6.9.

2.4. Leaf-Gas Exchange Measurements

Leaf–gas exchange measurements on beech seedlings were performed from August to October, and captured the drought response of beech. We measured light-saturated photosynthesis (A_{sat}) and stomatal conductance (g_s) using a portable leaf–gas exchange system (Li-6400, LI-COR Inc., Lincoln, NE, USA) equipped with a light source (Li-6400-02B LED, LI-COR Inc., Lincoln, NE, USA). One green leaf per beech tree was measured under predetermined saturated light conditions (PAR = 1200 µmol m⁻²s⁻¹), an average leaf temperature of 27.7 °C, and average relative humidity of 54%.

2.5. Hydraulic-Redistribution Experiment

Prior to the experiment, the VWC of beech compartments was maintained at roughly 15% by irrigation. Each mesocosm was then exposed to drought conditions by withholding irrigation for 25 days. On the night from 27 to 28 August, 2 L (i.e., 5.8 Lm^{-2}) of deuterated water ($\delta^2 \text{H} \approx +6000\%$) was injected into the bottom-soil zone of the fir compartment of 3 mesocosms containing trees and the unplanted control mesocosm. For each fir compartment, 6 evenly spread-out irrigation tubes (Figure 1) were used as injection ports in order to ensure the homogeneous distribution of deuterated water. The two remaining mesocosms containing trees received no label injection. Isotopic composition of soil moisture in the beech compartment was measured on 15 separate days during the 10 following weeks. Measurements generally took place during the morning hours until noon. Each beech compartment was re-watered on 29 October with 1.5 L of tap water in order to weaken the water-potential gradient between the fir and beech compartments, and, thus, the required conditions for HR.

2.6. Deuterated-Water Tracing

We used a membrane-inlet water-vapor sampling technique in soil coupled to a cavity ringdown laser spectrometer [71,72] to trace ${}^{2}\text{H}_{2}\text{O}$ transport from the fir to the beech compartment. For this purpose, the beech compartment of each mesocosm was equipped with gas-permeable tubing (Accurel PP V8/2HF polypropylene tubing, Membrana GmbH, Germany: 30 cm long sections buried at 5 and 10 cm depth) that was flushed with synthetic dry air (20% O₂ in N₂) during measurements (Figure 2). The dry air absorbed moisture upon passing the gas-permeable section. This approach resembled in situ soil air probes [72], but with double the length of gas-permeable tubes in each beech compartment. Accurel tubes are suitable for isotopic measurements, as they do not cause isotopic fractionation across a wide range of soil-moisture contents, and render possible vapor measurements in equilibrium with soil water [71].

A Picarro L 2130-i cavity ringdown spectrometer was used to analyze the stable isotopic composition (δ^2 H) of the gas stream. On measurement days, gas streams exiting each of the 6 mesocosms were continuously measured for at least 15 min. To minimize carryover artefacts, lines were flushed for 20 min between each measurement, and the values for the first 5 min of each measurement were ignored. Stable isotope data reported in the results section of this study refer to the average values of the remaining measurement time. Results of each day were scale-normalized to 2 in-house standards (δ^2 H = -235.0 ± 1.8 ‰ and 1.8 ± 0.9 ‰ VSMOW) that were calibrated against international reference materials (VSMOW2, SLAP2).

HR was quantified in this study as a percentage of excess deuterium (*d*) added to the fir compartment by labelling, that was recovered in the beech compartment. In other words, ²H excess recovery (Δd) corresponds to the needed amount of ²H from labelled water to reach the measured isotopic composition of a defined volume of water. We used isotopic measurements to calculate a mixing ratio between labelled water (i.e., $\delta^2 H = +6000\%$) and background moisture using the isotopic composition of the control mesocosm as a baseline to account for natural variability. This mixing ratio is used to determine the amount of excess ²H in beech compartments, relative to excess ²H in the label water based on the VWC (θ) of the beech compartment and the amount of added label water (Equation 2):

$$\Delta d = \frac{\Delta d_{beech}}{\Delta d_{label}} \times 100 = \frac{\left(\varepsilon^2 H_{beech} - \varepsilon^2 H_{control}\right) \frac{m_{beech}}{m_O} \theta}{\left(\varepsilon^2 H_{label} - \varepsilon^2 H_{control}\right) \frac{m_{label}}{m_O} M_{label}} \times 100,\tag{2}$$

where Δd is the percentage of ²H excess recovered in beech compartments, Δd_{beech} and Δd_{label} are the ²H excess [g] in the respective water reservoir (i.e., beech compartment or deuterated label water), $\varepsilon^2 H_{beech}$, $\varepsilon^2 H_{beech}$, and $\varepsilon^2 H_{beech}$ are the isotopic enrichment of the respective water reservoir (given in atomic % of deuterium calculated from δ^2 H values), $\frac{m_{beech}}{m_O}$ and $\frac{m_{label}}{m_O}$ are the molar weight ratio between hydrogen and oxygen in the respective water reservoir at the measured isotopic composition, θ is

the VWC of beech compartments, and M_{label} is the absolute amount of label water added to the fir compartment [g].

3. Results and Discussion

After 25 days of drought treatment, the VWC in the planted beech compartments declined to $6\% \pm 2\%$, while VWC in the unplanted control compartment was still at 16%. Soil-water potential in the beech compartments of the mesocosms ranged between pF values of ca. 2.0 and 3.5 (Figure 3c). Net beech photosynthesis had declined to an average of 8 µmol m⁻² s⁻¹ (Figure 3a), and stomatal conductance showed values of around 0.14 mol m⁻² s⁻¹ (Figure 3b). After the fir compartments were labelled with 2 L of deuterated water, the net photosynthesis and stomatal conductance of beech continued to decline to 4 µmol m⁻² s⁻¹ and 0.05 mol m⁻² s⁻¹, respectively, one week after labelling.



Figure 3. (a) Time series of light-saturated photosynthesis (A_{sat}) and (b) stomatal conductance (g_s) of beech seedlings as measurement averages ± standard error (n = 5). (c) Soil matrix potential in beech compartments (daily means) as pF-values, log(|hPa|) for three labelled (black, grey, light grey), two unlabelled (green, light green), and one unplanted control (light blue) mesocosms. Solid vertical line: time of deuterium labelling; dotted vertical line: re-watering of beech compartments. Very high pF values of some replicates from 10 April onwards were likely related to volumetric-water-content (VWC) measurement bias in very dry soil conditions.

These data showed that the beech trees experienced severe drought stress during the experiment that was not significantly counteracted by the silver-fir seedlings. Previous studies also reported a similar development for mature trees in the field under severe drought conditions at mixed fir and beech cultivation [66].

In unlabeled mesocosms and the labelled control mesocosm, δ^2 H values of soil moisture in the beech compartment remained between –120‰ and –80‰ VSMOW throughout the entire incubation period (Figure 4a). Labelled mesocosms showed similar or slightly enriched values for the first three weeks after labelling. In addition, ²H excess recovery in the beech compartments was negligible during that time. This suggested that no detectable HR occurred for three weeks after label injection.



Figure 4. (a) Time series of δ^2 H in soil moisture in beech compartment of ²H-labeled mesocosms (grey), planted unlabelled mesocosms (green), and labelled control mesocosm without plants (light blue). Dashed lines: time series of each respective replicate; solid lines: their mean values. Re-watering was with unlabelled water. (b) Time series of ²H excess recovery in beech compartments expressed as percentage of ²H excess added by deuterated water to fir compartment. Isotopic composition of control mesocosms' beech compartment used as baseline for excess calculation. Grid was spaced at 14 day intervals.

During this initial three-week period, soil pF further increased in the beech compartments (Figure 3c). Values for δ^2 H in the beech compartments started to notably rise afterwards (Figure 4a). On the basis of ²H excess recovery, we could assume that traceable amounts of water were reallocated to the beech compartment through the fir roots (Figure 4b). Hence, HR occurred in this experiment with a delay of ca. 2–3 weeks after label application. However, this delayed HR occurred at pF values above the permanent wilting point (i.e., pF = 4.2 according to Amelung et al. [73]; Figure 3c), and partially resulted in the wilting of the beech seedlings. Seedling wilting is also represented by the concurrent drop of photosynthetic activity and stomatal conductance to values close to zero (Figure 3a,b).

Prior to re-watering, ²H excess recovered in the beech compartments amounted to an average of 0.2% of ²H excess added with the 2 L of deuterated water (Figure 4b). Re-watering of the beech compartments with 1.5 L of water at the end of October clearly diluted the δ^2 H signal (Figure 4a). Furthermore, pF values increased above the permanent wilting point. However, ²H excess recovery was initially doubled by re-watering, followed by rapid decline (Figure 4b). We assumed that the unexpected doubling of ²H excess upon re-watering was caused by overestimated VWC measurements directly after re-watering the beech compartments (water amount was part of the calculation of ²H excess recovery; see Equation 2). As indicated by δ^2 H dynamics, re-watering seemed to eliminate the large gradient in water potential, which was needed to sustain HR through water loss from the fir roots.

Overall, δ^2 H was correlated to pF values in the beech compartments (Figure 5). This suggests that water-potential gradients largely drove HR by the silver-fir roots, and HR was stronger at high pF values. A similar observation was made in a loblolly pine stand, where hydraulic redistribution was determined by reverse root flow and was shown to increase with soil drought [74]. However, in split-root experiments for Norway spruce, European beech, and English oak, Hafner et al. [27] observed hydraulic redistribution already under moderate drought (water potential of ca. -0.5 MPa/-5000 hPa/pF value of 3.7) after only several days. In these conditions, we could not find evidence to support HR at similar time scales. Still, our results were consistent with the observation that, in

a mature mixed beech–fir forest, the presence of fir slightly improved the xylem-sap flow density of beech compared to a beech monoculture [66]. However, this improvement was not enough to counteract the negative effect of drought on xylem-sap flow density. In addition, the positive effect of the presence of fir on xylem-sap flow density in beech upon drought was reversed when the forest stand was subjected to precipitation after drought. These data indicated that cocultivation of beech and fir may not enhance the resilience of beech to drought events through HR.



Figure 5. Relationship between δ^2 H and pF values in beech compartments of labelled mesocosms. Only incubation period from labelling until re-watering was considered.

4. Conclusions

We set up a split-root experiment with silver-fir and European beech seedlings using ²H labelled water to track HR from wet- to dry-soil zones through silver-fir roots. In situ measurements of the isotopic composition of soil moisture showed detectable HR at 2–3 weeks after label application. A positive correlation between soil pF values and the isotopic composition of soil moisture confirmed that changes in water potential drive HR. Considering that only 0.2% of the added ²H could be recovered at the permanent wilting point, we concluded that the admixing of silver fir to European beech stands likely does not increase the resilience of beech monocultures to severe drought conditions. The delayed onset of HR of 2–3 weeks in this study led us to recommend that future field studies allow for a longer monitoring time to fully detect hydraulic redistribution.

Author Contributions: Conceptualization, H.R., F.Y., P.T., and M.D.; methodology, P.T., F.Y., H.R., M.D., N.K.R.; data processing, P.T., F.Y., S.R., R.R.; writing—original-draft preparation, P.T., F.Y., M.D., H.R., S.R.; writing—review and editing, all authors; visualization, P.T., S.R.; supervision, H.R., M.D., N.K.R. All authors have read and agreed to the published version of the manuscript.

Funding: This study was funded by the Federal Ministry of Food and Agriculture (BMEL) within the "Waldklimafonds" program and the "Buchen–Tannen–Mischwälder zur Anpassung von Wirtschaftswäldern an Extremereignisse des Klimawandels (BuTaKli)" project; their support is gratefully acknowledged. Additional funding was provided by the German Federal Ministry of Education and Research (BMBF) through the Helmholtz Association and its ATMO research program. RR and NKR acknowledge support from the German Research Foundation through its Emmy Noether Program (RU 1657/2-1).

Acknowledgments: We would like to thank Andreas Gast, Nils Risse, and Stefanie Dumberger for their assistance with the experiment setup and leaf–gas exchange measurements. We would also like to acknowledge the two anonymous reviewers whose comments have greatly improved the manuscript.

Conflicts of Interest: The authors declare no conflict of interest.

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