

Toward Reproducible Enzyme Modeling with Isothermal Titration Calorimetry

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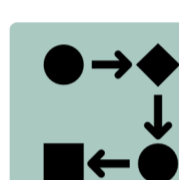
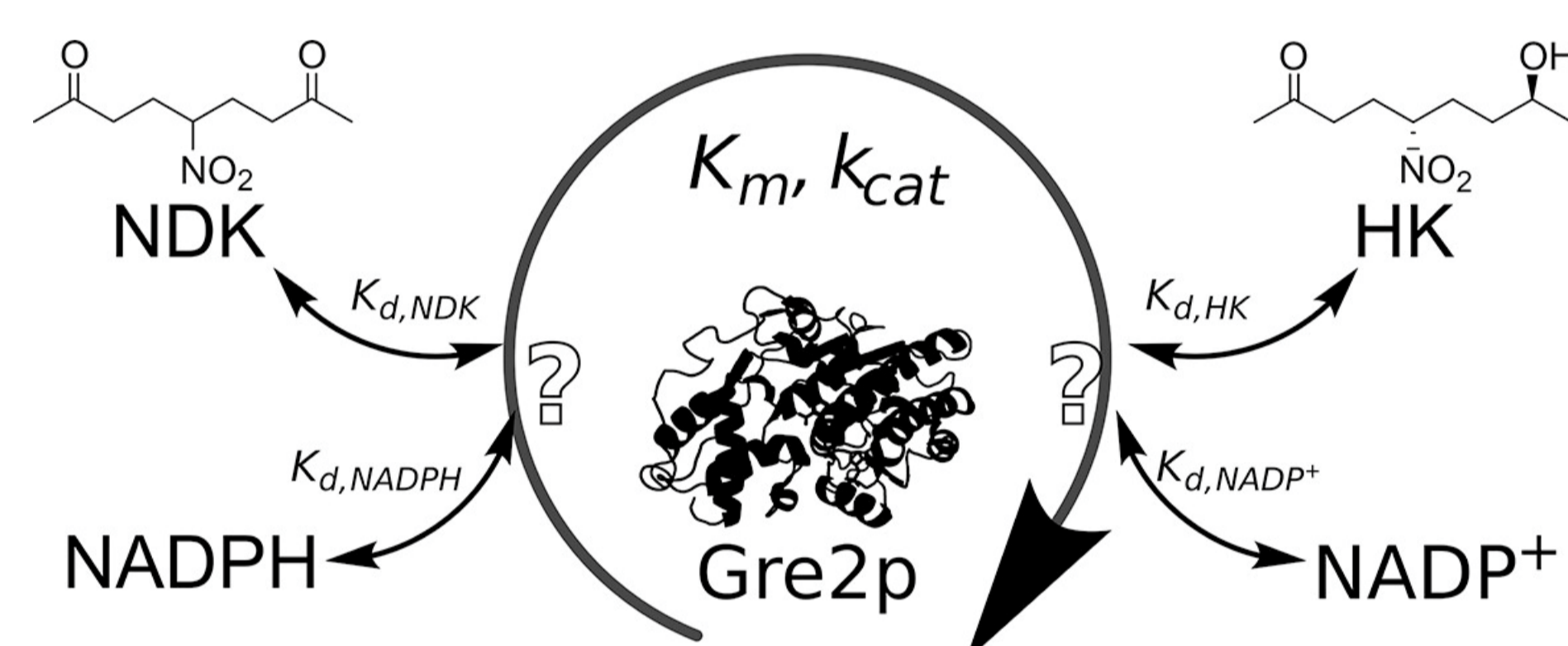
Relevance

To apply enzymes in technical processes, a detailed understanding of the molecular mechanisms is required. Kinetic and thermodynamic parameters of enzyme catalysis are crucial to plan, model, and implement biocatalytic processes more efficiently. While the kinetic parameters, K_m and k_{cat} , are often accessible by optical methods, the determination of thermodynamic parameters requires more sophisticated methods. Isothermal titration calorimetry (ITC) allows the label-free and highly sensitive analysis of kinetic and thermodynamic parameters of individual steps in the catalytic cycle of an enzyme reaction.



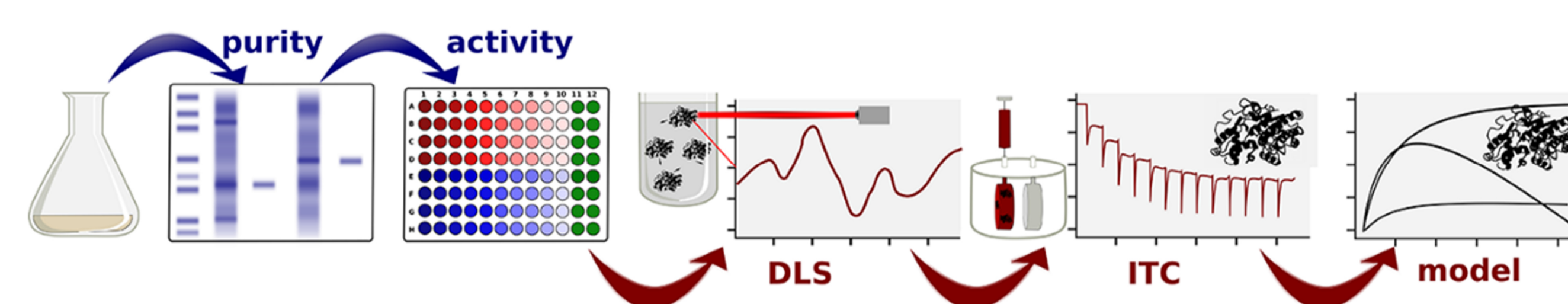
Aim

We aimed to establish an ITC-dependent work flow to determine both the kinetic and the thermodynamic data for a NADPH-dependent enzyme Gre2p. This workflow should function as blueprint for future investigations of different enzymes. For this purpose, it is important to ensure reproducibility by the scientific community.



Methods

Traditional approaches to enzyme kinetic data (blue) primarily use spectrophotometric activity measurements. The implementation of DLS and ITC for quality control and mechanistic insights, respectively, leads to an increase in data quality to enable robust modeling.



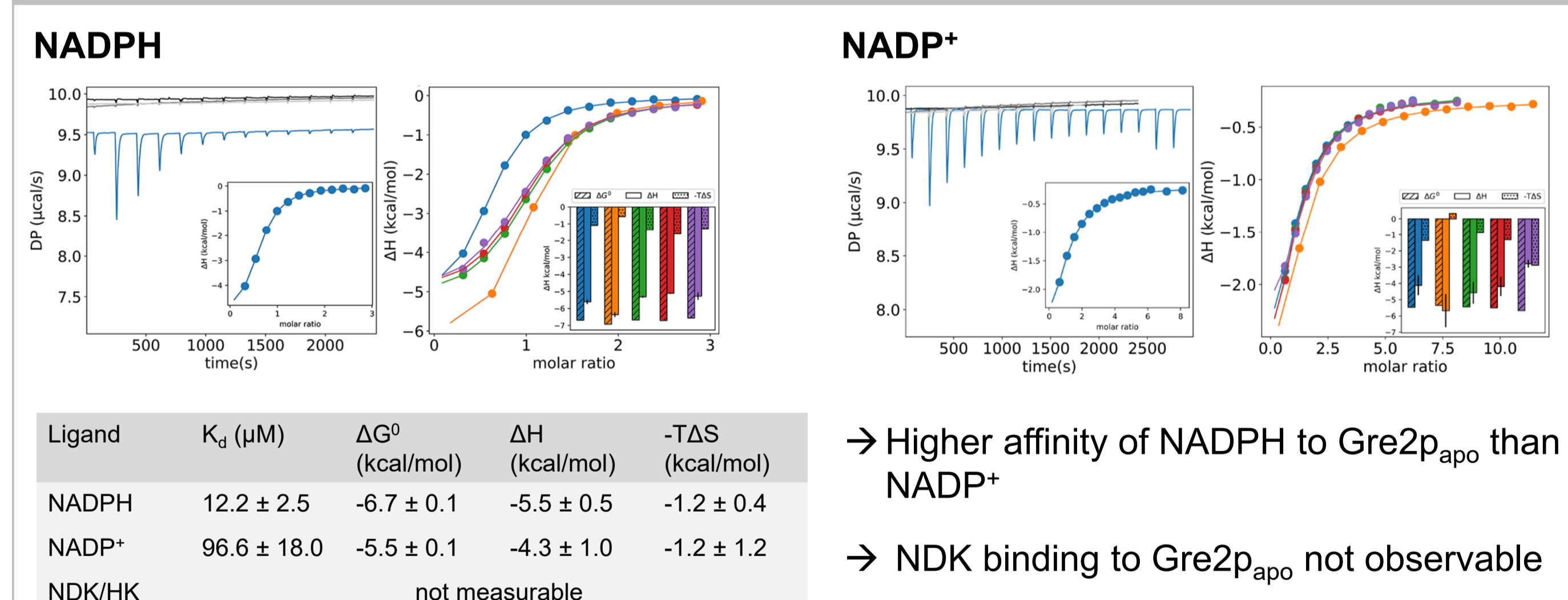
To achieve reproducibility, standard operating procedures (SOPs), data analysis and modeling workflows were published open and F.A.I.R. (findable, accessible, interoperable, and reusable).



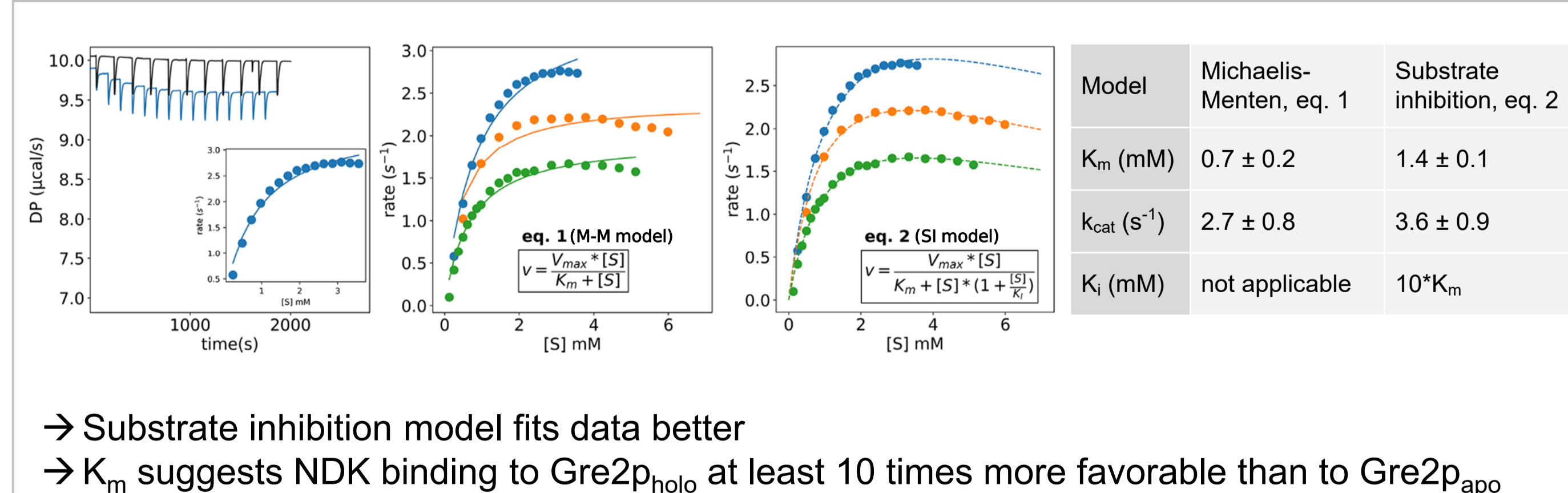
Findings

To untangle the reaction mechanism, traditional ITC binding experiments (ITC-BIND), multiple injection experiments (ITC-MIM) and single injection experiments (ITC-SIM) were performed.

ITC-BIND



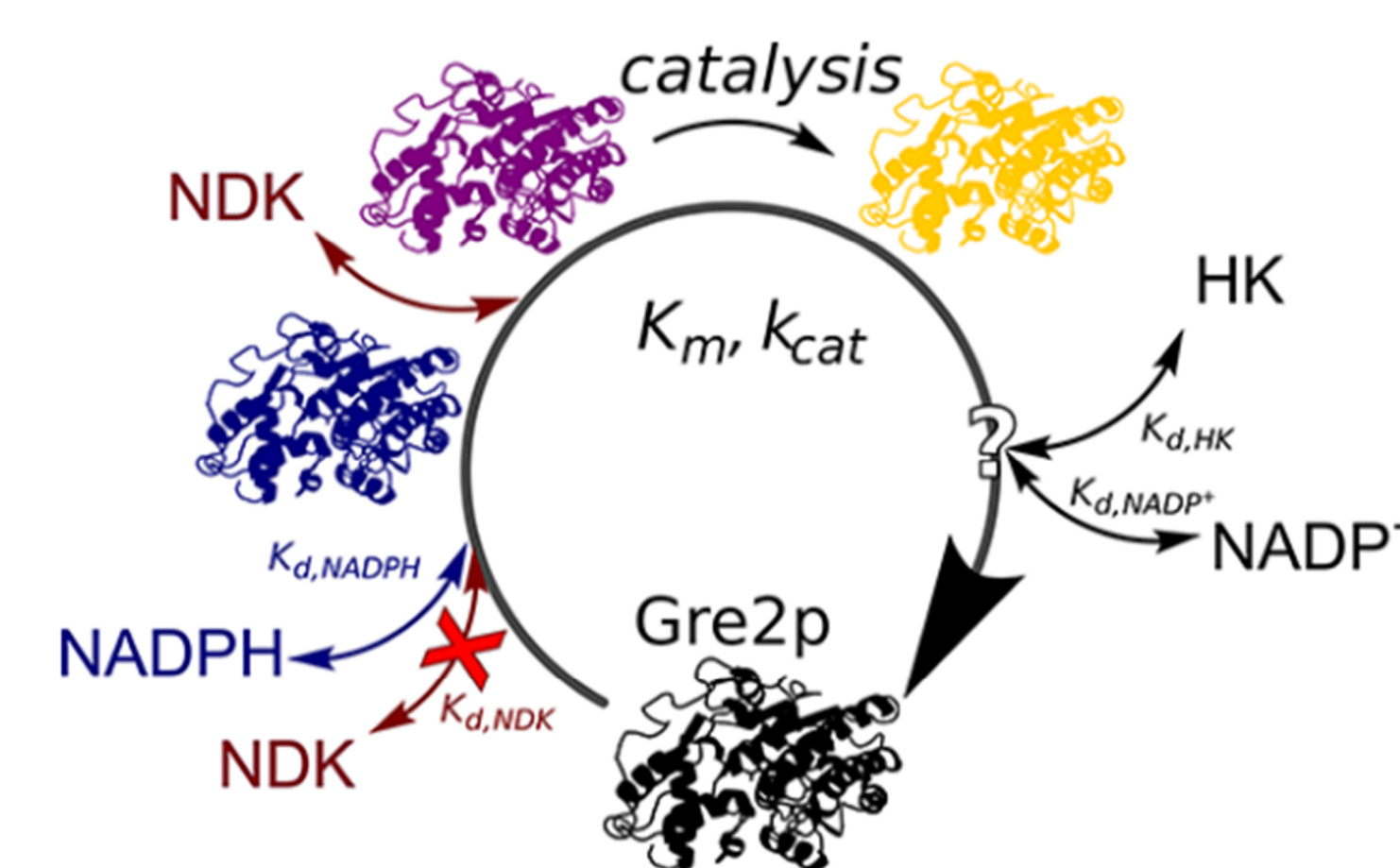
ITC-MIM



Proposed mechanism

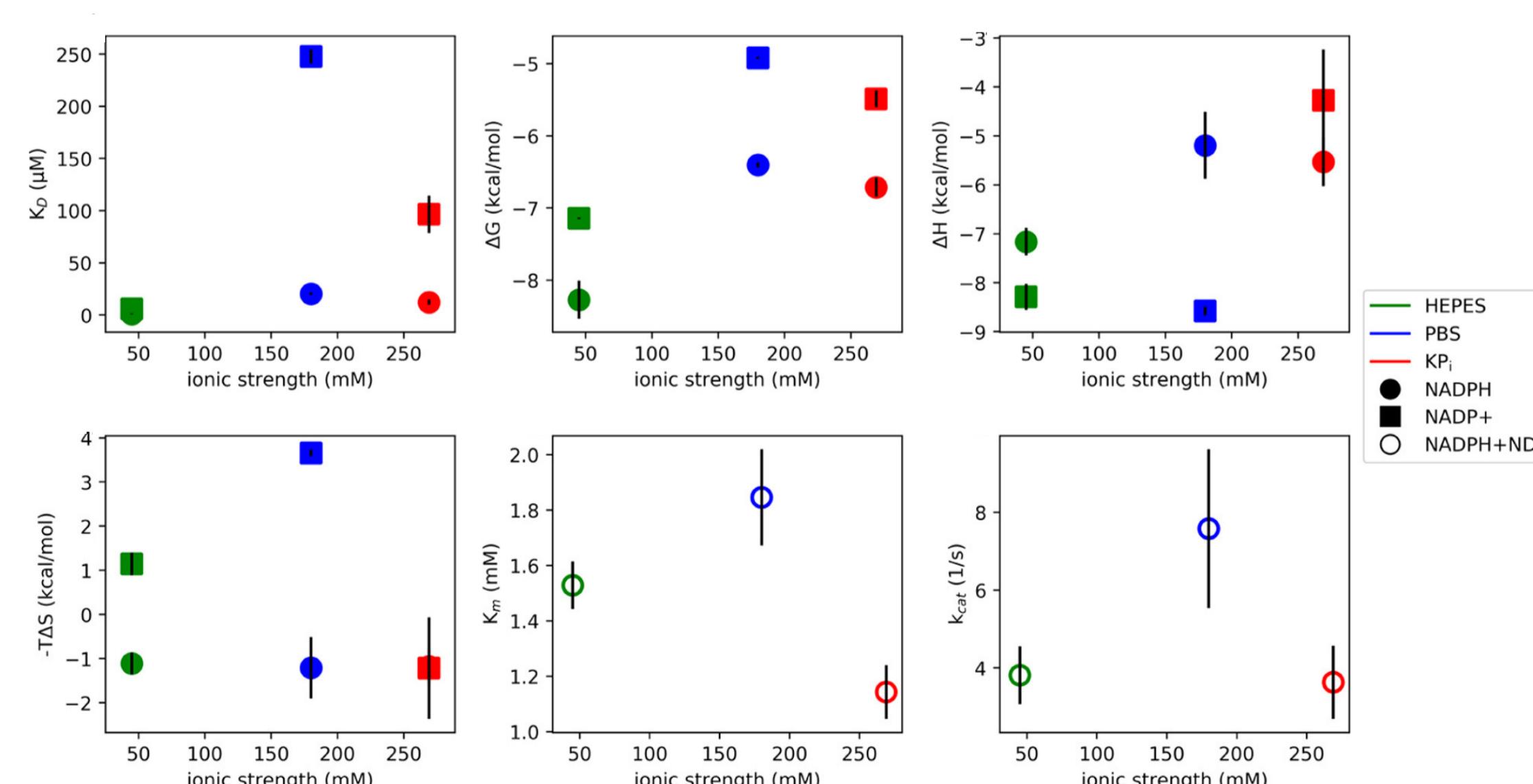
Ordered sequential:

- NADPH binds first to Gre2p_{apo} (black)
- Gre2p_{holo} (blue) has increased affinity to the substrate NDK which is represented in part by K_m
- Order of unbinding of HK and NADP⁺ cannot be determined experimentally



Impact of reaction buffer

Using different buffers changes the binding and kinetic parameters. Three buffers with low enthalpy of ionization of different ionic strengths were chosen for initial impressions.



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Sources

Publication: Ott F., Rabe K. S., Niemeyer C. M., Gygli G. (2021) Toward Reproducible Enzyme Modeling with Isothermal Titration Calorimetry, ACS Catalysis, 10695-10704.

Repository: Data, workflows and standard operating procedures are publicly available at fairdomhub.org.

