

# Does surface wettability influence bacterial adhesion?

## Annika Rieder<sup>1</sup>, Tatjana Ladnorg<sup>2</sup>, Christof Wöll<sup>2</sup>, Reinhard Fischer<sup>3</sup>, Ursula Obst<sup>1</sup>, Thomas Schwartz<sup>1</sup>

<sup>1</sup> Karlsruhe Institute of Technology, Institute of Functional Interfaces (IFG), Department of Microbiology of Natural and Technical Interfaces <sup>2</sup> Karlsruhe Institute of Technology, Institute of Functional Interfaces (IFG), Department of Chemistry of Oxydic and Organic Interfaces <sup>3</sup> Karlsruhe Institute of Technology, Institute for Applied Biosciences, Department of Microbiology

## Problem and Objective

Bacteria establish biofilms on a great variety of natural and synthetic surfaces. Especially in industry and medicine biofilms can cause various problems. In order to influence bacterial adhesion and manipulate biofilm formation the surface properties of a material need to be changed.

Hydrophobins are potential candidates for the large-scale surface modifications and change of surface hydrophobicity.

## Introduction – What are hydrophobins?



- highly surface active fungal proteins
- approximately 100 amino acids long (molecular mass 10kDa) amphiphillic
- self-assemble on interfaces into highly stable monolayers verv efficient
- non-toxic and non-immunogenic



# Characterization of hydrophobin-coated surfaces

#### Surface coating

The materials were incubated in hydrophobin solution (10 µM) and subsequently treated with 2% SDS at 80°C for coating stabilization (β-sheet shift).

#### Change of surface hydrophobicity

The surface hydrophobicity was determined by contact angel measurements (CA). A long incubation time (16h) and high incubation temperature (80°C) increased the surface hydrophobicity of hydrophobin-coated surfaces significantly (starting material: glass, CA 11°).

#### Homogeneity of hydrophobin coating

The hydrophobin-coating was specifically detected with His-tag directed fluorescent labeled antibodies and analyzed by epifluorescence microscopy.

It was essential to incubate the materials for 16 hours at 80°C in the protein-solution to form a homogenous hydrophobin-layer after SDS treatment. Atomic force microscopy confirmed these results.

#### Adsorption characteristics of hydrophobins

The adsorption behavior of hydrophobins was analyzed with quartz crystal microbalance with dissipation monitoring (QCM-D). The layer thickness was estimated using Voigt equation.

	layer thickness [nm]
Hydrophobin H*A	17 ± 1
Hydrophobin H*B	15 ± 3

# **Biofilm formation**

The influence of hydrophobin-coatings and changed surface hydrophobicity on different stages of biofilm development was analyzed.

#### Initial bacterial adhesion

The adhesion of a GFP-tagged Escherichia coli was monitored for two days.

#### A preference of initial bacterial adhesion on hydrophobic surfaces was detected.



### Mature Biofilm

Biofilms were grown on hydrophobin-coated and uncoated glass slides in a biofilm reactor in natural wastewater effluent for four weeks.

A similar bacterial population was found on hydrophobincoated and uncoated glassslides.

The effects of hydrophobic surfaces are time dependent.





Influence of incubation time on coating hydrophobicity and homogeneity



AFM images of homogenic hydrophobin layer

# **Conclusion and Outlook**

 Recombinant fusion-hydrophobins are well suited for easy large-scale surface coatings. They adhere in a temperature and time dependent manner into stable monolayers and change surface hydrophobicity.

#### Surface wettability has an impact on primary bacterial adhesion but not on biofilm formation.



· Recombinant hydrophobins modified with antimicrobial agents will be used to influence biofilm formation.

This work was funded by a grant of the BMBF

KIT - University of the State of Baden-Wuerttemberg and National Research Center of the Helmholtz Association