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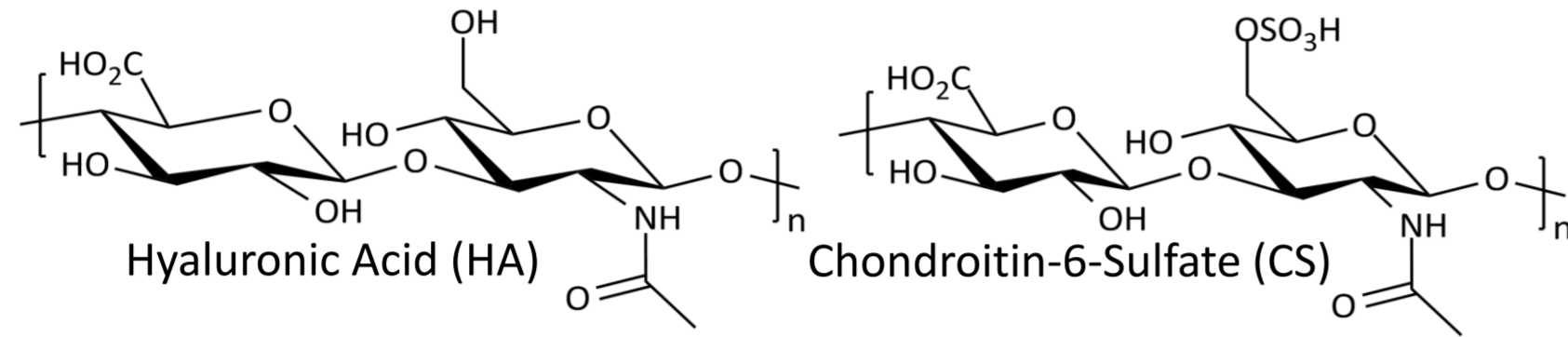
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## Introduction

- Polysaccharides are resistant to the adhesion of proteins, mammalian cells and bacteria<sup>[1]</sup>
- Against marine biofoulers, the coatings perform worse than expected due to the complexation of bivalent ions<sup>[2]</sup>
- Glycosaminoglycans, found in the extracellular matrix, highly hydrophilic, differ in sulfatation of CS, both have only one carboxylic-acid moiety per disaccharide unit. The use of CS is bioinspired due to its presence in fish mucus and its potential contribution to protection of the skin of fish<sup>[3]</sup>
- Amphiphilic surface-coatings have a higher anti-fouling performance than those which are only hydrophilic or hydrophobic<sup>[4]</sup>



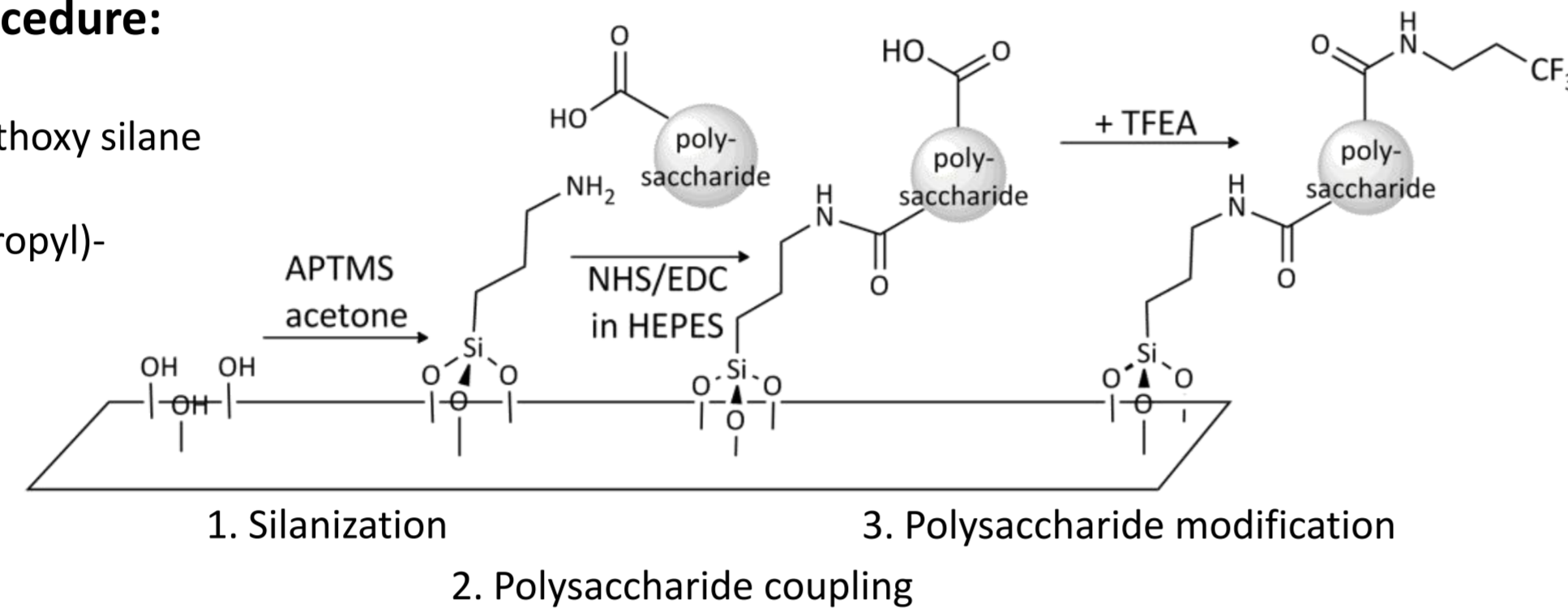
## Motivation and Hypothesis

- Polysaccharides are biocompatible and non-toxic
- Capping of free carboxyl-groups prevents complexation of bivalent cations like Ca<sup>2+</sup> and hence syneresis of the films
- A hydrophobic protection molecule (fluorinated amine) establishes amphiphilic properties in the highly hydrophilic polysaccharide network
- Additionally, the contact angle is shifted towards the minimum in the Baier curve
- Standard assays with a range of different foulers to correlate the change in surface chemistry with the different settlement preferences: biofilm building microorganisms, hard and soft macrofoulers

## Surface Preparation

### Polysaccharide coupling procedure:

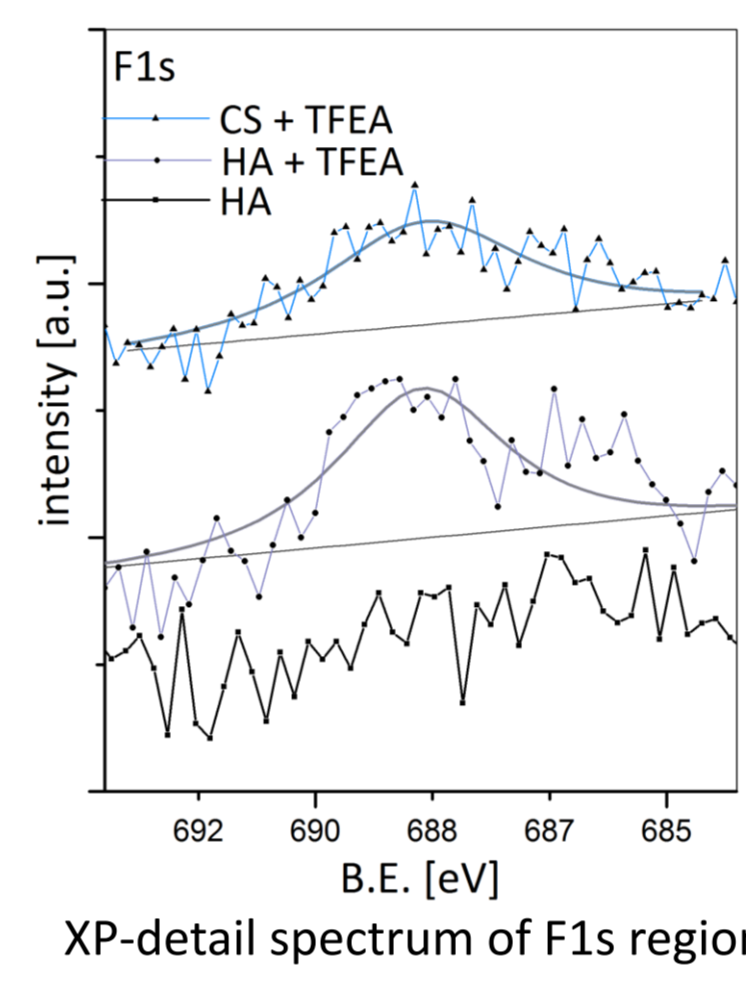
APTMS – 3-Aminopropyltrimethoxy silane  
NHS – N-Hydroxysuccinimid  
EDC – N-(3-Dimethyl amino propyl)-3-ethyl carbodiimid  
TFEA – Trifluoroethylamine



### Characterization:

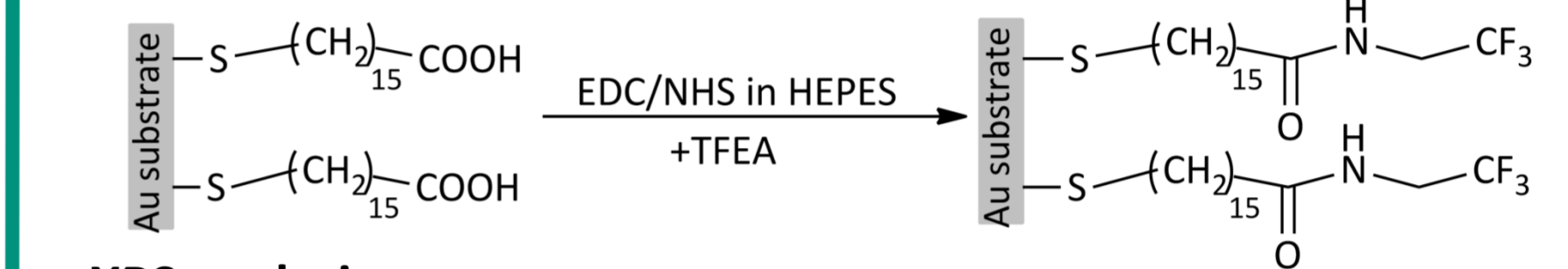
Coating	APTMS	HA	CS	HA + TFEA	CS + TFEA
Contact angle [°]	35 ± 5	< 10	11 ± 3	25 ± 5	28 ± 3
Ellipsometric thickness [Å]	11.4 ± 4.0	26.6 ± 6.6	26.5 ± 7.1	28.2 ± 5.1	28.6 ± 4.0
Thickness from XPS [Å]*	n.a.	23.3 ± 2.5	18.6 ± 5.6	24.8 ± 4.2	23.2 ± 7.7

\* Thickness determined by attenuation of the Si2p signal of a representative APTMS layer

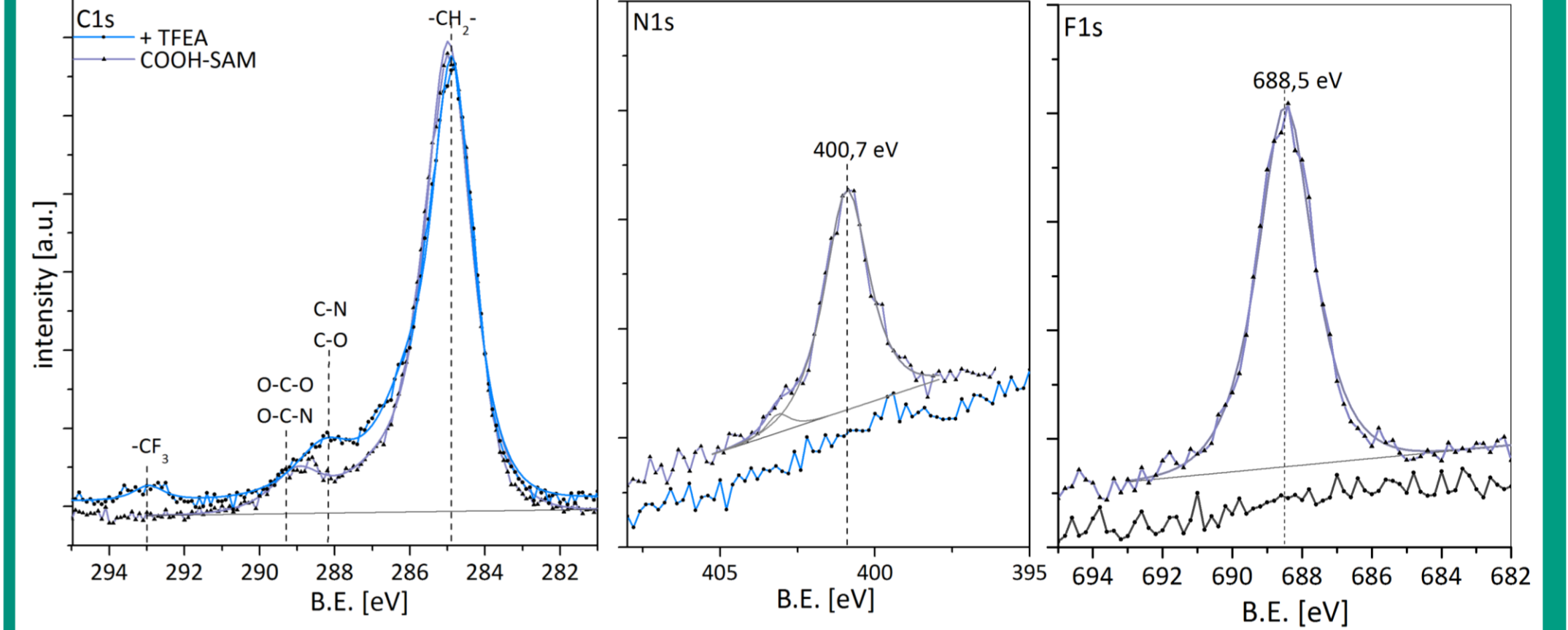


XP-detail spectrum of F1s region

### Proof of amine coupling with simplified SAM-System:



### XPS analysis:



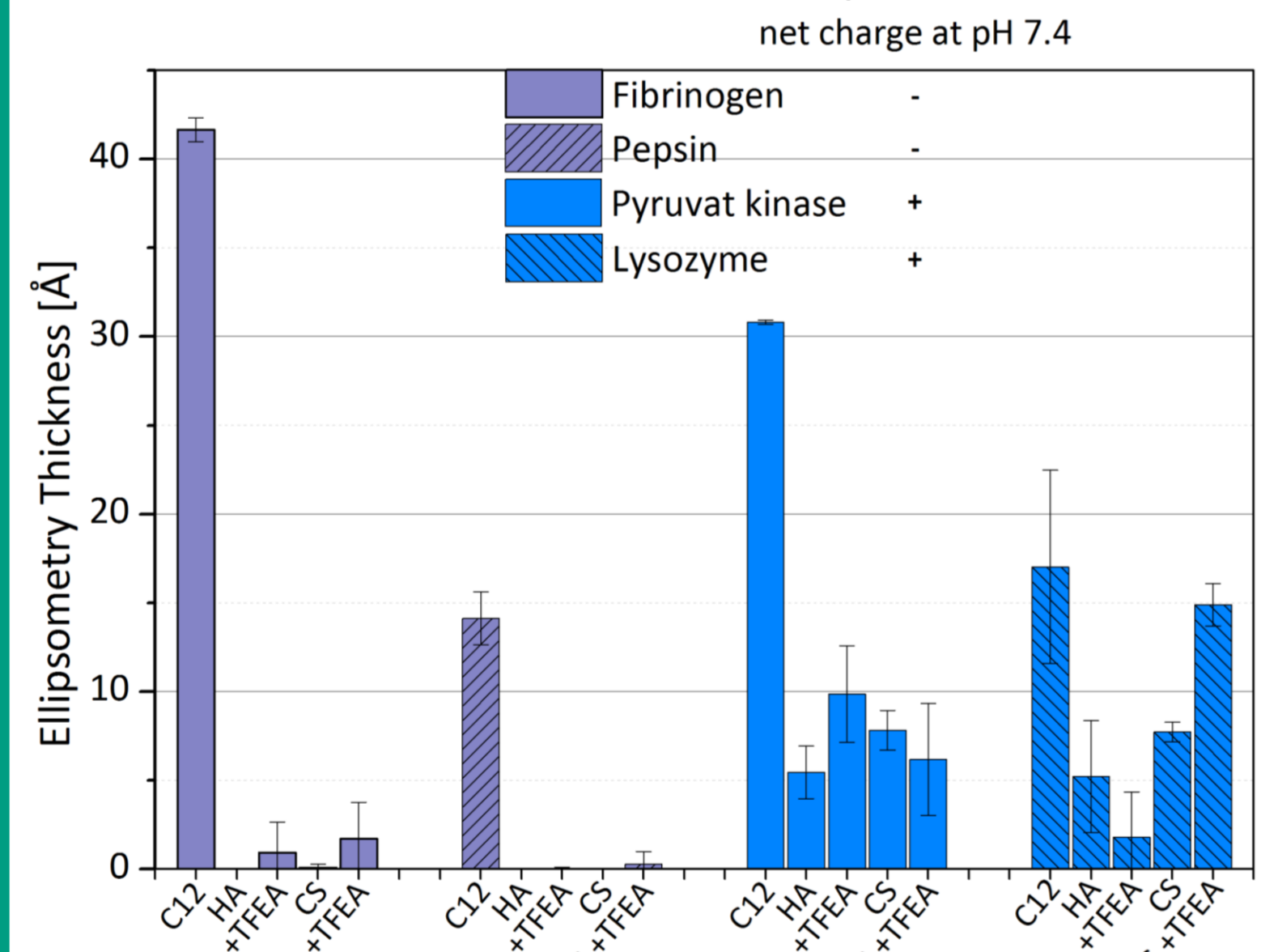
→ Conditions are suitable for intended modification of free carboxyl groups

## Biological Evaluations

### Lab adhesion experiments: Proteins and biofoulers

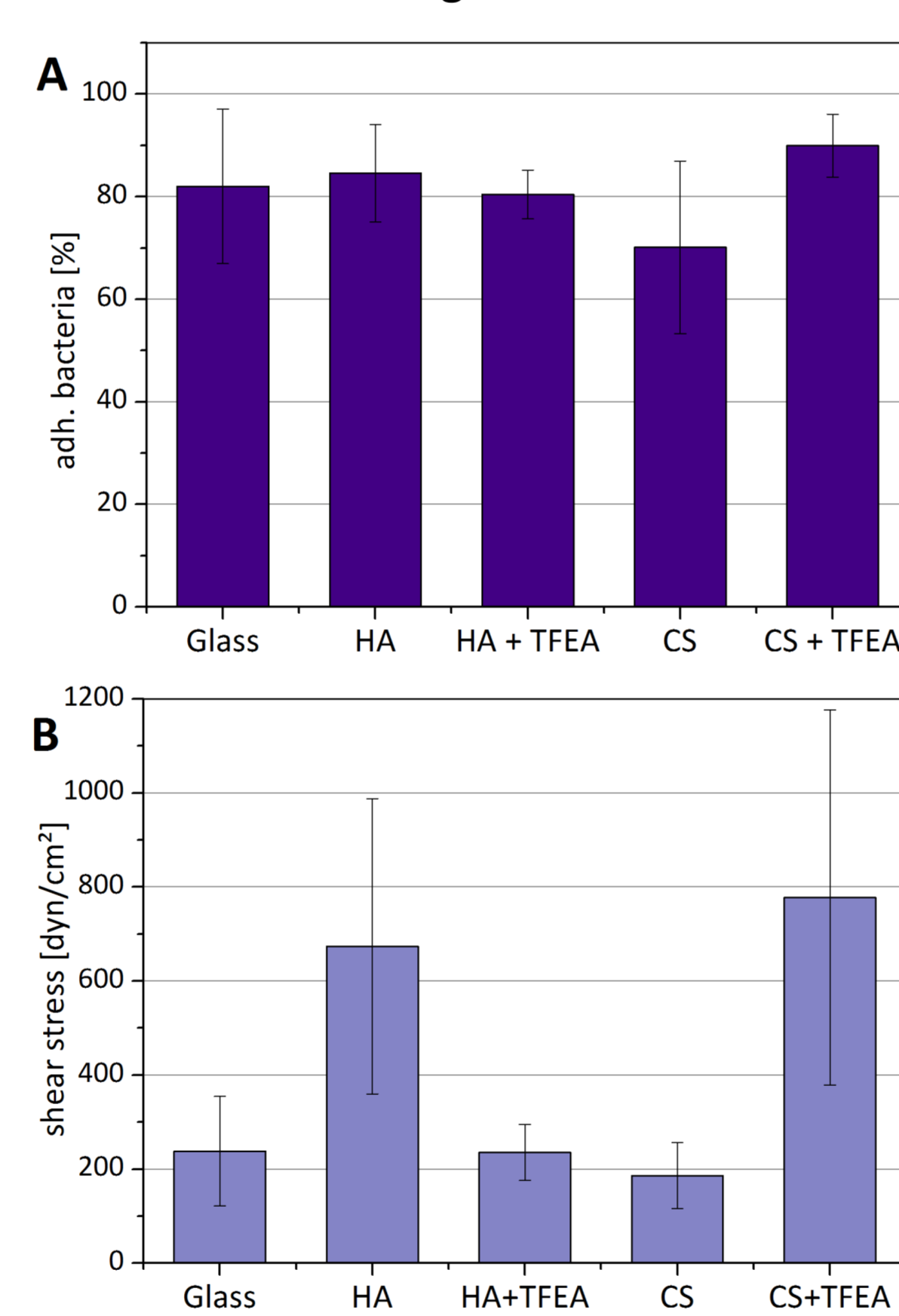
- Protein affinity test:<sup>[5]</sup>  
Determination of protein film thickness by spectral ellipsometry
- Bacteria:  
Microfluidic *Cobetia marina* assay (M.P. Arpa Sancet)
- Algae:  
*Navicula perminuta* settlement and detachment assay (J. Finlay, University of Birmingham)  
*Ulva linza* settlement and detachment assay (J. Finlay, University of Birmingham)
- Barnacles:  
*Balanus amphitrite* settlement assay (N.Aldred, University of Newcastle)

### Protein adhesion assay<sup>[5]</sup>



- All polysaccharide coatings are resistant against both negatively charged proteins
- Positively charged proteins adhere more readily compared to the negative ones, but in most cases weakly compared to the not-resistant C<sub>12</sub>-SAM standard

### Adhesion strength of *Cobetia marina*



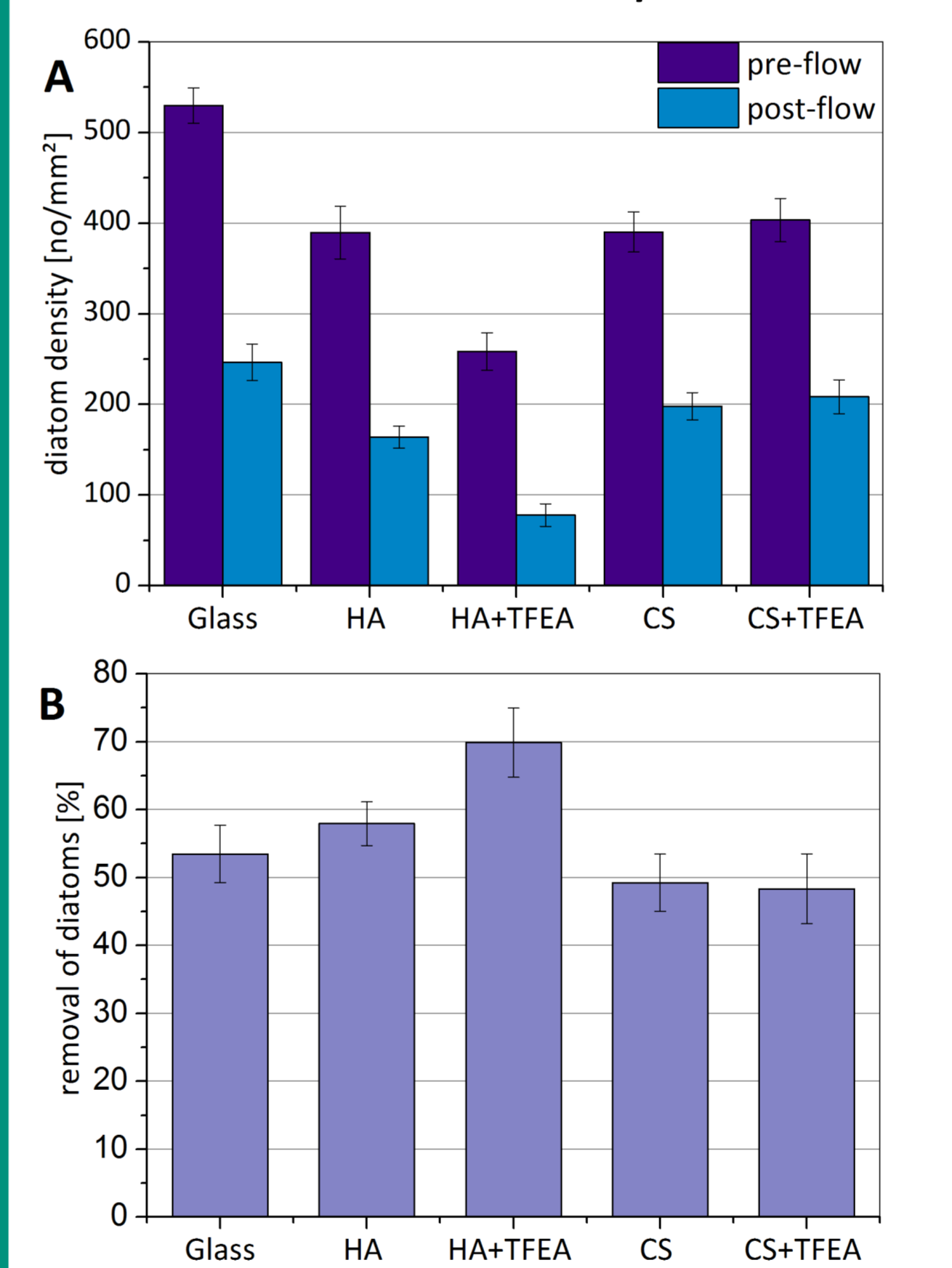
A - Initial settlement after incubation  
B - Critical shear stress to remove 50% of adherent bacteria

- Initial settlement is not influenced by different surface chemistries
- Adhesion strength is weakened on TFEA modified HA surfaces
- Native CS performs very well, behaviour after TFEA capping is reversed

### Field tests in Melbourne, Florida (USA)

- Settlement after 48h incubation in sea water
- Quantification of organisms bigger than 10µm by optical microscopy

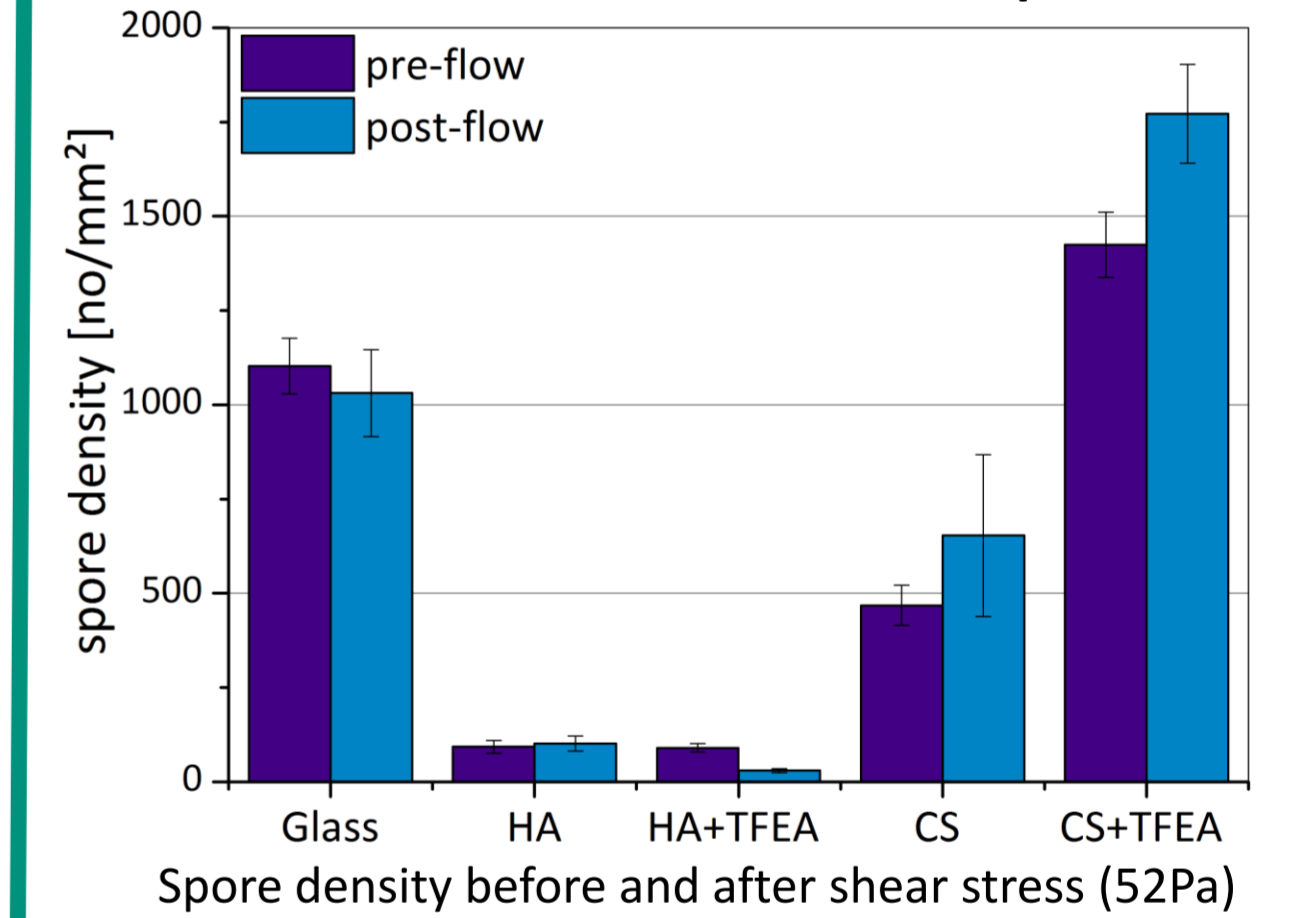
### Attachment of *Navicula perminuta*



A - Comparison of diatom density before and after shear stress (32Pa)  
B - Percentage removal

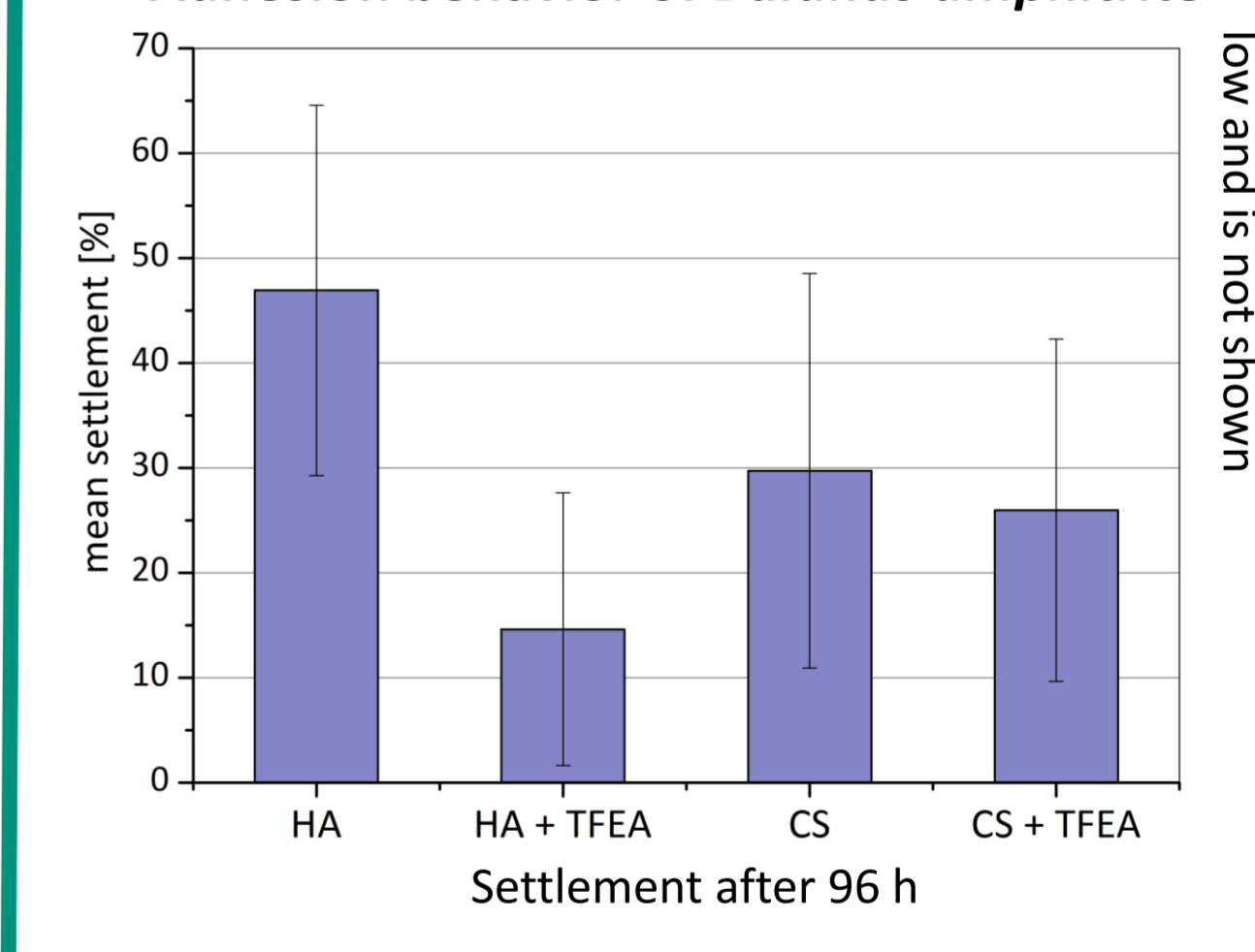
- TFEA capping weakens attachment for HA
- Attachment on CS surfaces is in range of glass standard, modification shows no effect

### Settlement of *Ulva linza* zoospores



- Settlement is very low on both HA coatings
- Modification makes CS films more attractive and adhesion strength increases

### Adhesion behavior of *Balanus amphitrite*



- Settlement is strongly reduced on modified HA
- Settlement comparable on both CS surfaces

## Literature

- [1] Morra, M.; Cassinelli, C. J. *Biomater. Sci.-Polym. Ed.* **1999**, *10*, 1107.
- [2] Cao, X. Y.; Pettit, M. E.; Conlan, S. L.; Wagner, W.; Ho, A. D.; Clare, A. S.; Callow, J. A.; Callow, M. E.; Grunze, M.; Rosenhahn, A. *Biomacromolecules* **2009**, *10*, 907.
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- [4] Krishnan, S.; Wang, N.; Ober, C. K.; Finlay, J. A.; Callow, M. E.; Callow, J. A.; Hexemer, A.; Sohn, K. E.; Kramer, E. J.; Fischer, D. A. *Biomacromolecules* **2006**, *7*, 1449.
- [5] Prime, K. L.; Whitesides, G. M. *Science* **1991**, *252*, 1164.

## Conclusions

- Amide modification does not worsen good protein resistance
- Modification with TFEA has a positive impact on HA coatings: decreased adhesion strength of *C.marina*, lower settlement and decreased adhesion strength of *N.perminuta*, *U.linza* and *B.amphitrite*
- Opposite behavior or no change in settlement of the different species is observed for CS coatings (likely related to the negative charge carrying sulfate-group)

## Acknowledgements

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