

Introduction

Peptaibols

- Natural membrane-active peptides isolated from fungi
- Abundant in Aib (U) (α -aminoisobutyric acid), possess a C-terminal alcohol and N-terminal acetylation/alkylation
- Display wide range of antimicrobial activities
- Able to lyse lipid membranes by pore formation

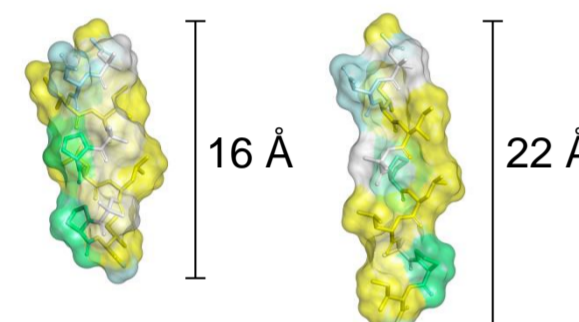
Harzianin HK-VI (HZ wt) is an ultra-short peptaibol (11-mer) isolated from *T. pseudokoningii* [1] with the sequence:



Research aim

To solve the structure of membrane-bound HZ and get insights into its interactions with lipid bilayers. How can the short HZ span the bilayer with the expected α -helical structure?

HZ as α -helix and 3_{10} -helix

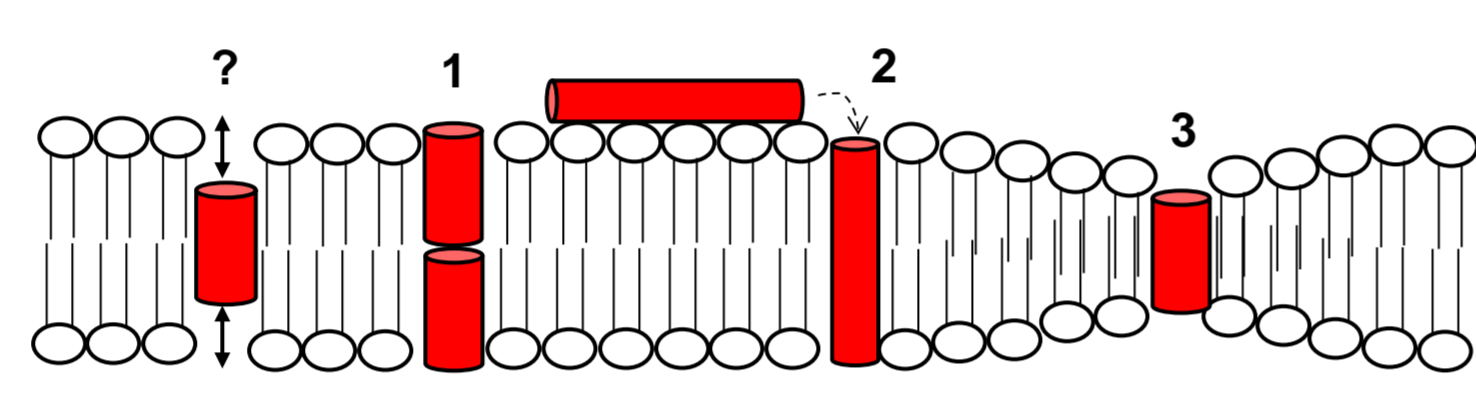


Inconsistency between peptide length and bilayer thickness.

D_b : bilayer thickness; $2D_c$: thickness of hydrocarbon region

¹T=30°C [2]

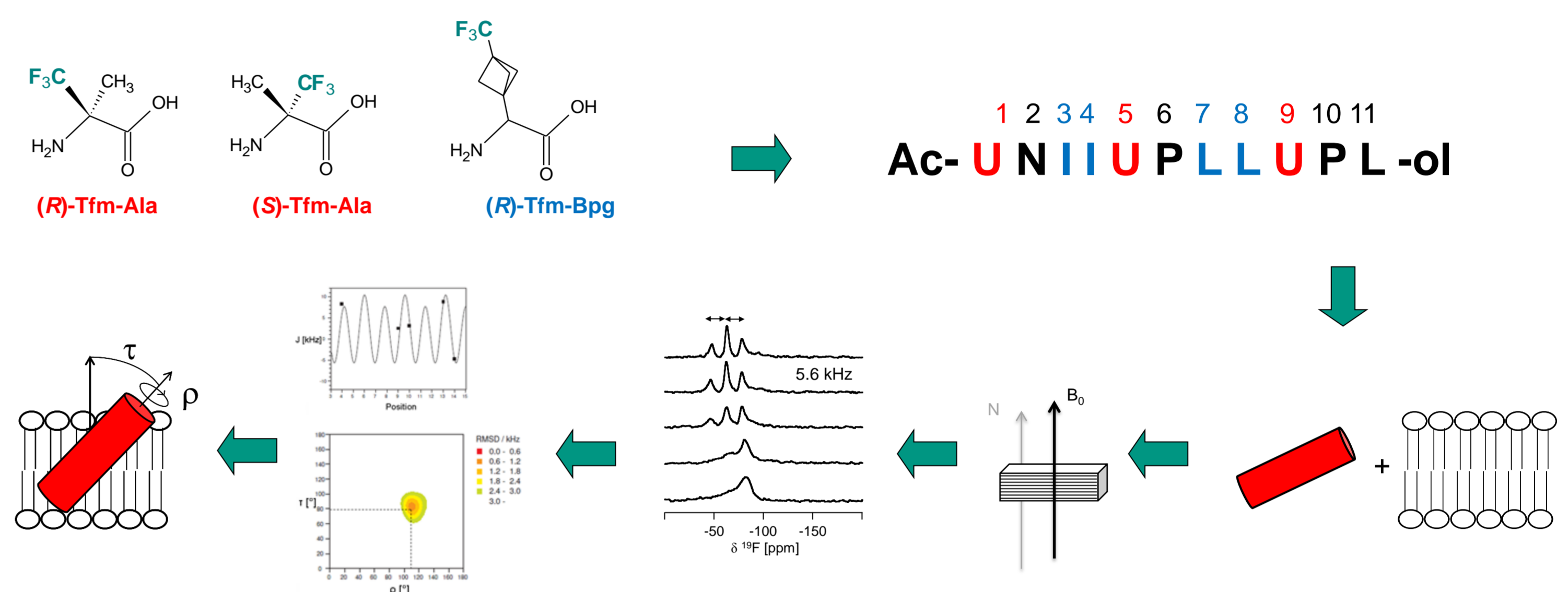
Lipid	D_b [Å]	$2D_c$ [Å]
DLPC	32.6	21.7
DMPC	36.7	25.7
POPC	39.1	28.8
DPhPC	35.4	27.2



Suggested mechanisms of peptide/bilayer interactions for short pore-forming peptides: arranging as double-layered channels (1); conformational change to extended conformations; (2) membrane thinning (3).

Methods

- Solid-state ¹⁹F-NMR and synchrotron circular dichroism (SRCD) spectroscopy to determine the structure and alignment of HZ wt in lipid bilayers
- ¹⁹F-NMR is enabled by one-at-a-time incorporation of synthetic α -trifluoromethylated amino acids: (*R*)-, (*S*)-Tfm-Ala [3,4] and (*R*)-Tfm-Bpg [5]
- Synthetic peptides are reconstituted in mechanically aligned (oriented) lipid bilayers
- From the ¹⁹F-NMR dipolar couplings, the **structure, orientation** and **dynamics** is determined [6]



- Oriented SRCD from non-labelled peptide (HZ wt) with the synchrotron UV/VIS light source ANKA complements the results for the overall alignment of peptides

Results

Antimicrobial tests

Bacteria	MIC [μ g/ml]	
	HZ wt	HZ Tfm analogues
<i>E. coli</i> K12	> 256	> 256
<i>S. aureus</i> DSM 1104	128	> 256
<i>S. xylophilus</i> DSM 20267	> 256	> 256
<i>E. faecalis</i> DSM 2570	> 256	> 256
<i>B. subtilis</i> ATCC 6633	> 256	> 256
Fungi		
<i>A. nidulans</i> (GR5)	> 256	> 256
<i>C. tropicalis</i>	> 256	> 256
<i>M. oryzae</i> (Guy 11)	32	32
<i>T. harzianum</i>	> 256	> 256

Antimicrobial activity (MIC) of HZ wt and ¹⁹F-labeled analogues.

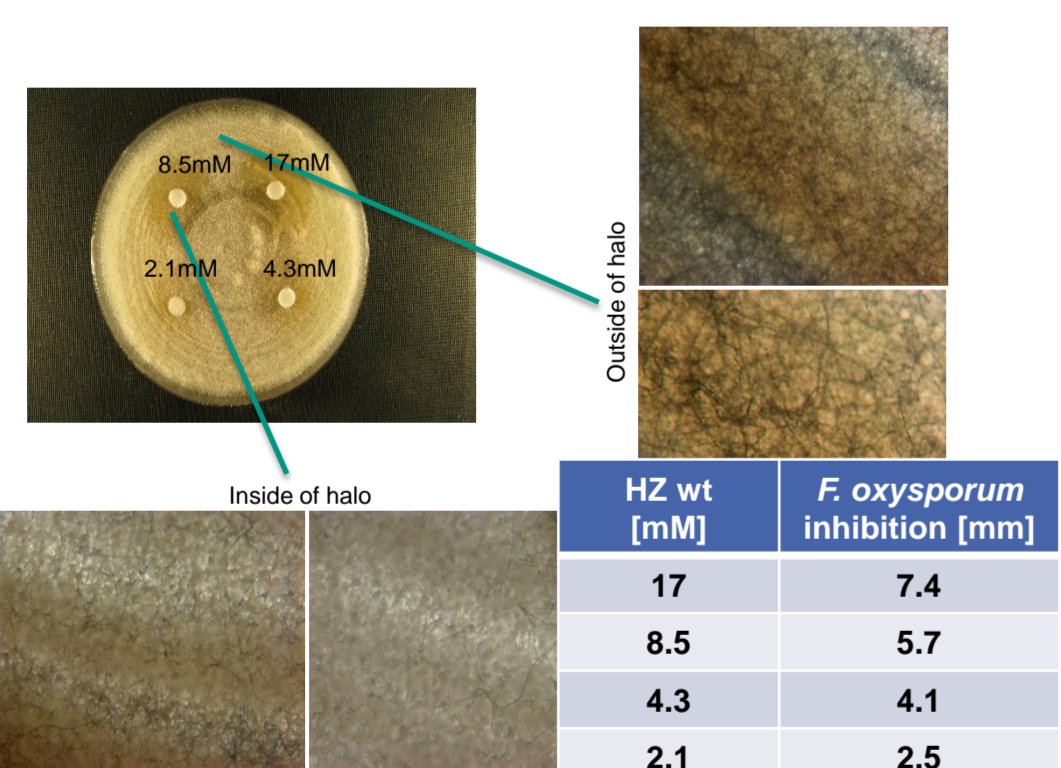
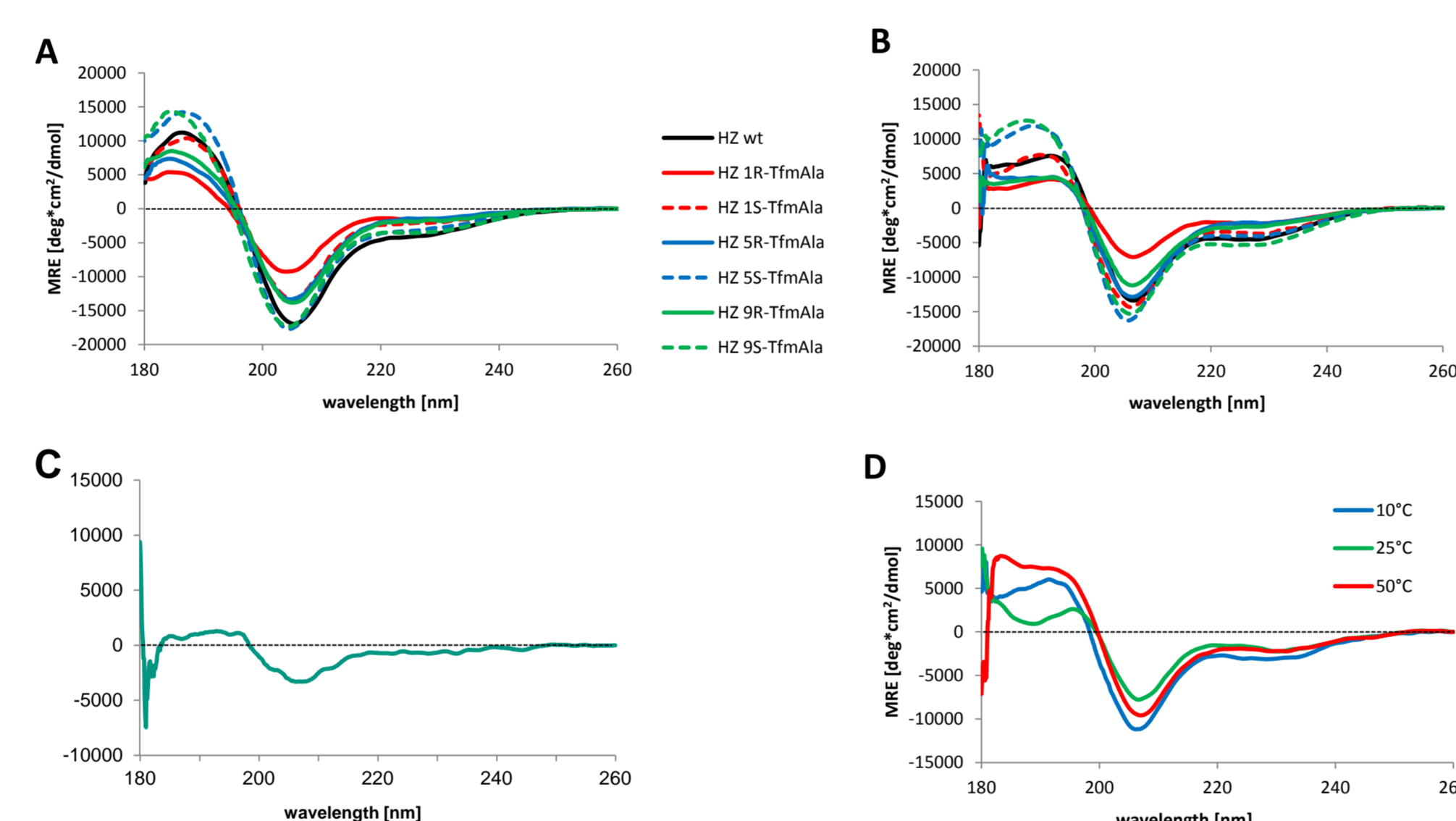


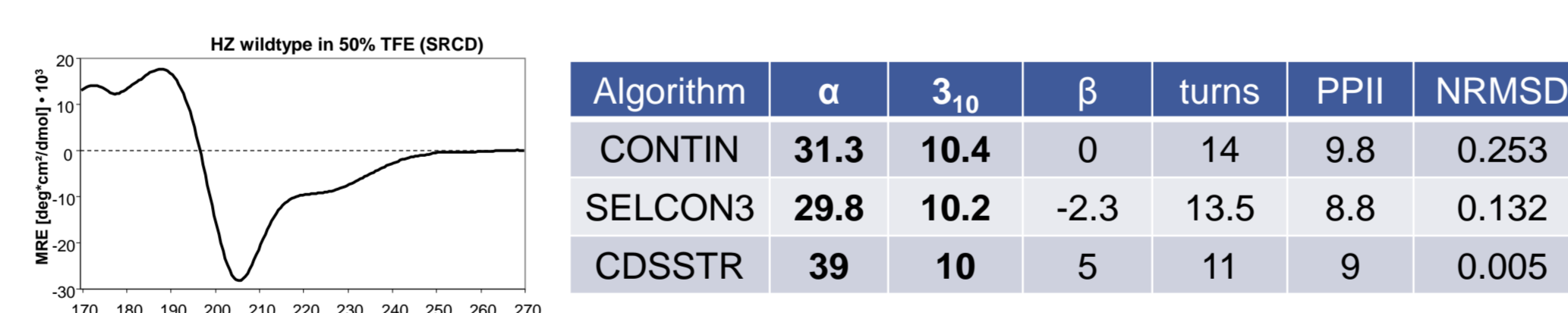
Plate diffusion assay testing HZ wt against *F. oxysporum*.

No pronounced antibacterial activity
Low antifungal effect against selected plant pathogenic fungi

Structure determination by CD



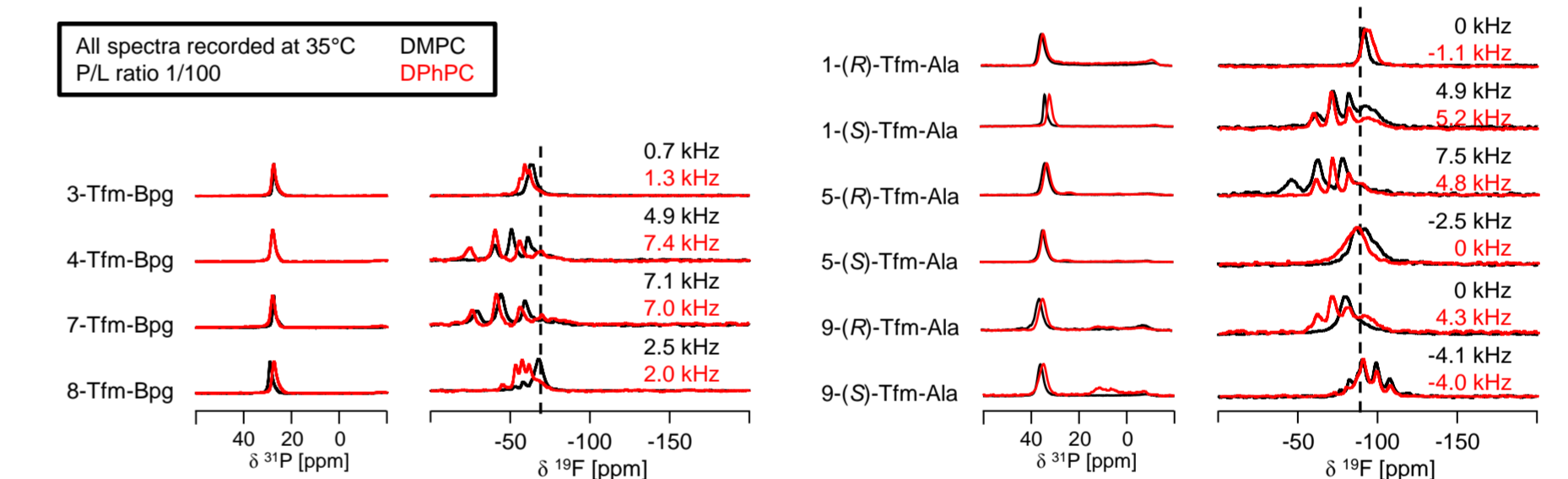
CD spectra of HZ peptides in organic solvent (TFE) (A); in detergent micelles (DPC, P/D 1/200) (B); in phospholipid liposomes (DPhPC (C) and DMPC (D), P/L 1/20).



Structural analysis of the SRCD spectrum of HZ wt in 50% TFE.

3_{10} -helical structure in various membrane models, whereas deconvolution suggests a predominant α -helical conformation

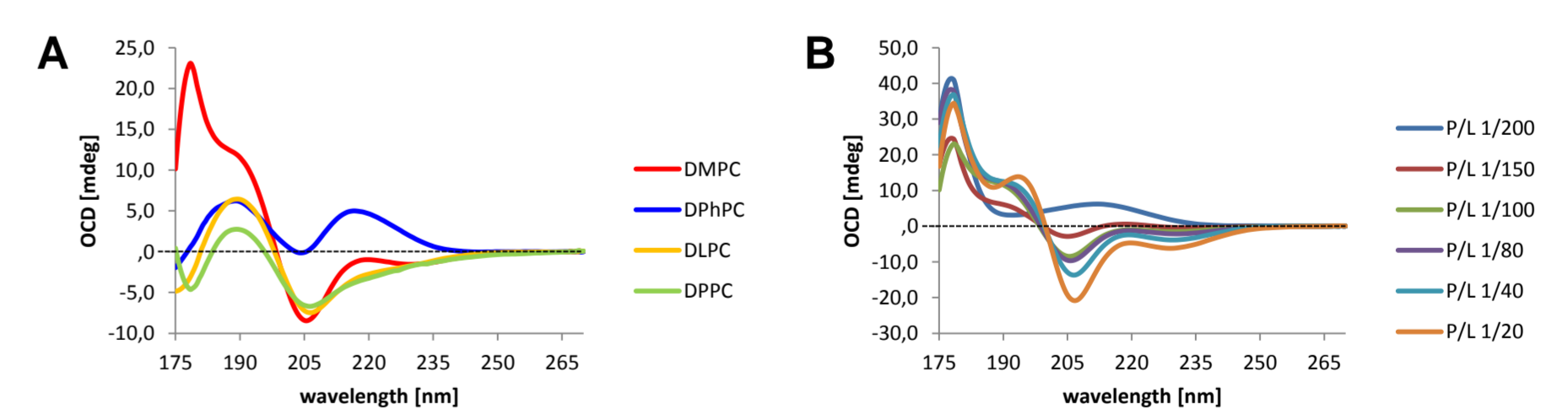
Peptide orientation by ssNMR and OCD



Solid-state ³¹P- and ¹⁹F-NMR spectra and observed ¹⁹F-NMR dipolar splittings of ¹⁹F-labeled HZ analogues in oriented DMPC and DPhPC bilayers; dotted line: isotropic position.

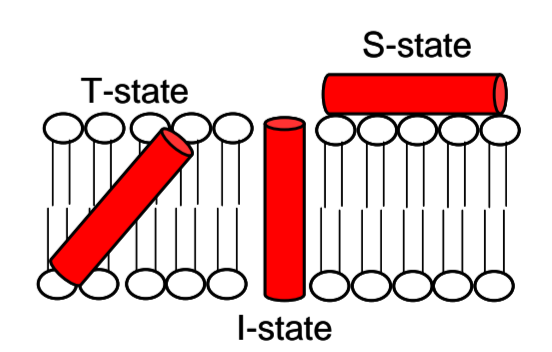
Structure model	Lipid	τ [°]	ρ [°]	S_{mol}	RMSD
3_{10} -helix (ideal)	DMPC	126	172	0.4	2.05
	DPhPC	78	10	0.5	2.0
α -helix (ideal)	DMPC	92	116	0.5	2.23
	DPhPC	96	102	0.5	0.59
β -bend ribbon spiral [7]	DMPC	102	118	0.6	1.38
	DPhPC	124	106	0.4	0.87

Putative alignment (τ and ρ angles) and dynamics (S_{mol}) of HZ in DMPC and DPhPC lipid bilayers assuming different models for secondary structure.



SR-OCD spectra of HZ wt in oriented phosphatidylcholine membranes of different composition (P/L 1/100) (A); HZ wt in oriented DMPC bilayers at varying PL (B).

DMPC: β -bend ribbon and S-state
DPhPC: α -helix and S-state
OCD suggests two different states



Outlook

- Structure determination of HZ wt by NMR in solution
- Synthesis of ¹⁵N-labeled HZ peptides to get more information on peptide alignment in lipid bilayers
- Analysis of membrane thinning (²H-NMR, MD simulations)
- Channel conductance measurements

Acknowledgements

This work has been carried out with the financial aid of the DFG/ANR program. Special thanks to S. Roth, J. Bürck for their support with SR-CD/OCD measurements at the UV-CD12 synchrotron beamline at ANKA.

References:

- [1] S. Rebuffat, S. Hilmi, Y. Prigent, C. Goulard, B. Bodo, J. Chem. Soc., Perkin Trans. 1 (1996) 2021; [2] N. Kučerka, M. Nieh, J. Katsaras, BBA Biomembr 1808 (2011) 2761-2771; [3] F. Huguénot, T. Brigaud, J. Org. Chem. 71 (2006) 7075-7078; [4] D. Maisch, P. Wadhvani, S. Afonin, C. Böttcher, B. Koksich, A.S. Ulrich, J. Am. Chem. Soc. 131 (2009) 15596-15597; [5] P.K. Mikhailuk, S. Afonin, A.N. Chernega, E.B. Rusanov, M. Platonov, G. Dubinina, M. Berditsch, A.S. Ulrich, I.V. Komarov, Angewandte Chem. 118 (2006), 5787-5789; [6] A.S. Ulrich, Progress in Nuclear Magnetic Resonance Spectroscopy 46 (2005) 1-21; [7] I. Segalas, Y. Prigent, D. Davoust, B. Bodo, S. Rebuffat, Biopolymers (1999) 71-85.